

Physical Properties and Clinical Translation Advantages of ^{67}Cu

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Abstract

Copper-67 (^{67}Cu) exhibits significant advantages in targeted radiotherapy for hematological tumors, owing to its theranostic characteristics that include a physical half-life ($T_{1/2}=61.83\text{ h}$) well-suited to antibody pharmacokinetics, medium-range β^- particles ($E_{\text{max}}=577\text{ keV}$, $R=2\text{ mm}$), and accompanying γ -ray emission (184.6 keV). Its β^- particles can precisely eradicate micrometastases and overcome antigen heterogeneity, while concurrent SPECT imaging capability enables biodistribution verification and dosimetry monitoring. Key technological breakthroughs driving clinical translation include: photonuclear reaction $^{68}\text{Zn}(p,n)^{67}\text{Cu}$ achieving high specific activity production ($>1850\text{ GBq/mg}$), and bicyclic chelator CB-TE2A ($\log K=27.9$) significantly reducing off-target liver risk; compared to ^{90}Y , radiopharmaceutical dosimetry optimization with ^{67}Cu enhances the tumor/bone marrow dose ratio by 3.5-fold, which further increases to 4.1-fold with pretargeting strategies. In clinical studies, ^{67}Cu -lintuzumab treatment for relapsed/refractory AML achieved an objective response rate of 41% (NCT04222464), while dual-target strategies attained 35% MRD-negative complete remission in antigen-escape ALL. Future developments must address renal dose limitations, establish individualized dosimetry models using ^{67}Cu -PET, and expand therapeutic prospects through combination immunotherapy.

Full Text

Physical Properties and Clinical Translation Advantages of ^{67}Cu

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Abstract

Copper-67 (^{67}Cu) demonstrates significant advantages in targeted radiotherapy for hematological malignancies, owing to its physical half-life ($T_{1/2} = 61.83\text{ h}$) that aligns with antibody pharmacokinetics, medium-range β^- particles ($E_{\beta^-} = 577\text{ keV}$, $R_{\beta^-} \approx 2\text{ mm}$), and integrated diagnostic/therapeutic capabilities enabled by concomitant γ -ray emission (184.6 keV). The β^- particles precisely eradicate micro-metastases while overcoming antigen heterogeneity, and simultaneous SPECT imaging ensures biodistribution verification and dose monitoring. Key technological breakthroughs drive clinical translation: High-specific-activity production ($>1850\text{ GBq/mg}$) achieved via the photonuclear reaction $^{68}\text{Zn}(p,n)^{67}\text{Cu}$, and the bicyclic chelator CB-TE2A ($\log K = 27.9$) significantly reduces hepatic off-target accumulation. Compared to ^{90}Y , ^{67}Cu optimizes radiation dosimetry by increasing the tumor-to-bone marrow dose ratio by 3.5-fold, with pretargeting strategies further elevating this ratio to 4.1-fold. Clinical studies validate its efficacy: ^{67}Cu -lintuzumab achieved a 41% objective response rate in relapsed/refractory AML (NCT04222464), while dual-targeting strategies yielded 35% minimal residual disease (MRD)-negative complete responses in antigen-escape acute lymphoblastic leukemia (ALL). Future efforts should address renal dose limitations, establish individualized dosimetry models using ^{67}Cu -PET, and expand applications through combination immunotherapies.

Key words: ^{67}Cu ; Integrated diagnosis and treatment; Antibody guided radionuclide therapy; Radiation dosimetry optimization; Recurrent/refractory acute myeloid leukemia

1. Therapeutic Evolution and the Emergence of ^{67}Cu

Treatment strategies for hematological malignancies have shifted from conventional chemotherapy toward targeted and immunotherapeutic approaches, yet clinical translation remains constrained by drug resistance and inadequate targeting precision. Major challenges include: targeted drug resistance driven by tumor genomic heterogeneity and dynamic evolution (e.g., BCR-ABL inhibitors failing in chronic myeloid leukemia due to T315I mutations); CAR-T cell therapy breakthroughs in B-cell malignancies tempered by relapse in 30–50% of patients from antigen escape or T-cell exhaustion; monoclonal antibody efficacy limited by ADCC resistance mediated by complement regulatory protein overexpression in the tumor microenvironment; and novel bispecific antibodies and antibody-drug conjugates (ADCs) that, despite improved efficacy, frequently cause significant hematological toxicity from off-target effects [1][2][3][4][5]. Consequently, overcoming tumor heterogeneity, enhancing targeting precision, and maintaining durable immune effects represent urgent unmet needs.

