



Soviet-era science, translated into English

Reports of the Academy of Sciences of the USSR

PHYSICAL CHEMISTRY

1964

SovietRxiv

View the original and related papers at <https://sovietrxiv.org/items/ru-196401.15010>

Source: Math-Net.Ru and CyberLeninka. Machine translation. Verify with the original.

Abstract**Full Text**

Reports of the Academy of Sciences of the USSR
1964. Volume 155, No. 6

PHYSICAL CHEMISTRY

A. T. Vartanyan

**SPECTRAL STUDY OF THE INTERACTION
OF SOLID LAYERS OF PORPHIN DYES
WITH HYDRAZINE VAPORS**

(Presented by Academician A. N. Terenin on 22 X 1963)

Hemin and hematin are substances that form complex compounds with many solvents, with characteristic absorption spectra. They have an affinity for nitrogen-containing bases. If a little hydrazine hydrate is added to a solution of hemin in 0.1 *N* NaOH, then, owing to the formation of the corresponding hemochrome, the solution becomes red ^(1,2). Under the action of vapors of anhydrous hydrazine (N₂H₄) on solid layers of dyes of certain classes, unstable molecular compounds are formed ⁽³⁻⁵⁾. Depending on the part of the dye molecule in which the interaction is localized, either a colored or a colorless compound ("quasi-leuco base") is formed. It is also known that solutions of chlorophyll in a number of organic bases—piperidine ⁽⁶⁾, benzylamine ⁽⁷⁾, pyridine, nicotine, phenylhydrazine ⁽⁸⁾—have an absorption maximum at 642 *mμ*. It was of interest to study the direct interaction of solid layers of a series of porphin dyes with N₂H₄ vapors.

Experiments showed that, upon admission of N₂H₄ vapors, the spectrum of protoporphin does not change. From this important fact it follows that N₂H₄ molecules are localized neither at the vinyl groups of the pigment molecule nor at the nitrogen atoms of the pyrrole rings. Interaction with the carboxyl groups cannot affect the spectrum.

In contrast to the spectrum of protoporphyrin, the spectrum of the hematoporphyrin layer is very sensitive to N₂H₄ vapors. Curves 1 and 2 in Fig. 1 refer to a hematoporphyrin layer in vacuum and in N₂H₄ vapors, respectively. Under vacuum conditions the absorption maxima are at 502, 530, 560, and 600 *mμ*. After admitting the vapors, the new maxima are at 505, 536, 574, and 625 *mμ*, and their height increases regularly toward shorter wavelengths. The band at 405 *mμ* remains almost unchanged (curves 3 and 4). The same spectrum was observed for hematoporphyrin in 0.1 *N* KOH solution ⁽¹⁰⁾. It is noteworthy, however, that curve 2 is similar to curve 5, which belongs to a layer of protoporphyrin, except that it is shifted somewhat relative to curve 5 toward shorter

Fig. 2. Absorption spectra. 1 —pheophytin *a* in vacuum; 2 —the same in N_2H_4 vapors; 3 —chlorophyll *a* in vacuum; 4 —the same in N_2H_4 vapors; 5 —methyl chlorophyllide *a* in vacuum; 6 —the same in N_2H_4 vapors

Figure 2: Fig. 2. Absorption spectra. 1 —pheophytin *a* in vacuum; 2 —the same in N_2H_4 vapors; 3 —chlorophyll *a* in vacuum; 4 —the same in N_2H_4 vapors; 5 —methyl chlorophyllide *a* in vacuum; 6 —the same in N_2H_4 vapors

that the N_2H_4 molecules, in their interaction with hemin, are localized at the iron atom. This is also confirmed by magnetic measurements¹². In addition, as our measurements of absorption spectra in the visible and ultraviolet regions have shown, N_2H_4 vapors do not act on a layer of metal-free phthalocyanine, but do act, although weakly, on layers of magnesium and iron phthalocyanines. This agrees with the results of a study of the action of N_2H_4 vapors on infrared-spectra of phthalocyanine layers⁽¹³⁾. Thus, also in the direct interaction of N_2H_4 vapors with a hemin layer, the spectrum is transformed into the spectrum of the corresponding hemochrome, in which the band maxima at 410, 525, and 555 $m\mu$ prove to be shifted relative to the hemin maxima by 10 $m\mu$ toward longer wavelengths. As for the band at 325 $m\mu$, it is apparently the result of the splitting of the band at 400 $m\mu$ into two independent bands as a consequence of the above-noted narrowing of the bands upon formation of the hemochrome (Fig. 1, 7-9).

Fig. 2. Absorption spectra. **1** —pheophytin *a* in vacuum; **2** —the same in N_2H_4 vapors; **3** —chlorophyll *a* in vacuum; **4** —the same in N_2H_4 vapors; **5** —methyl chlorophyllide *a* in vacuum; **6** —the same in N_2H_4 vapors.

The results of measuring the spectra of hematin layers in vacuum (Fig. 1, 8) and in N_2H_4 vapors (Fig. 1, 9)* lead to the same conclusions.

