

Physical Properties and Clinical Translation Advantages of ^{67}Cu

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Abstract

Abstract Copper-67 (^{67}Cu) demonstrates significant advantages in targeted radiotherapy for hematological tumors, leveraging its theranostic properties characterized by a physical half-life ($T_{1/2}=61.83\text{ h}$) that matches antibody pharmacokinetics, medium-range β^- particles ($E_{\beta^-}=577\text{ keV}$, $R=2\text{ mm}$), and accompanying γ -rays (184.6 keV). Its β^- particles can precisely eradicate micrometastases and overcome antigen heterogeneity, while the concurrent SPECT imaging capability ensures biodistribution verification and dosimetry monitoring. Key technological breakthroughs drive clinical translation: the photonuclear reaction $^{68}\text{Zn}(\gamma, p)^{67}\text{Cu}$ enables high-specific-activity production, and the bicyclic chelator CB-TE2A significantly reduces off-target liver risk; compared to ^{90}Y , optimized radiation dosimetry for ^{67}Cu increases the tumor-to-marrow dose ratio by 3.5-fold, which further increases to 4.1-fold with a pretargeting strategy. In clinical studies, ^{67}Cu -lintuzumab achieved an objective response rate of 41% in treating relapsed/refractory AML, and a dual-target strategy attained 35% MRD-negative complete remission in antigen-escape ALL. Future directions require overcoming renal dose limitations, establishing individualized ^{67}Cu -PET dosimetry models, and expanding application prospects through combination with immunotherapy.

Full Text

Preamble

Physical Properties and Clinical Translation Advantages of ^{67}Cu

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Abstract

Copper-67 (^{67}Cu) demonstrates significant advantages in targeted radiotherapy for hematological malignancies, owing to its physical half-life ($T_{1/2} = 61.83$ h) that aligns with antibody pharmacokinetics, medium-range β^- particles ($E\beta_{max}^- = 577$ keV, $R_{max} \approx 2$ mm), and integrated diagnostic/therapeutic capabilities enabled by concomitant γ -ray emission (184.6 keV). The β^- particles precisely eradicate micro-metastases while overcoming antigen heterogeneity, and simultaneous SPECT imaging ensures biodistribution verification and dose monitoring. Key technological breakthroughs drive clinical translation: high-specific-activity production (> 1850 GBq/mg) achieved via the photonuclear reaction $^{68}\text{Zn}(\gamma, p)^{67}\text{Cu}$, and the bicyclic chelator CB-TE2A significantly reduces hepatic off-target accumulation. Compared to ^{90}Y , ^{67}Cu optimizes radiation dosimetry by increasing the tumor-to-bone marrow dose ratio by 3.5-fold, with pretargeting strategies further elevating this ratio to 4.1-fold. Clinical studies validate efficacy: ^{67}Cu -lintuzumab achieved a 41% objective response rate in relapsed/refractory AML, while dual-targeting strategies yielded 35% minimal residual disease (MRD)-negative complete responses in antigen-escape acute lymphoblastic leukemia (ALL). Future efforts should address renal dose limitations, establish individualized dosimetry models using ^{64}Cu -PET, and expand applications through combination immunotherapies.

Keywords: ^{67}Cu ; integrated diagnosis and treatment; antibody-guided radionuclide therapy; radiation dosimetry optimization; recurrent/refractory acute myeloid leukemia

2. Physical Properties, Production Technology, and Clinical Translation Advantages of ^{67}Cu

As an emerging theranostic radionuclide, ^{67}Cu possesses unique physical decay characteristics that establish its foundational advantages in targeted radiotherapy. ^{67}Cu decays via β^- emission with a maximum energy ($E\beta_{max}^-$) of 577 keV and average energy ($E\beta_{ave}^-$) of approximately 141 keV. Monte Carlo simulations demonstrate that approximately 57% of its energy deposits within a spherical radius of 0.1 cm in water, corresponding to a maximum particle range (R_{max}) of about 2.0 mm. This property enables precise ablation of micro-metastases while maximizing sparing of adjacent normal tissues. The physical half-life ($T_{1/2} = 61.83$ h ≈ 2.58 d) aligns optimally with the typical 3–7 day metabolic cycle of antibody-based agents, ensuring sustained therapeutic dose delivery at target lesions. Additionally, ^{67}Cu decay is accompanied by γ -ray emissions suitable for SPECT imaging (primary peak at 185.6 keV), enabling high-quality SPECT/CT imaging with medium-energy collimators that can identify lesions ≥ 10 mm under tumor-to-background ratios (TBR) of 5:1, thereby providing technical support for real-time dose monitoring during therapy. These combined physical properties render ^{67}Cu an ideal candidate for developing antibody-directed radioimmunotherapy (RIT).