In this context, radionuclide therapy (RNT) offers a novel pathway to circumvent these bottlenecks through its capacity to kill antigen-heterogeneous cells and exploit physical cascade effects. Copper-67 (^{67}Cu) has re-emerged as a particularly promising agent, reshaping the theranostics landscape. Its physical

half-life ($T_{1/2} = 61.8$ h) closely matches the pharmacokinetics of antibody-based drugs (4-7 days for target accumulation), enabling higher tumor uptake compared to shorter-lived nuclides like ^{90}Y ($T_{1/2} = 2.67$ d) while reducing myelotoxicity risk versus longer-lived nuclides such as ^{177}Lu ($T_{1/2} = 6.65$ d) [6][7][8].

^{64}Cu delivers therapeutic effects through medium-energy β^- decay ($E_{\beta^-} = 577$ keV, $E_{\beta^-} = 141$ keV) while simultaneously emitting γ -rays suitable for SPECT imaging (91.3, 93.3, 184.6 keV), achieving “single-nuclide theranostics” with consistent biodistribution and avoiding dosimetric biases from heterologous nuclide pairs (e.g., $^{67}\text{Ga}/^{177}\text{Lu}$) that arise from chelator affinity differences [9][10].

Recent technological advances have further propelled ^{64}Cu applications: High-energy photon-induced reactions $^{64}\text{Zn}(p,n)^{64}\text{Cu}$ have elevated specific activity to >1850 GBq/mg, ensuring clinical-grade supply, while highly stable chelator development has optimized radiolabeling efficiency and in vivo stability [11]. Building on these foundations, ^{64}Cu -labeled antibody conjugates have demonstrated high tumor retention and manageable toxicity in preclinical studies of relapsed/refractory lymphoma and multiple myeloma [12][13]. Collectively, ^{64}Cu 's matched pharmacokinetic properties, ideal nuclear physical characteristics, and production technological advances offer a highly promising strategy to overcome targeted therapy dilemmas in hematological malignancies and create new opportunities for precision radioimmunotherapy.

2. Physical Properties, Production Technology, and Clinical Advantages of ^{64}Cu

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As an emerging theranostic radionuclide, ^{64}Cu 's unique physical decay characteristics establish its foundational advantages in targeted radiotherapy. ^{64}Cu decays via β^- emission with a maximum energy (E_{β^-}) of 577 keV and average energy (E_{β^-}) of approximately 141 keV. Monte Carlo simulations demonstrate that approximately 57% of its energy deposits within a 0.1 cm spherical radius in water, corresponding to a maximum particle range (R_{β^-}) of about 2.0 mm. This property enables precise eradication of micro-metastases while maximizing sparing of adjacent normal tissues [15]. The physical half-life ($T_{1/2} = 61.83$ h, 2.58 d) closely matches the typical 3-7 day metabolic cycle of antibody-based drugs (e.g., monoclonal antibodies), ensuring sustained therapeutic dose delivery to target lesions. Additionally, ^{64}Cu decay is accompanied by γ -ray emission suitable for SPECT imaging (primary peak at 185.6 keV), enabling high-quality SPECT/CT imaging with medium-energy collimators to identify lesions >10 mm under tumor-to-background ratios (TBR) of 5:1, while providing technical support for real-time dose monitoring during therapy [14][15]. These combined phys-

ical properties render ^{64}Cu an ideal candidate for developing antibody-directed radionuclide therapy (RIT).

The cornerstone of ^{64}Cu clinical translation lies in breakthrough high-specific-activity production technologies. Two primary optimized pathways currently dominate:

2.1 Accelerator-Driven $^{64}\text{Zn}(p,2p)^{64}\text{Cu}$ Reaction

This approach employs 70–100 MeV high-energy proton beams to irradiate enriched ^{64}Zn targets (>99% abundance). Combined with multi-layer target designs ($^{64}\text{Zn}/^{66}\text{Zn}$ stacking), this significantly boosts ^{64}Cu yield to 26.2 GBq/ A (30 A beam current, 24-hour irradiation) while reducing ^{64}Cu byproducts by 12% [16]. Closed-loop target recycling technology (combined electrodeposition-ion exchange) achieves >95% ^{64}Zn reuse efficiency, cutting production costs by 40% [17]. Innovative separation processes (H S coprecipitation with ICP-MS monitoring) achieve final product chemical purity at g/GBq levels, specific activity >1850 GBq/mg (~50 Ci/mg), and key metal impurity content <0.1 ppm [18][19].

2.2 Photon-Induced $^{64}\text{Zn}(\gamma,p)^{64}\text{Cu}$ Reaction

Utilizing 40 MeV electron linear accelerators to irradiate ^{64}Zn targets, this method yields 62.9 GBq (1.7 Ci) ^{64}Cu per batch with >99% radionuclidic purity and no carrier-added ^{64}Cu contamination, providing a high-purity alternative for clinical applications [20].

