

Phenotypic plasticity of *Artemisia ordosica* seedlings in response to different levels of calcium carbonate in soil postprint

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Abstract

Plant phenotypic plasticity is a common feature that is crucial for explaining interspecific competition, dynamics and biological evolution of plant communities. In this study, we tested the effects of soil CaCO₃ (calcium carbonate) on the phenotypic plasticity of a psammophyte, *Artemisia ordosica*, an important plant species on sandy lands in arid and semi-arid areas of China, by performing pot experiments under different CaCO₃ contents with a two-factor randomized block design and two orthogonal designs. We analyzed the growth responses (including plant height, root length, shoot-leaf biomass and root biomass) of *A. ordosica* seedlings to different soil CaCO₃ contents. The results revealed that, with a greater soil CaCO₃ content, *A. ordosica* seedlings gradually grew more slowly, with their relative growth rates of plant height, root length, shoot-leaf biomass and root biomass all decreasing significantly. Root N/P ratios showed significant negative correlations with the relative growth rates of plant height, shoot-leaf biomass and root length of *A. ordosica* seedlings; however, the relative growth rate of root length increased significantly with the root P concentration increased, showing a positive correlation. These results demonstrate that soil CaCO₃ reduces the local P availability in soil, which produces a non-adaptive phenotypic plasticity to *A. ordosica* seedlings. This study should prove useful for planning and promoting the restoration of damaged/degraded vegetation in arid and semi-arid areas of China.

Full Text

Preamble

Phenotypic plasticity of *Artemisia ordosica* seedlings in response to different levels of calcium carbonate in soil

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Abstract: Plant phenotypic plasticity is a common feature that is crucial for explaining interspecific competition, community dynamics, and biological evolution in plant communities. In this study, we investigated the effects of soil CaCO₃ (calcium carbonate) on the phenotypic plasticity of *Artemisia ordosica*, a psammophyte species that plays an important role on sandy lands in arid and semi-arid regions of China. We conducted pot experiments using a two-factor randomized block design and two orthogonal designs with varying soil CaCO₃ contents, analyzing growth responses including plant height, root length, shoot-leaf biomass, and root biomass. The results revealed that as soil CaCO₃ content increased, *A. ordosica* seedlings exhibited progressively slower growth, with significant decreases in relative growth rates for plant height, root length, shoot-leaf biomass, and root biomass.

Root N/P ratios showed significant negative correlations with the relative growth rates of plant height, shoot-leaf biomass, and root length in *A. ordosica* seedlings. However, the relative growth rate of root length increased significantly with rising root P concentration, demonstrating a positive correlation. These findings indicate that soil CaCO₃ reduces local P availability, thereby inducing non-adaptive phenotypic plasticity in *A. ordosica* seedlings. This study provides valuable insights for planning and promoting restoration of damaged or degraded vegetation in arid and semi-arid areas of China.

Keywords: *Artemisia ordosica*; N/P ratio; phenotypic plasticity; relative growth rate; soil CaCO₃; soil P availability; arid and semi-arid areas

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1 Introduction

Phenotypic plasticity refers to the phenotypic variation produced by a single genotype in response to different local environmental conditions (Bradshaw, 1965; Lauri et al., 2016; Li et al., 2016). It is inherently a developmental and multivariate phenomenon (Pigliucci et al., 1997). Various environmental factors—including light, temperature, water, heat, and wind—can individually or jointly induce phenotypic plasticity (Sultan, 2001a, b; Pigliucci and Kolodynska, 2002). Importantly, the plasticity of specific functional traits may itself be heritable and thus capable of independent evolution under natural selection (Jain and Bradshaw, 1966; Stearns, 1989; D' Ambrosio and Colagè, 2017). Quantifying phenotypic plasticity in plants can reveal relationships between individual plasticity and ecological and evolutionary patterns (Schlichting, 1986; Bradshaw and Hardwick, 1989; Pigliucci, 2005). Moreover, as sessile organisms, plants rely on phenotypic plasticity to cope with sudden environmental changes (Via et al.,

1995; Pigliucci, 2002; Bossdorf and Pigliucci, 2009), which can facilitate species invasions (Geng et al., 2004) and influence patterns of evolutionary diversity (Ghalambor et al., 2007; Fusco and Minelli, 2010; Storz et al., 2010).

Soils at latitudes 10°–40° in both hemispheres generally contain CaCO₃ (Schinas and Rowell, 1977). Abbas et al. (1991) classified plant species as calcicole (thriving in lime-rich soil), calcifuge (not growing well in lime-rich soil), or CaCO₃-indifferent based on their survival ability in CaCO₃-rich soils. Nearly 57 years ago, Lee and Woolhouse (1971) investigated bicarbonate effects on root growth in calcicole and calcifuge grasses, finding that calcifuge species were more strongly inhibited by soil CaCO₃ than calcicole species. Interestingly, Pedersen et al. (2011) demonstrated in a pot experiment that both the calcicole species *Anisantha tectorum* and the calcifuge species *Rumex acetosella* grew slower in subsoil containing approximately 10% CaCO₃ than in topsoil with less than 1% CaCO₃. Our previous study suggested that the ratio of soil CaCO₃ to available P could significantly influence plant density in *Artemisia ordosica* populations (Zhao et al., 2012), indicating that soil CaCO₃ might induce phenotypic plasticity in this species.