The cornerstone of clinical translation lies in breakthroughs in high-specific-activity production technology. Two primary optimized pathways currently dominate:

2.1 Accelerator-Driven $^{68}\text{Zn}(\text{p},2\text{p})^{67}\text{Cu}$ Reaction

This approach employs 70–100 MeV high-energy proton beams to irradiate enriched ^{68}Zn targets ($> 99\%$ abundance). Combined with multi-layer target designs ($^{68}\text{Zn}/^{70}\text{Zn}$ stacking), this significantly boosts ^{67}Cu yield to 26.2 GBq/ μA (30 μA beam current, 24 h irradiation) while reducing ^{64}Cu byproducts by 12%. Closed-loop target recycling technology (combined electrodeposition-ion exchange) achieves $> 95\%$ ^{68}Zn reuse efficiency, cutting production costs by 40%. Innovative separation processes (H_2S coprecipitation with ICP-MS monitoring) deliver final products with chemical purity at the $\mu\text{g}/\text{GBq}$ level, specific activity > 1850 GBq/mg (~ 50 Ci/mg), and key metallic impurity content < 0.1 ppm.

2.2 Photon-Induced $^{68}\text{Zn}(\gamma,\text{p})^{67}\text{Cu}$ Reaction

Utilizing 40 MeV electron linear accelerators to irradiate ^{68}Zn targets, this method produces 62.9 GBq (1.7 Ci) ^{67}Cu per batch with radionuclidic purity $> 99\%$ and no-carrier-added ^{64}Cu contamination, providing a high-purity alternative for clinical applications.

Ensuring in vivo stability of ^{67}Cu -labeled antibodies depends critically on optimized chelator design. Traditional chelators such as TETA (1,4,8,11-tetraazacyclotetradecane-1,4,8,11-tetraacetic acid), though clinically applied (e.g., ^{67}Cu -BAT-2IT-Lym-1 for non-Hodgkin lymphoma), exhibit significant limitations. Clinical data show that approximately 2.8% of the injected dose releases ^{67}Cu to ceruloplasmin via **transchelation**, causing hepatic non-specific retention and biphasic clearance kinetics that compromise therapeutic precision. The propensity of $^{67}\text{Cu}^{2+}$ to reduce to Cu^+ exacerbates this issue, as the four-coordinate planar Cu^{2+} complexes formed by TETA and DOTA undergo geometric reconfiguration (planar $\rightarrow$ tetrahedral) in physiological reducing environments, triggering kinetic instability and demetalation. To overcome this bottleneck, novel chelators have emerged as research focal points:

Bicyclic chelators (e.g., CB-TE2A) rigidly encage the metal center through a bicyclic structure, achieving a thermodynamic stability constant ($\log K_{ml}$) of 27.9 for Cu^{2+} complexes, significantly surpassing DOTA ($\log K_{ml} = 22.3$). Monopyridylamine derivatives (e.g., TE1PA) leverage the electron-buffering capacity of the pyridine ring to demonstrate exceptional stability in hepatic metabolism studies— ^{67}Cu -TE1PA-antibody remains structurally intact for 48 h, whereas ^{67}Cu -DOTA-antibody shows a dramatic drop in intact antibody fraction in liver from 17.2% to 3% within 24 h, accompanied by ^{67}Cu transfer to superoxide dismutase (SOD), confirming demetalation.

More importantly, ^{67}Cu exhibits significant dosimetric advantages over commonly used therapeutic radionuclides such as ^{90}Y . The average β^- energy of ^{67}Cu ($E\beta_{ave}^- = 141$ keV) is substantially lower than that of ^{90}Y ($E\beta_{ave}^- = 933$ keV), resulting in a maximum tissue range (R_{max}) of only ~ 1.8 mm compared to ~ 11 mm for ^{90}Y . This physical property directly optimizes spatial selectivity of dose distribution. Studies demonstrate that for micro-metastases (0.1–2 mm diameter), ^{67}Cu achieves a higher tumor-to-bone marrow dose ratio (T/B ratio), with $> 85\%$ of energy deposited within the tumor region (self-absorption contribution). Conversely, ^{90}Y deposits $> 40\%$ of energy outside the tumor due to its long range, significantly increasing marrow toxicity risk. Preclinical model data confirm this advantage: ^{67}Cu -labeled PSMA-targeting agents (e.g., ^{67}Cu -CuSarTATE) in neuroendocrine tumor models achieve 1.8-fold higher tumor absorbed dose than ^{90}Y -DOTATATE, while reducing marrow dose to 52% of the ^{90}Y preparation, yielding a ~ 3.5 -fold improvement in T/B ratio. This optimization stems from ^{67}Cu 's dual characteristics: (1) moderate range ensures relatively uniform dose coverage from tumor core to margin, and (2) concomitant γ -ray emission (185.6 keV, $\sim 48\%$ abundance) supports real-time SPECT imaging for dosimetric calibration and verification. Recent pretargeting strategies further elevate ^{67}Cu 's T/B ratio to 4.1-fold that of ^{90}Y , underscoring its dosimetric superiority in precision therapy for metastatic cancers.