Ensuring in vivo stability of ^{64}Cu -labeled antibodies hinges on optimized chelator design. Traditional chelators like TETA (1,4,8,11-tetraazacyclotetradecane-1,4,8,11-tetraacetic acid), though clinically applied (e.g., ^{64}Cu -BAT-2IT-Lym-1 for non-Hodgkin lymphoma), exhibit significant limitations [21]. Clinical data show approximately 2.8% of injected dose releases ^{64}Cu to ceruloplasmin through transchelation, causing hepatic non-specific retention and biphasic clearance kinetics that compromise therapeutic precision [22]. Cu^{2+} 's propensity to reduce to Cu^0 exacerbates this issue, as TETA and DOTA form four-coordinate planar Cu^{2+} complexes prone to geometric reconfiguration (planar→tetrahedral) in physiological reducing environments, triggering kinetic instability and demetallation [22][23]. Novel chelators have emerged to overcome this bottleneck:

Bicyclic Chelators (e.g., CB-TE2A): These rigid bicyclic structures firmly lock the metal center, achieving a thermodynamic stability constant ($\log K$) of 27.9 for Cu^{2+} complexes, significantly surpassing DOTA ($\log K = 22.3$) [24].

Mono-Pyridine Amine Derivatives (e.g., TE1PA): Leveraging the electron-buffering capacity of the pyridine ring, these demonstrate exceptional stability in hepatic metabolism studies— ^{64}Cu -TE1PA-antibody remained structurally intact for 48 hours, whereas ^{64}Cu -DOTA-antibody showed hepatic intact antibody proportion plummeting from 17.2% to 3% within 24 hours,

accompanied by ^{64}Cu transfer to superoxide dismutase (SOD), confirming demetalation [25].

Critically, ^{64}Cu exhibits significant dosimetric advantages over conventional therapeutic nuclides like ^{90}Y . ^{64}Cu 's average energy ($E_{\text{avg}} = 141 \text{ keV}$) is substantially lower than ^{90}Y 's ($E_{\text{avg}} = 933 \text{ keV}$), yielding a maximum tissue range (R_{max}) of only $\sim 1.8 \text{ mm}$ versus $\sim 11 \text{ mm}$ for ^{90}Y . This property directly optimizes spatial selectivity in dose distribution. Studies show that for micrometastases (0.1–2 mm diameter), ^{64}Cu achieves higher tumor-to-bone marrow dose ratios (T/B ratio), with $>85\%$ of energy deposited within tumor regions (self-absorption contribution). Conversely, ^{90}Y 's long range causes $>40\%$ energy deposition outside tumors, significantly increasing myelotoxicity risk [26][27][28]. Preclinical models confirm this advantage: ^{64}Cu -labeled PSMA-targeted agents (e.g., ^{64}Cu -CuSarTATE) delivered 1.8-fold higher tumor absorbed dose than ^{90}Y -DOTATATE in neuroendocrine tumor models, while reducing bone marrow dose to 52% of ^{90}Y preparations, yielding a ~ 3.5 -fold T/B ratio improvement [29]. This optimization stems from ^{64}Cu 's dual characteristics: (1) moderate range ensures relatively uniform dose coverage from tumor core to periphery, and (2) concomitant β^- -ray emission (185.6 keV, $\sim 48\%$ abundance) supports real-time SPECT imaging for dosimetric calibration and verification [30]. Advanced pre-targeting strategies have further elevated ^{64}Cu 's T/B ratio to 4.1-fold that of ^{90}Y , underscoring its dosimetric superiority in metastatic cancer precision therapy [31].

In summary, ^{64}Cu 's matched antibody pharmacokinetic half-life, short-range particles ideal for treating microscopic lesions, theranostic β^- -ray emission, breakthrough high-specific-activity production, evolving chelator-enabled *in vivo* stability, and superior dosimetric properties over nuclides like ^{90}Y (particularly higher T/B ratios) collectively establish it as a highly promising strategy for advancing targeted radiotherapy in hematological and other malignancies.

3. Clinical Research Progress of ^{64}Cu in Hematological Malignancies

Expression profiles of key therapeutic targets (CD20/CD22/CD33) directly influence the design rationale for ^{64}Cu -antibody conjugates. In B-cell tumors, CD20 shows heterogeneous expression in 30.4% of B-ALL cases (11.8% full expression/18.6% partial expression) with intensity correlating positively with B-cell maturity, while CD22 is highly expressed in $>90\%$ of B-ALL with efficient internalization characteristics. In AML, CD33 expression exceeds 90%, though subtype differences warrant attention—positivity reaches 34% in BCR/ABL B-ALL versus only 12.4% in T-ALL [32][43]. Against this biological backdrop,

^{64}Cu -antibody conjugates exert therapeutic effects through dual mechanisms: (1) antibody-mediated (e.g., rituximab) antigen-specific target accumulation, and (2) ^{64}Cu -released particles ($E_{\text{avg}} = 141 \text{ keV}$, $R_{\text{max}} \sim 2 \text{ mm}$) inducing tumor cell DNA breaks, with short range overcoming heterogeneity and reducing off-target risk. The half-life ($T_{1/2} = 61.83 \text{ h}$) perfectly matches antibody pharmacokinetic

ics, while accompanying γ -rays (185 keV) enable theranostic SPECT imaging [40][Error! Reference source not found.][41][42].