N_2H_4 vapors also interact with layers of green-leaf pigments related to blood pigments. Upon interaction of layers of pheophytin *a*, chlorophyll *a*, and methyl chlorophyllide *a* (Fig. 2) with N_2H_4 vapors, the main red maxima shift toward shorter wavelengths, while absorption in the region 405-420 $m\mu$ increases noticeably. Pheophytin is characterized by disappearance of the band at 320 $m\mu$ and enhancement of absorption at 504 $m\mu$ (Fig. 2, 1 and 2). If the spectrum of protoporphyrin is compared with the spectra of hematoporphyrin and pheophytin in N_2H_4 vapors, then in the region shorter than 550 $m\mu$ a similarity is observed between the spectra. For such a similarity to be observed, localization of the N_2H_4 molecule at the oxygen atom in the five-membered

* Through an unfortunate misunderstanding, in papers^(14,15) for hematin we gave results relating to another compound.

ring of the pheophytin molecule, leading to the reaction:

structural fragment of pheophytin with $\text{HC}=\text{C}=\text{O} + \text{N}_2\text{H}_4 \rightarrow$ corresponding fragment with $\text{C}=\text{O}^-$ and

Since a shift of the main red band is also observed for pheophytin*, it is obvious that it cannot be due to the interaction of the N_2H_4 molecule with the central magnesium atom in the chlorophyll molecule. The appearance in the spectrum of a chlorophyll layer exposed to N_2H_4 vapors of a band with a maximum at 640 m (Fig. 2, 4) is apparently connected with the above reaction. The band at 640 m in the spectrum of chlorophyll *a* solutions in organic bases may have the same origin. According to Krasnovskii and Brin⁽⁸⁾, the shift may be associated with ionization of the "acidic" groups of the pigment molecule.

The difference between the frequencies of the maxima of chlorophyll *a* (624 and 672 m) and methyl chlorophyllide *a* (627 and 676 m), equal to 1150 cm^{-1} , decreases after admission of N_2H_4 vapors to 1050 cm^{-1} . This value is close to the difference between the frequencies of the absorption maxima of hemin and hematin in N_2H_4 vapors (525 and 555 m), which is equal to 1030 cm^{-1} . The possibility is not excluded that chlorophyll may interact weakly with the N_2H_4 molecule also by the hemochrome type.

Bilirubin—the coloring substance of bile—is also in close genetic relationship to hemin. The length of the chain of conjugated bonds in its molecule is approximately half that in the molecules considered above. The spectrum of a bilirubin layer consists of a main band (apparently double) in the region of 460 m and weak bands with maxima at 240, 285, and 320 m. In the presence of N_2H_4 vapors, the maximum of the main band shifts to 440 m, its height decreases by approximately 25%, and the half-width of the band increases by 20%. As was to be expected, N_2H_4 vapors do not exert a very strong effect on the general form of the spectrum, since N_2H_4 molecules can be localized at OH groups at the site of chain rupture. Interaction with OH groups was previously shown by us for aurin, fluorescein, and gallein⁽⁵⁾.

Thus, analysis of the absorption spectra of layers of the pigments studied, kept in vacuum and in hydrazine vapors, makes it possible in each individual case to indicate the site of direct interaction of pigment and hydrazine molecules and to establish its nature.

Received
17 X 1963

CITED LITERATURE

1. L. Heilmeyer, *Medizinische Spektrophotometrie*, Jena, 1933, p. 116.
2. R. Lemberg, J. W. Legge, *Hematin Compounds and Bile Pigments*, N. Y., 1949, p. 174.

3. A. T. Vartanyan, ZhFKh, **35**, 2241 (1961).
4. A. T. Vartanyan, ZhFKh, **36**, 1890 (1962).
5. A. T. Vartanyan, ZhFKh, **36**, 2118 (1962).
6. E. Katz, E. C. Wassink, *Enzymol.*, **7**, 97 (1939).
7. R. Livingstone, W. F. Watson, J. McArdle, *J. Am. Chem. Soc.*, **71**, 1542 (1949).
8. A. A. Krasnovskii, G. P. Brin, DAN, **89**, 527 (1953).
9. A. T. Vartanyan, *Izv. AN SSSR, ser. fiz.*, **16**, 169 (1952).
10. L. Heilmeyer, *Medizinische Spektralphotometrie*, Jena, 1933, p. 144.
11. F. Gund, *J. Soc. Dyers Colour.*, **69**, 671 (1953).
12. L. Pauling, C. D. Coryell, *Proc. Nat. Acad. Sci., U.S.A.*, **22**, 159 (1936).
13. A. N. Sidorov, A. N. Terenin, DAN, **104**, 575 (1955).
14. A. T. Vartanyan, *Izv. AN SSSR, ser. fiz.*, **27**, 37 (1963).
15. A. T. Vartanyan, DAN, **143**, 1317 (1962).

* In work (⁸), no such shift was found.

Note: Figure translations are in progress. See original paper for figures.

Source: Math-Net.Ru and CyberLeninka. Machine translation. Verify with the original.