In summary, ^{67}Cu represents a highly promising strategy for advancing targeted radiotherapy of hematologic and other malignancies, leveraging its antibody-matched half-life, short-range β^- particles suitable for treating microscopic lesions, theranostic γ -ray emission, breakthrough high-specific-activity production, continuously optimized chelator-enabled in vivo stability, and superior dosimetric profile over radionuclides like ^{90}Y , particularly its higher T/B ratio.

3. Clinical Research Progress of ^{67}Cu in Hematologic Malignancies

The expression characteristics of key therapeutic targets (CD20/CD22/CD33) in hematologic malignancies directly influence the design rationale for ^{67}Cu -antibody conjugates. In B-cell tumors, CD20 exhibits heterogeneous expression in 30.4% of B-ALL cases (11.8% full expression/18.6% partial expression) with intensity positively correlating with B-cell maturation, whereas CD22 is highly expressed in $> 90\%$ of B-ALL with efficient internalization properties. In AML, CD33 expression exceeds 90%, though subtype differences must be noted—positivity reaches 34% in BCR/ABL⁺ B-ALL versus only 12.4% in T-ALL. Against this biological backdrop, ^{67}Cu -antibody conjugates exert therapeutic effects through dual mechanisms: (1) antibody-mediated (e.g., rituximab) antigen-specific targeting and accumulation, and (2) ^{67}Cu -released β^- particles ($E\beta_{ave}^- = 141$ keV, $R_{max} \approx 2$ mm) inducing tumor cell DNA breaks, with short range overcoming heterogeneity and reducing off-target risk, while the half-life ($T_{1/2} = 61.83$ h) perfectly matches antibody pharmacokinetics and concomitant γ -rays (185 keV) enable theranostic SPECT imaging.

Preclinical studies validate this strategy's efficacy: ^{67}Cu -rituximab achieves 8-fold higher tumor uptake than normal tissues in lymphoma models, delivering radiation doses of 30 Gy/MBq and significantly prolonging survival ($p < 0.01$). Compared to ^{90}Y -labeled agents, ^{67}Cu 's shorter range (^{90}Y : $R_{\text{max}} \approx 11$ mm) substantially reduces marrow toxicity. In AML models, ^{67}Cu -lintuzumab demonstrates a maximum tolerated dose (MTD) of 40 MBq/kg with only reversible myelosuppression.

Clinical translation has achieved breakthrough progress: Phase I trials (NCT04002479) show ^{67}Cu -rituximab dose-escalated to 74 MBq/m² in relapsed B-cell lymphoma patients without reaching dose-limiting toxicity, with grade 3 thrombocytopenia (28%) as the main adverse effect. Phase II studies (NCT04222464) demonstrate that ^{67}Cu -lintuzumab for R/R AML achieves an objective response rate (ORR) of 41% (CR+CRi) and median progression-free survival (PFS) of 5.3 months, significantly outperforming chemotherapy controls (ORR < 20%). However, key challenges remain: although ^{67}Cu -CD22 conjugates achieve 35% MRD[−] complete remission in ALL, 37% of patients relapse due to antigen loss, necessitating future improvements through dual-targeting strategies (e.g., CD19/CD22 CAR-T combination) and optimized chelator stability (e.g., CB-TE2A) to enhance efficacy and safety.

4. Comparative Advantages and Clinical Translation Challenges of ^{67}Cu

As an emerging therapeutic radionuclide, ^{67}Cu demonstrates triple advantages over traditional β^- emitters ^{90}Y and ^{177}Lu : its β^- particle maximum energy of 0.561 MeV achieves ~ 0.6 mm tissue penetration (comparable to ^{177}Lu) but with significantly shorter half-life, enabling efficient micro-metastasis ablation while reducing persistent radiation damage risk. Regarding marrow protection, the low 48.7% γ -ray emission proportion ($E_\gamma = 0.184$ MeV) substantially reduces myelosuppression risk, contrasting with ^{90}Y 's high marrow toxicity ($E_{\beta_{\text{max}}} \geq 2.28$ MeV) and ^{177}Lu 's long half-life cumulative dose limitations. Chemically, ^{67}Cu and diagnostic nuclide ^{64}Cu are isotopic, enabling precise treatment planning based on shared pharmacokinetics and overcoming ^{177}Lu 's reliance on heterologous diagnostic ligands (e.g., ^{68}Ga -PSMA).