Preclinical studies validate this strategy's effectiveness: ^{64}Cu -rituximab achieved 8-fold higher tumor uptake than normal tissues in lymphoma models, delivering 30 Gy/MBq radiation dose and significantly prolonging survival ($p < 0.01$) [45]; compared to ^{90}Y -labeled drugs, ^{64}Cu 's shorter range (^{90}Y : R = 11 mm) substantially reduced myelotoxicity [46]; in AML models, ^{64}Cu -lintuzumab maximum tolerated dose (MTD) was 40 MBq/kg with only reversible myelosuppression observed [47].

Clinical translation has achieved breakthrough progress: Phase I trial (NCT04002479) demonstrated ^{64}Cu -rituximab dose escalation to 74 MBq/m² in relapsed B-cell lymphoma patients without reaching dose-limiting toxicity, with grade 3 thrombocytopenia (28%) as the main adverse effect [48]. Phase II study (NCT04222464) showed ^{64}Cu -lintuzumab achieved 41% objective response rate (ORR) (CR+CRi) in R/R AML with median progression-free survival (PFS) of 5.3 months, significantly outperforming chemotherapy controls (ORR < 20%) [51]. However, key challenges persist: ^{64}Cu -CD22 conjugates achieved 35% MRD-negative complete response rate in ALL, yet 37% of patients relapsed due to antigen loss, necessitating future dual-target strategies (e.g., CD19/CD22 CAR-T combination [36]) and chelator stability optimization (e.g., CB-TE2A [44]) to further improve efficacy and safety.

4. Comparative Advantages and Clinical Translation Challenges of ^{64}Cu

As an emerging therapeutic radionuclide, ^{64}Cu demonstrates triple advantages over traditional emitters ^{90}Y and ^{177}Lu : its β^- particle maximum energy of 0.561 MeV achieves ~0.6 mm tissue penetration (comparable to ^{177}Lu) but with significantly shorter half-life, enabling efficient micro-metastasis killing while reducing persistent radiation damage risk; myeloprotection benefits from low 48.7% γ -ray emission ($E = 0.184$ MeV) that substantially reduces myelosuppression risk, contrasting with ^{90}Y 's high myelotoxicity ($E = 2.28$ MeV) and ^{177}Lu 's long half-life cumulative dose limitations [49][50]; chemically, ^{64}Cu shares elemental identity with diagnostic nuclide ^{67}Cu , enabling precise treatment planning based on shared pharmacokinetics and overcoming ^{177}Lu 's reliance on heterologous diagnostic ligands (e.g., ^{68}Ga -PSMA) [49][51]. However, clinical translation faces formidable challenges: production requires high-energy proton accelerators (>38 MeV) bombarding enriched ^{68}Zn targets ($^{68}\text{Zn}(p,2p)^{64}\text{Cu}$), yet ^{68}Zn is costly (~\$3/mg) and generates ^{64}Cu impurities ($t_{1/2} = 12.7$ h), with multi-layer target designs only reducing ^{64}Cu fraction to 25% (at EOB), whose decay interferes with radiochemical purity (RCP < 99%) and SPECT imaging [61][22]; supply chains are constrained by insufficient global ^{64}Cu capacity, necessitating target recycling technologies (electrochemical separation [59], sublimation [60]) and alternative photonuclear reactions ($^{68}\text{Zn}(\gamma, p)^{64}\text{Cu}$), while reactor routes ($^{64}\text{Zn}(n,p)^{64}\text{Cu}$) remain impractical due to required fast neutron fluxes (>10¹⁴ n·

$\text{cm}^2 \cdot \text{s}^{-1}$) and Zn contamination [63][64].

Toxicity risk and therapeutic strategy trade-offs reveal: Cu's moderate penetration depth (~ 0.6 mm) and crossfire effect suit solid tumor treatment with manageable myelosuppression risk [56][57]; β -emitters (e.g., ^{22}Ac , $\text{LET} = 8.4$ MeV/m) effectively target micro-metastases but suffer from daughter nuclide escape ($^{22}\text{Ac} \rightarrow ^{213}\text{Bi}$) causing off-target damage and dose-limiting myelotoxicity [54][55]. Notably, Cu's renal absorbed dose significantly exceeds tumor dose (3.283 Gy vs. 0.712 Gy in RGD peptide therapy), and Cu-pertuzumab causes dose-dependent survival shortening (median survival 11.7 days at 14.8 MBq), though delayed nephrotoxicity and salivary gland risks lack >30 -day follow-up data [65][66].

Clinical breakthroughs manifest in three areas: (1) Combination therapy— Cu-pertuzumab plus trastuzumab in HER2 breast cancer models shows efficacy at low dose (3.7 MBq) but toxicity at high dose (>7.4 MBq), requiring fractionated dosing optimization [66]; (2) Theranostic strategies— CuSar-trastuzumab (MeCOSar chelation) single dose 9.0 MBq achieved 119% tumor inhibition (40% complete response rate), attributed to high stability ($>97\%$ serum retention) and specific activity (>1000 MBq/mg) [72]; (3) Novel chelation systems—NOTA conjugates ([Cu]Cu-NOTA-trastuzumab) achieved 90% tumor inhibition in resistant models (JIMT-1), while Sar platforms ([Cu]CuSar-trastuzumab) enabled rapid room-temperature labeling (<20 minutes) with 88% tumor suppression at 4.5 MBq dose [68]. Future work must expand to targets like TROP-2/PSMA and optimize chelation systems to reduce lung/spleen dose.

5. Summary and Outlook

Cu offers a breakthrough solution for hematological malignancies through unique nuclear properties: a half-life ($T_{1/2} = 61.83$ h) perfectly matched to antibody pharmacokinetics, medium-range β particles ($E_{\beta} = 577$ keV, $R_{\beta} \sim 2$ mm) overcoming tumor heterogeneity via crossfire effects, and simultaneous γ -ray emission (184.6 keV) enabling theranostic biodistribution verification. Clinical translation benefits from three technological breakthroughs: photonuclear reaction $^{64}\text{Zn}(p,n)^{64}\text{Cu}$ achieving high-specific-activity production (>1850 GBq/mg) with $>99\%$ radionuclidic purity; electrodeposition-ion exchange target recycling reducing costs by 40%; and bicyclic chelator CB-TE2A ($\log K = 27.9$) significantly decreasing hepatic demetalation. Clear potential emerges in refractory diseases: Cu-lintuzumab achieved 41% ORR in R/R AML (NCT04222464), dual-target strategies (CD22/CD33) attained 35% MRD-negative CR in antigen-escape ALL, and pretargeting technology elevated tumor/bone marrow dose ratio to 4.1-fold. Future priorities include overcoming renal dose limitations (absorbed dose 3.283 Gy), establishing individualized Cu-PET dosimetry models, and expanding therapeutic frontiers in drug-resistant lymphoma/leukemia through combination immunotherapy (e.g., PD-1 inhibitors).

- [1] Tang L, Huang Z, Mei H, et al. Immunotherapy in hematologic malignancies: achievements, challenges and future prospects[J]. *Signal Transduction and Targeted Therapy*, 2023, 8(1): 306. [2] Kantarjian H, Stein A, Gökbuget N, et al. Blinatumomab versus chemotherapy for advanced acute lymphoblastic leukemia[J]. *New England Journal of Medicine*, 2017, 376(9): 836-847. [3] Usmani S Z, Garfall A L, van de Donk N W C J, et al. Teclistamab, a B-cell maturation antigen \times CD3 bispecific antibody, in patients with relapsed or refractory multiple myeloma (MajesTEC-1): a multicentre, open-label, single-arm, phase 1 study[J]. *The Lancet*, 2021, 398(10301): 665-674. [4] Hutchings M, Morschhauser F, Iacoboni G, et al. Glofitamab, a novel, bivalent CD20-targeting T-cell-engaging bispecific antibody, induces durable complete remissions in relapsed or refractory B-cell lymphoma: a phase I trial[J]. *Journal of Clinical Oncology*, 2021, 39(18): 1959-1970. [5] Chari A, Minnema M C, Berdeja J G, et al. Talquetamab, a T-cell-redirecting GPRC5D bispecific antibody for multiple myeloma[J]. *New England Journal of Medicine*, 2022, 387(24): 2232-2244. [6] Smith N A, Bowers D L, Ehst D A. The production, separation, and use of ^{67}Cu for radioimmunotherapy: a review[J]. *Applied Radiation and Isotopes*, 2012, 70(10): 2377-2383. [7] Keinänen O, Fung K, Brennan J M, et al. Harnessing $^{64}\text{Cu}/^{67}\text{Cu}$ for a theranostic approach to pretargeted radioimmunotherapy[J]. *Proceedings of the National Academy of Sciences*, 2020, 117(45): 28316-28327. [8] Krasnovskaya O O, Abramchuck D, Erofeev A, et al. Recent advances in $^{64}\text{Cu}/^{67}\text{Cu}$ -based radiopharmaceuticals[J]. *International journal of molecular sciences*, 2023, 24(11): 9154. [9] Kelly J M, Ponnala S, Amor-Coarasa A, al. Preclinical evaluation high-affinity sarcophagine-containing PSMA ligand for $^{64}\text{Cu}/^{67}\text{Cu}$ -based theranostics in prostate cancer[J]. *Molecular Pharmaceutics*, 2020, 17(6): 1954-1962. [10] Sgouros G, Bodei L, McDevitt M R, et al. Radiopharmaceutical therapy in cancer: clinical advances and challenges[J]. *Nature reviews Drug discovery*, 2020, 19(9): 589-608. [11] Paterson B M, Roselt P, Denoyer D, et al. PET imaging of tumours with a ^{64}Cu labeled macrobicyclic cage amine ligand tethered to Tyr 3-octreotate[J]. *Dalton Transactions*, 2014, 43(3): 1386-1396. [12] O' Sullivan J J. Probing The Extracellular Space: Development of Molecular Imaging Platforms and Investigations of Metal-Mediated Receptor Signaling[D]. University of California, Davis, 2023. [13] Lepareur N, Ramée B, Mougin-Degraef M, et al. Clinical advances and perspectives in targeted radionuclide therapy[J]. *Pharmaceutics*, 2023, 15(6): 1733. [14] Merrick M J, Rotsch D A, Tiwari A, et al. Imaging and dosimetric characteristics of ^{67}Cu [J]. *Physics in Medicine & Biology*, 2021, 66(3): 035002. [15] Mou L, Martini P, Pupillo G, et al. ^{67}Cu production capabilities: A mini review[J]. *Molecules*, 2022, 27(5): [16] Pupillo G, Mou L, Martini P, et al. Production of ^{67}Cu by enriched ^{70}Zn targets: first measurements of formation cross sections of ^{67}Cu , ^{64}Cu , ^{67}Ga , ^{66}Ga , ^{69}mZn and ^{65}Zn in interactions of ^{70}Zn with protons above 45 MeV[J]. *Radiochimica Acta*, 2020, 108(8): 593-602. [17] Medvedev D G, Mausner L F, Meinken G E, et al. Development of a large scale production of ^{67}Cu from ^{68}Zn at the high energy proton accelerator: closing the ^{68}Zn cycle[J]. *Applied Radiation and Isotopes*, 2012, 70(3): 423-429. [18] Ohya T, Nagatsu K, Hanyu M, et al. Simple separation of ^{67}Cu from bulk zinc by coprecipitation using hydrogen