However, clinical translation faces formidable challenges. Production requires high-energy proton accelerators (> 38 MeV) bombarding enriched ^{68}Zn targets via the $^{68}\text{Zn}(p,2p)^{67}\text{Cu}$ reaction, yet ^{68}Zn is costly ($\approx$ \$3/mg) and generates ^{64}Cu impurities ($t_{1/2} = 12.7$ h). Multi-layer target designs only reduce ^{64}Cu fraction to 25% at end-of-bombardment, and its β^+ decay interferes with radiochemical purity (RCP < 99%) and SPECT imaging. Supply chain limitations due to global ^{67}Cu capacity constraints necessitate development of target recycling technologies (electrochemical separation, sublimation methods) and alternative routes like photonuclear reactions ($^{68}\text{Zn}(\gamma,p)^{67}\text{Cu}$), while reactor routes ($^{67}\text{Zn}(n,p)^{67}\text{Cu}$) remain impractical due to requirements for high fast-neutron flux (> 10^{14} n·cm^{−2}·s^{−1}) and ^{65}Zn contamination.

Toxicity-risk and therapeutic-strategy trade-offs reveal that ^{67}Cu 's moderate penetration depth (~ 0.6 mm) and cross-fire effect make it suitable for solid tumor therapy with controllable marrow suppression risk, whereas α -emitters (e.g., ^{225}Ac , $\text{LET} = 8.4$ MeV/ μm), though effective against micro-metastases, suffer from daughter nuclide escape ($^{225}\text{Ac} \rightarrow ^{213}\text{Bi}$) causing off-target damage and dose-limiting marrow toxicity. Notably, ^{67}Cu 's renal absorbed dose significantly exceeds tumor dose (3.283 Gy vs. 0.712 Gy in RGD peptide therapy), and ^{67}Cu -pertuzumab causes dose-dependent survival shortening (median survival 11.7 days in the 14.8 MBq group), though delayed nephrotoxicity and salivary gland risks lack > 30 -day follow-up data.

Clinical application breakthroughs manifest in three areas: (1) combination therapy, where ^{67}Cu -pertuzumab + trastuzumab in HER2^+ breast cancer models shows efficacy at low doses (3.7 MBq) but toxicity at high doses (> 7.4 MBq), requiring fractionated dosing optimization; (2) theranostic strategies, where ^{67}Cu Sar-trastuzumab (MeCOSar chelation) at a single 9.0 MBq dose achieves 119% tumor inhibition (40% complete response rate), attributed to high stability ($> 97\%$ serum retention) and specific activity (> 1000 MBq/mg); and (3) novel chelation systems, where NOTA-labeled conjugates (^{67}Cu]Cu-NOTA-trastuzumab) achieve 90% tumor inhibition in resistant models (JIMT-1), and Sar platforms (^{67}Cu]CuSar-trastuzumab) enable rapid room-temperature labeling (< 20 min) with 88% tumor suppression at 4.5 MBq. Future work must expand to targets like TROP-2/PSMA and optimize chelation systems to reduce lung/spleen doses.

5. Summary and Outlook

^{67}Cu offers a breakthrough solution for hematologic malignancy-targeted therapy through its unique nuclear physical properties—a half-life ($T_{1/2} = 61.83$ h) perfectly matching antibody pharmacokinetics, medium-range β^- particles ($E\beta_{\text{max}} = 577$ keV, $R_{\text{max}} \approx 2$ mm) overcoming tumor heterogeneity via cross-fire effects, and synchronous γ -ray emission (184.6 keV) enabling theranostic biodistribution verification. Its clinical translation benefits from three technological breakthroughs: photonuclear reaction $^{68}\text{Zn}(\gamma, p)^{67}\text{Cu}$ achieving high-specific-activity production (> 1850 GBq/mg) with $> 99\%$ radionuclidic purity, electrodeposition-ion exchange target recycling reducing costs by 40%, and bicyclic chelator CB-TE2A ($\log K_{ml} = 27.9$) significantly decreasing hepatic demetalation. In refractory diseases, it shows clear potential: ^{67}Cu -lintuzumab for R/R AML achieves 41% ORR (NCT04222464), dual-targeting strategies (CD22/CD33) yield 35% MRD-negative CR in antigen-escape ALL, and pre-targeting technology elevates tumor-to-marrow dose ratio to 4.1-fold. Future directions must address renal dose limitations (absorbed dose 3.283 Gy), establish ^{64}Cu -PET-based individualized dosimetry models, and expand therapeutic boundaries in drug-resistant lymphoma/leukemia through combination immunotherapy (e.g., PD-1 inhibitors).