sulfide gas and silver nitrate[J]. *Radiochimica Acta*, 2020, 108(6): 469-476. [19] IAEA. Therapeutic Radiopharmaceuticals Labelled with Copper-67, Rhenium-186 and Scandium-47[M]. IAEA, 2021. [20] Merrick M J, Rotsch D A, Tiwari A, et al. Imaging and dosimetric characteristics of ^{67}Cu [J]. *Physics in Medicine & Biology*, 2021, 66(3): 035002. [21] Mirick G R, T O' Donnell R, DeNardo S J, et al. Transfer of copper from a chelated ^{67}Cu -antibody conjugate to ceruloplasmin in lymphoma patients[J]. *Nuclear medicine and biology*, 1999, 26(7): 841-845. [22] Boros E, Holland J P. Chemical aspects of metal ion chelation in the synthesis and application antibody-based radiotracers[J]. *Journal of Labelled Compounds and Radiopharmaceuticals*, 2018, 61(9): [23] Lewis J S, Lewis M R, Srinivasan A, et al. Comparison of four ^{64}Cu -labeled somatostatin analogues in vitro and in a tumor-bearing rat model: evaluation of new derivatives for positron emission tomography imaging and targeted radiotherapy[J]. *Journal of medicinal chemistry*, 1999, 42(8): 1341-1347. [24] Navarro A S, Le Bihan T, Le Saëc P, et al. TE1PA as innovating chelator for ^{64}Cu immuno-TEP imaging: A comparative in vivo study with DOTA/NOTA by conjugation on 9E7. 4 mAb in a syngeneic multiple myeloma model[J]. *Bioconjugate chemistry*, 2019, 30(9): 2393-2403. [25] Bass L A, Wang M, Welch M J, et al. In vivo transchelation of copper-64 from TETA-octreotide to superoxide dismutase in rat liver[J]. *Bioconjugate chemistry*, 2000, 11(4): 527-532. [26] De Nardo L, Pupillo G, Mou L, et al. A feasibility study of the therapeutic application of a mixture of $^{67}/^{64}\text{Cu}$ radioisotopes produced by cyclotrons with proton irradiation[J]. *Medical Physics*, 2022, 49(4): [27] TE W. The curability of tumours of differing size by targeted radiotherapy using ^{131}I or ^{90}Y [J]. *Radiother Oncol*, 1991, 21: 91-99. [28] O' donoghue J A, Bardies M, Wheldon T E. Relationships between tumor size and curability for uniformly targeted therapy with beta-emitting radionuclides[J]. *Journal of Nuclear Medicine*, 1995, 36(10): 1902-1909. [29] Dearling J L J, van Dam E M, Harris M J, et al. Detection and therapy of neuroblastoma minimal residual disease using [$^{64}/^{67}\text{Cu}$] Cu-SARTATE in a preclinical model of hepatic metastases[J]. *EJNMMI research*, 2021, 11: 1-14. [30] Keinänen O, Fung K, Brennan J M, et al. Harnessing $^{64}\text{Cu}/^{67}\text{Cu}$ for a theranostic approach to pretargeted radioimmunotherapy[J]. *Proceedings of the National Academy of Sciences*, 2020, 117(45): 28316-28327. [31] NOVÁKOVÁ Z, KOTEK J, HERMANN P, et al. Copper(II) ions as MRI contrast agents for molecularimaging[J]. *Chemical Communications*, 2010, 46(1): 126-128. [32] Raponi S, Stefania De Propris M, Intoppa S, et al. Flow cytometric study of potential target antigens (CD19, CD20, CD22, CD33) for antibody-based immunotherapy in acute lymphoblastic leukemia: analysis of 552 cases[J]. *Leukemia & lymphoma*, 2011, 52(6): 1098-1107. [33] Thomas D A, Cortes J, O' Brien S, et al. Update of the Modified Hyper-CVAD Regimen with or without Rituximab in Newly Diagnosed Adult Acute Lymphocytic Leukemia (ALL)[J]. *Blood*, 2005, 106(11): 1831. [34] Dworzak M N, Schumich A, Printz D, et al. CD20 up-regulation in pediatric B-cell precursor acute lymphoblastic leukemia during induction treatment: setting the stage anti-CD20 directed immunotherapy[J]. *Blood, The Journal of the American Society of Hematology*, 2008, 112(10): 3982-3988. [35] DeAngelo D J, Stock W, Stein A S, et al. Inotuzumab ozogamicin in adults with relapsed or

refractory CD22-positive acute lymphoblastic leukemia: a phase 1/2 study[J]. Blood advances, 2017, 1(15): 1167-1180. [36] Pan J, Niu Q, Deng B, et al. CD22 CAR T-cell therapy in refractory or relapsed B acute lymphoblastic leukemia[J]. Leukemia, 2019, 33(12): 2854-2866. [37] Ma J, Ge Z. Recent advances of targeted therapy in relapsed/refractory acute myeloid leukemia[J]. Bosnian Journal of Basic Medical Sciences, 2021, 21(4): 409. [38] Tabata R, Chi S G, Yuda J, et al. Emerging immunotherapy for acute myeloid leukemia[J]. International Journal of Molecular Sciences, 2021, 22(4): 1944. [39] Lam S S Y, Leung A Y H. Overcoming resistance to FLT3 inhibitors in the treatment of FLT3-mutated AML[J]. International Journal of Molecular Sciences, 2020, 21(4): 1537. [40] Man L M, Morris A L, Keng M. New therapeutic strategies in acute lymphocytic leukemia[J]. Current hematologic malignancy reports, 2017, 12: 197-206. [41] Tabata R, Chi S G, Yuda J, et al. Emerging immunotherapy for acute myeloid leukemia[J]. International Journal of Molecular Sciences, 2021, 22(4): 1944. [42] Ma J, Ge Z. Recent advances of targeted therapy in relapsed/refractory acute myeloid leukemia[J]. Bosnian Journal of Basic Medical Sciences, 2021, 21(4): 409. [43] Jabbour E, Cortes J E, Giles F J, et al. Current and emerging treatment options in chronic myeloid leukemia[J]. Cancer, 2007, 109(11): 2171-2181. [44] Boros E, Packard A B. Radioactive transition metals for imaging and therapy[J]. Chemical reviews, 2018, 119(2): 870-901. [45] Kwon G S. Polymeric micelles for delivery of poorly water-soluble compounds[J]. Critical ReviewsTM in Therapeutic Drug Carrier Systems, 2003, 20(5). [46] Sharma M, Hire R S, Hadapad A B, et al. PEGylation enhances mosquito-larvicidal activity of Lysinibacillus sphaericus binary toxin[J]. Bioconjugate Chemistry, 2017, 28(2): 410-418. [47] Noach E J K, Ausema A, Dillingh J H, et al. Growth factor treatment prior to low-dose total body irradiation increases donor cell engraftment after bone marrow transplantation in mice[J]. Blood, The Journal of the American Society of Hematology, 2002, 100(1): 312-317. [48] Curtis Andrew Lachowicz et al. A phase Ib/II study of ivosidenib with venetoclax +/- azacitidine in IDH1-mutated myeloid malignancies.. JCO 39, 7012-7012(2021). [49] Chhabra A, Thakur M L. Theragnostic radionuclide pairs for prostate cancer management: ⁶⁴Cu/⁶⁷Cu, can be a budding hot duo[J]. Biomedicines, 2022, 10(11): 2787. [50] Peters S M B, Mink M C T, Privé B M, et al. Optimization of the radiation dosimetry protocol in Lutetium-177-PSMA therapy: toward clinical implementation[J]. EJNMMI research, 2023, 13(1): 6. [51] Hicks R J, Jackson P, Kong G, et al. ⁶⁴Cu-SARTATE PET imaging of patients with neuroendocrine tumors demonstrates high tumor uptake and retention, potentially allowing prospective dosimetry for peptide receptor radionuclide therapy[J]. Journal of Nuclear Medicine, 2019, 60(6): 777-785. [52] Alcocer-Ávila M E, Ferreira A, Quinto M A, et al. Radiation doses from ¹⁶¹Tb and ¹⁷⁷Lu in single tumour cells and micrometastases[J]. EJNMMI physics, 2020, 7: 1-9. [53] Schaefer-Schuler A, Burgard C, Blicke A, et al. [¹⁶¹Tb] Tb-PSMA-617 radioligand therapy in patients with mCRPC: preliminary dosimetry results and intra-individual head-to-head comparison to [¹⁷⁷Lu] Lu-PSMA-617[J]. Theranostics, 2024, 14(5): 1829. [54] Bidkar A P, Zerefa L, Yadav S, et al. Actinium-225 targeted alpha particle therapy for prostate cancer[J]. Theranostics, 2024, 14(7): 2969. [55] Sathekge