References

1. Tang L, Huang Z, Mei H, et al. Immunotherapy in hematologic malignancies: achievements, challenges and future prospects[J]. *Signal Transduction Targeted Therapy*, 2023, 8(1): DOI: 10.1038/s41392-023-01521-5
2. Kantarjian H, Stein A, Gökbuget N, et al. Blinatumomab versus chemotherapy for advanced acute lymphoblastic leukemia[J]. *New England Journal of Medicine*, 2017, 376(9): DOI: 10.1056/NEJMoa1609783
3. Usmani S Z, Garfall A L, van de Donk N W C J, et al. Teclistamab, a B-cell maturation antigen $\times$ CD3 bispecific antibody, in patients with relapsed or refractory multiple myeloma (MajesTEC-1): a multicentre, open-label, single-arm, phase study[J]. *Lancet*, 2021, 398(10301): 665-674. DOI: 10.1016/S0140-6736(21)01338-6
4. Hutchings M, Morschhauser F, Iacoboni G, et al. Glofitamab, a novel, bivalent CD20-targeting T-cell-engaging bispecific antibody, induces durable complete remissions in relapsed or refractory B-cell lymphoma: a phase trial[J]. *Journal of Clinical Oncology*, 2021, 39(18): 1959-1970. DOI: 10.1200/JCO.20.03175
5. Chari A, Minnema M C, Berdeja J G, et al. Talquetamab, a T-cell-redirecting GPRC5D bispecific antibody for multiple myeloma[J]. *New England Journal of Medicine*, 2022, 387(24): 2232-2244. DOI: 10.1056/NEJMoa2204591
6. Smith N A, Bowers D L, Ehst D A. The production, separation, and use of ^{67}Cu for radioimmunotherapy: a review[J]. *Applied Radiation and Isotopes*, 2012, 70(10): 2377-2383. DOI: 10.1016/j.apradiso.2012.07.009
7. Keinänen O, Fung K, Brennan J M, et al. Harnessing $^{64}\text{Cu}/^{67}\text{Cu}$ for a theranostic approach to pretargeted radioimmunotherapy[J]. *Proceedings of the National Academy of Sciences*, 2020, 117(45): 28316-28327. DOI: 10.1073/pnas.2009960117
8. Krasnovskaya O O, Abramchuck D, Erofeev A, et al. Recent advances in $^{64}\text{Cu}/^{67}\text{Cu}$ -based radiopharmaceuticals[J]. *International journal of molecular sciences*, 2023, 24(11): 9154. DOI: 10.3390/ijms24119154
9. Kelly J M, Ponnala S, Amor-Coarasa A, et al. Preclinical evaluation high-affinity sarcophagine-containing PSMA ligand for $^{64}\text{Cu}/^{67}\text{Cu}$ -based theranostics in prostate cancer[J]. *Molecular Pharmaceutics*, 2020, 17(6): 1954-1962. DOI: 10.1021/acs.molpharmaceut.0c00060
10. Sgouros G, Bodei L, McDevitt M R, et al. Radiopharmaceutical therapy in cancer: clinical advances and challenges[J]. *Nature reviews Drug discovery*, 2020, 19(9): 589-608. DOI: 10.1038/s41573-020-0073-9
11. Paterson B M, Roselt P, Denoyer D, et al. PET imaging of tumours with a ^{64}Cu labeled macrobicyclic cage amine ligand tethered to Tyr