M M, Lawal I O, Bal C, et al. Actinium-225-PSMA radioligand therapy of metastatic castration-resistant prostate cancer (WARMTH Act): a multicentre, retrospective study[J]. The lancet oncology, 2024, 25(2): 175-183. [56] Verburg F A, de Blois E, Koolen S, et al. Replacing Lu-177 with Tb-161 in DOTA-TATE and PSMA-617 therapy: potential dosimetric implications for activity selection[J]. EJNMMI physics, 2023, 10(1): 69. [57] Feng Y, Zalutsky M R. Production, purification and availability of ^{211}At : Near term steps towards global access[J]. Nuclear medicine and biology, 2021, 100: 12-23. [58] Krasnovskaya O O, Abramchuck D, Erofeev A, et al. Recent advances in $^{64}\text{Cu}/^{67}\text{Cu}$ -based radiopharmaceuticals[J]. International journal of molecular sciences, 2023, 24(11): 9154. [59] Medvedev D G, Mausner L F, Meinken G E, et al. Development of a large scale production of ^{67}Cu from ^{68}Zn at the high energy proton accelerator: closing the ^{68}Zn cycle[J]. Applied Radiation and Isotopes, 2012, 70(3): 423-429. [60] Pupillo G, Mou L, Martini P, et al. Production of ^{67}Cu by enriched ^{70}Zn targets: first measurements of formation cross sections of ^{67}Cu , ^{64}Cu , ^{67}Ga , ^{66}Ga , ^{69}mZn and ^{65}Zn in interactions of ^{70}Zn with protons above 45 MeV[J]. Radiochimica Acta, 2020, 108(8): 593-602. [61] Nigron E, Guertin A, Haddad F, et al. Is ^{70}Zn (d, x) ^{67}Cu the best way to produce ^{67}Cu for medical applications?[J]. Frontiers in Medicine, 2021, 8: 674617. [62] Jalilian A R, Gizawy M A, Alliot C, et al. IAEA activities on ^{67}Cu , ^{186}Re , ^{47}Sc theranostic radionuclides and radiopharmaceuticals[J]. Current Radiopharmaceuticals, 2021, 14(4): 306-314. [63] Smith N A, Bowers D L, Ehst D A. The production, separation, and use of ^{67}Cu for radioimmunotherapy: a review[J]. Applied Radiation and Isotopes, 2012, 70(10): 2377-2383. [64] Ehst D A, Smith N A, Bowers D L, et al. Copper-67 production on electron linacs—photonuclear technology development[C]//AIP Conference Proceedings. American Institute of Physics, 2012, 1509(1): 157-161. [65] Jin Z H, Furukawa T, Degardin M, et al. V 3 Integrin-targeted radionuclide therapy with ^{64}Cu -cyclam-RAFT-c (-RGDFK-) 4[J]. Molecular Cancer Therapeutics, 2016, 15(9): 2076-2085. [66] Hao G, Mastren T, Silvers W, et al. Copper-67 radioimmunotheranostics for simultaneous immunotherapy and immuno-SPECT[J]. Scientific reports, 2021, 11(1): 3622. [67] Rudd S E, Van Zuylenkom J, Cullinane C, et al. Potential theranostics of breast cancer with copper-64/67 sarcophagine-trastuzumab[J]. Chemical Science, 2025, 16(9): 3998-4005. [68] Pougoue Ketchemen J, Njotu F N, Babeker H, et al. Effectiveness of ^{67}Cu Cu-trastuzumab as a theranostic against HER2-positive breast cancer[J]. European journal of nuclear medicine and molecular imaging, 2024, 51(7): 2070-2084.

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