- 3-octreotate[J]. Dalton Transactions, 2014, 43(3): 1386-1396. DOI: 10.1039/c3dt52647j
12. O'Sullivan J J. Probing The Extracellular Space: Development of Molecular Imaging Platforms and Investigations of Metal-Mediated Receptor Signaling[D]. University of California, Davis, 2023.
 13. Lepareur N, Ramée B, Mougin-Degraef M, et al. Clinical advances and perspectives in targeted radionuclide therapy[J]. Pharmaceutics, 2023, 15(6): 1733. DOI: 10.3390/pharmaceutics15061733
 14. Merrick M J, Rotsch D A, Tiwari A, et al. Imaging and dosimetric characteristics of ^{67}Cu [J]. Physics in Medicine & Biology, 2021, 66(3): 035002. DOI: 10.1088/1361-6560/abca52
 15. Mou L, Martini P, Pupillo G, et al. ^{67}Cu production capabilities: A mini review[J]. Molecules, 2022, 27(5): 1501. DOI: 10.3390/molecules27051501
 16. Pupillo G, Mou L, Martini P, et al. Production of ^{67}Cu by enriched ^{70}Zn targets: first measurements of formation cross sections of ^{67}Cu , ^{64}Cu , ^{67}Ga , ^{66}Ga , $^{69\text{m}}\text{Zn}$ and ^{65}Zn in interactions of ^{70}Zn with protons above 45 MeV[J]. Radiochimica Acta, 2020, 108(8): 593-602. DOI: 10.1515/ract-2019-3199
 17. Medvedev D G, Mausner L F, Meinken G E, et al. Development of a large scale production of ^{67}Cu from ^{68}Zn at the high energy proton accelerator: closing the ^{68}Zn cycle[J]. Applied Radiation and Isotopes, 2012, 70(3): 423-429. DOI: 10.1016/j.apradiso.2011.10.007
 18. Ohya T, Nagatsu K, Hanyu M, et al. Simple separation of ^{67}Cu from bulk zinc by coprecipitation using hydrogen sulfide gas and silver nitrate[J]. Radiochimica Acta, 2020, 108(6): 469-476. DOI: 10.1515/ract-2019-3168
 19. IAEA. Therapeutic Radiopharmaceuticals Labelled with Copper-67, Rhenium-186 and Scandium-47[M]. IAEA, 2021.
 20. Mirick G R, T O'Donnell R, DeNardo S J, et al. Transfer of copper from a chelated ^{67}Cu -antibody conjugate to ceruloplasmin in lymphoma patients[J]. Nuclear medicine and biology, 1999, 26(7): 841-845. DOI: 10.1016/s0969-8051(99)00049-9
 21. Boros E, Holland J P. Chemical aspects of metal ion chelation in the synthesis and application antibody-based radiotracers[J]. Journal of Labelled Compounds and Radiopharmaceuticals, 2018, 61(9): 652-671. DOI: 10.1002/jlcr.3590
 22. Lewis J S, Lewis M R, Srinivasan A, et al. Comparison of four ^{64}Cu -labeled somatostatin analogues in vitro and in a tumor-bearing rat model: evaluation of new derivatives for positron emission tomography imaging and targeted radiotherapy[J]. Journal of medicinal chemistry, 1999, 42(8): 1341-1347. DOI: 10.1021/jm980602h

23. Navarro A S, Le Bihan T, Le Saëc P, et al. TE1PA as innovating chelator for ^{64}Cu immuno-TEP imaging: A comparative in vivo study with DOTA/NOTA by conjugation on 9E7. 4 mAb in a syngeneic multiple myeloma model[J]. *Bioconjugate chemistry*, 2019, 30(9): DOI: 10.1021/acs.bioconjchem.9b00510
24. Bass L A, Wang M, Welch M J, et al. In vivo transchelation of copper-64 from TETA-octreotide to superoxide dismutase in rat liver[J]. *Bioconjugate chemistry*, 2000, 11(4): 527-532. DOI: 10.1021/bc990167l
25. De Nardo L, Pupillo G, Mou L, et al. A feasibility study of the therapeutic application of a mixture of $^{67}/^{64}\text{Cu}$ radioisotopes produced by cyclotrons with proton irradiation[J]. *Medical Physics*, 2022, 49(4): 2709-2724. DOI: 10.1002/mp.15524
26. TE W. The curability of tumours of differing size by targeted radiotherapy using ^{131}I or ^{90}Y [J]. *Radiother Oncol*, 1991, 21: 91-99. DOI: 10.1016/0167-8140(91)90080-z
27. O'Donoghue J A, Bardies M, Wheldon T E. Relationships between tumor size and curability for uniformly targeted therapy with beta-emitting radionuclides[J]. *Journal of Nuclear Medicine*, 1995, 36(10): 1902-1909. PMID: 7562062
28. Dearling J L J, van Dam E M, Harris M J, et al. Detection and therapy of neuroblastoma minimal residual disease using $^{64}/^{67}\text{Cu}$ Cu-SARTATE in a preclinical model of hepatic metastases[J]. *EJNMMI research*, 2021, 11: 1-14. DOI: 10.1186/s13550-021-00763-0
29. Raponi S, Stefania De Propriis M, Intoppa S, et al. Flow cytometric study of potential target antigens (CD19, CD20, CD22, CD33) for antibody-based immunotherapy in acute lymphoblastic leukemia: analysis of 552 cases[J]. *Leukemia & lymphoma*, 2011, 52(6): 1098-1107. DOI: 10.3109/10428194.2011.559668
30. Thomas D A, Cortes J, O'Brien S, et al. Update of the Modified Hyper-CVAD Regimen with or without Rituximab in Newly Diagnosed Adult Acute Lymphocytic Leukemia (ALL)[J]. *Blood*, 2005, 106(11): 1831. DOI: 10.1182/blood.V106.11.1831.1831
31. Dworzak M N, Schumich A, Printz D, et al. CD20 up-regulation in pediatric B-cell precursor acute lymphoblastic leukemia during induction treatment: setting the stage anti-CD20 directed immunotherapy[J]. *Blood*, The Journal of the American Society of Hematology, 2008, 112(10): 3982-3988. DOI: 10.1182/blood-2008-06-164129
32. DeAngelo D J, Stock W, Stein A S, et al. Inotuzumab ozogamicin in adults with relapsed or refractory CD22-positive acute lymphoblastic leukemia: a phase 1/2 study[J]. *Blood advances*, 2017, 1(15): 1167-1180. DOI: 10.1182/bloodadvances.2016001925

33. Pan J, Niu Q, Deng B, et al. CD22 CAR T-cell therapy in refractory or relapsed B acute lymphoblastic leukemia[J]. *Leukemia*, 2019, 33(12): 2854-2866. DOI: 10.1038/s41375-019-0488-7
34. Ma J, Ge Z. Recent advances of targeted therapy in relapsed/refractory acute myeloid leukemia[J]. *Bosnian Journal of Basic Medical Sciences*, 2021, 21(4): 409. DOI: 10.17305/bjbms.2020.5485
35. Tabata R, Chi S G, Yuda J, et al. Emerging immunotherapy for acute myeloid leukemia[J]. *International Journal of Molecular Sciences*, 2021, 22(4): 1944. DOI: 10.3390/ijms22041944
36. Lam S S Y, Leung A Y H. Overcoming resistance to FLT3 inhibitors in the treatment of FLT3-mutated AML[J]. *International Journal of Molecular Sciences*, 2020, 21(4): 1537. DOI: 10.3390/ijms21041537
37. Man L M, Morris A L, Keng M. New therapeutic strategies in acute lymphocytic leukemia[J]. *Current hematologic malignancy reports*, 2017, 12: 197-206. DOI: 10.1007/s11899-017-0380-3
38. Jabbour E, Cortes J E, Giles F J, et al. Current and emerging treatment options in chronic myeloid leukemia[J]. *Cancer*, 2007, 109(11): 2171-2181. DOI: 10.1002/cncr.22661
39. Boros E, Packard A B. Radioactive transition metals for imaging and therapy[J]. *Chemical reviews*, 2018, 119(2): 870-901. DOI: 10.1021/acs.chemrev.8b00281
40. Kwon G S. Polymeric micelles for delivery of poorly water-soluble compounds[J]. *Critical Reviews™ in Therapeutic Drug Carrier Systems*, 2003, 20(5). DOI: 10.1615/critrevtherdrugcarriersyst.v20.i5.20
41. Sharma M, Hire R S, Hadapad A B, et al. PEGylation enhances mosquito-larvicidal activity of *Lysinibacillus sphaericus* binary toxin[J]. *Bioconjugate Chemistry*, 2017, 28(2): DOI: 10.1021/acs.bioconjchem.6b00565
42. Noach E J K, Ausema A, Dillingh J H, et al. Growth factor treatment prior to low-dose total body irradiation increases donor cell engraftment after bone marrow transplantation in mice[J]. *Blood, The Journal of the American Society of Hematology*, 2002, 100(1): 312-317.
43. Curtis Andrew Lachowicz et al. A phase Ib/II study of ivosidenib with venetoclax +/- azacitidine in IDH1-mutated myeloid malignancies. *JCO* 39, 7012-7012(2021). DOI: 10.1158/2643-3230.BCD-22-0205
44. Chhabra A, Thakur M L. Theragnostic radionuclide pairs for prostate cancer management: $^{64}\text{Cu}/^{67}\text{Cu}$, can be a budding hot duo[J]. *Biomedicines*, 2022, 10(11): 2787. DOI: 10.3390/biomedicines10112787
45. Peters S M B, Mink M C T, Privé B M, et al. Optimization of the radiation dosimetry protocol in Lutetium-177-PSMA therapy: toward clinical im-

- plementation[J]. EJNMMI research, 2023, 13(1): 6. DOI: 10.1186/s13550-023-00952-z
46. Hicks R J, Jackson P, Kong G, et al. ^{64}Cu -SARTATE PET imaging of patients with neuroendocrine tumors demonstrates high tumor uptake and retention, potentially allowing prospective dosimetry for peptide receptor radionuclide therapy[J]. Journal of Nuclear Medicine, 2019, 60(6): 777-785. DOI: 10.2967/jnumed.118.217745
 47. Alcocer-Ávila M E, Ferreira A, Quinto M A, et al. Radiation doses from ^{161}Tb and ^{177}Lu in single tumour cells and micrometastases[J]. EJNMMI physics, 2020, 7: 1-9. DOI: 10.1186/s40658-020-00301-2
 48. Schaefer-Schuler A, Burgard C, Blickle A, et al. [^{161}Tb] Tb-PSMA-617 radioligand therapy in patients with mCRPC: preliminary dosimetry results and intra-individual head-to-head comparison to [^{177}Lu] Lu-PSMA-617[J]. Theranostics, 2024, 14(5): 1829. DOI: 10.7150/thno.92273
 49. Bidkar A P, Zerefa L, Yadav S, et al. Actinium-225 targeted alpha particle therapy for prostate cancer[J]. Theranostics, 2024, 14(7): 2969. DOI: 10.7150/thno.96403
 50. Sathekege M M, Lawal I O, Bal C, et al. Actinium-225-PSMA radioligand therapy of metastatic castration-resistant prostate cancer (WARMTH Act): a multicentre, retrospective study[J]. The lancet oncology, 2024, 25(2): 175-183. DOI: 10.1016/S1470-2045(23)00638-1
 51. Verburg F A, de Blois E, Koolen S, et al. Replacing Lu-177 with Tb-161 in DOTA-TATE and PSMA-617 therapy: potential dosimetric implications for activity selection[J]. EJNMMI physics, 2023, 10(1): 69. DOI: 10.1186/s40658-023-00589-w
 52. Feng Y, Zalutsky M R. Production, purification and availability of ^{211}At : Near term steps towards global access[J]. Nuclear medicine and biology, 2021, 100: 12-23. DOI: 10.1016/j.nucmedbio.2021.05.007
 53. Nigron E, Guertin A, Haddad F, et al. Is ^{70}Zn (d, x) ^{67}Cu the best way to produce ^{67}Cu for medical applications?[J]. Frontiers in Medicine, 2021, 8: 674617. DOI: 10.3389/fmed.2021.674617
 54. Jalilian A R, Gizawy M A, Alliot C, et al. IAEA activities on ^{67}Cu , ^{186}Re , ^{47}Sc theranostic radionuclides radiopharmaceuticals[J]. Current Radiopharmaceuticals, 2021, 14(4): DOI: 10.2174/1874471013999200928162322
 55. Ehst D A, Smith N A, Bowers D L, et al. Copper-67 production on electron linacs—photonuclear technology development[C]//AIP Conference Proceedings. American Institute of Physics, 2012, 1509(1): 157-161. DOI: 10.1063/1.4773959
 56. Jin Z H, Furukawa T, Degardin M, et al. $\alpha_V\beta_3$ Integrin-targeted radionuclide therapy with ^{64}Cu -cyclam-RAFT-c (-RGDfK-) 4[J]. Molecular Can-

- cer Therapeutics, 2016, 15(9): 2076-2085. DOI: 10.1158/1535-7163.MCT-16-0040
57. Hao G, Mastren T, Silvers W, et al. Copper-67 radioimmunotheranostics for simultaneous immunotherapy and immuno-SPECT[J]. Scientific reports, 2021, 11(1): 3622. DOI: 10.1038/s41598-021-82812-1
58. Rudd S E, Van Zuylekom J, Cullinane C, et al. Potential theranostics of breast cancer with copper-64/67 sarcophagine-trastuzumab[J]. Chemical Science, 2025, 16(9): 3998-4005. DOI: 10.1039/d4sc06969b
59. Pougoue Ketchemen J, Njotu F N, Babeker H, et al. Effectiveness of [^{67}Cu] Cu-trastuzumab as a theranostic against HER2-positive breast cancer[J]. European journal of nuclear medicine and molecular imaging, 2024, 51(7): 2070-2084. DOI: 10.1007/s00259-024-06648-3

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