This raises an intriguing question: since CaCO₃ itself is a soil-forming substrate, how might it induce plant phenotypic plasticity? We hypothesized that soil CaCO₃ limits P availability, preventing P absorption by plants and thereby affecting N/P ratios, growth rates, and phenotypic plasticity. In this study, we selected *A. ordosica* for three pot experiments analyzing its phenotypic plasticity under varying soil CaCO₃ contents. Our aims were: (1) to examine the effect of soil CaCO₃ content on phenotypic plasticity in *A. ordosica*, and (2) to explore the mechanisms underlying the relationship between soil CaCO₃ content and plant phenotypic plasticity in arid and semi-arid environments.

2.1 Greenhouse pot experiments

Artemisia ordosica is a dominant psammophyte species that forms communities on sandy lands in arid and semi-arid areas of Northwest China. Restoration of *A. ordosica*-dominated plant communities is currently crucial for controlling degraded vegetation ecosystems in the study region (Wang et al., 2018).

The pot experiments were conducted in March 2012 at the greenhouse of the Department of Plant Biology and Ecology, Nankai University, China. Seeds of *A. ordosica* and experimental soil were collected from Bayanhot in Alxa Left Banner, Inner Mongolia Autonomous Region, China.

To investigate *A. ordosica* growth in soils with varying CaCO₃ contents, we performed three pot experiments using a two-factor randomized block design and two orthogonal designs. In the low soil CaCO₃ content orthogonal experiment with an L8 (4×24) array, soil CaCO₃ levels were 0 (control), 10, 25, and 50 g/kg. In the high soil CaCO₃ content orthogonal experiment with an L16 (45) array, soil CaCO₃ levels were 0 (control), 25, 50, and 100 g/kg, soil available P levels were 0, 3, 6, and 12 g/kg, and watering amounts were 200, 250, 300, and

350 mL/week. In the two-factor randomized block experiment with high soil CaCO content, soil CaCO levels were 10, 50, 75, and 100 g/kg, and watering levels were 250, 300, 350, and 400 mL/week, respectively. All three experiments included three replicates per treatment.

The seed sowing procedure was as follows: seeds were sown at 3-4 seeds per hole in seedling-raising pans filled with a 1:1 turf:perlite mixture. The pans were covered with a thin moist paper film to reduce evaporation until germination occurred. After 25 days, germinated seedlings were transferred to polyvinyl chloride (PVC) pots (170 mm × 200 mm; one seedling per pot) according to the experimental design. Each pot contained 2 kg of mixed soil at a 2:1 sandy soil:loam ratio, with the mixture containing 3.11 g/kg organic matter, 7.00 mg/kg available N, and 5.77 mg/kg available P.

To calculate relative growth rate (RGR) of potted seedlings, we measured all initial plant heights upon transfer to PVC pots. Additionally, 50 leftover *A. ordosica* seedlings from the seedling-raising pans served as initial controls. These seedlings were excavated, washed to remove root mud, blotted with filter paper, and measured for plant height, root length, and fresh weights of shoot-leaf and root biomass.

The *A. ordosica* seedlings were then grown for a 40-day monitoring period. To minimize water evaporation, we watered all seedlings by injecting water beneath the soil surface to a depth of 2 cm using a syringe. Greenhouse temperatures were maintained at 25°C during daytime and 18°C at night. Upon harvest, we measured and recorded plant height, root length, shoot-leaf biomass, and root biomass for every seedling.

2.2 Sample analyses

For seedlings in the high soil CaCO content experiment, plant N and P concentrations were determined using H SO -H O -semi-micro-Kjeldahl and H SO -H O -Mo-Sb-Vc-colorimetry methods, respectively (Bao, 2007). Each plant root sample was analyzed with three measurement replicates.

2.3 Calculation of RGR

We used 50 leftover *A. ordosica* seedlings for curve regressions with plant height as the independent variable and shoot-leaf biomass, root length, and root biomass as separate dependent variables. Because all regression equations were significant ($P < 0.001$), they were used to estimate pre-experiment shoot-leaf biomass, root length, and root biomass of the potted seedlings.

The RGR values for plant height, root length, shoot-leaf biomass, and root biomass of *A. ordosica* seedlings were calculated as follows:

$$\mu = \frac{\ln(M_t/M_0)}{t} \quad (1)$$

where r is the relative growth rate of plant height (cm/cm/d), root length (cm/cm/d), shoot-leaf biomass (g/g/d), or root biomass (g/g/d); M_0 is the observed pre-experiment plant height (cm) or shoot biomass (g), root length (cm), or root biomass (g) predicted from the regression equations; M_t is the post-experiment measurement of the four growth response variables; and t is the growing time (40 d).

2.4 Data analysis

Data were analyzed using SPSS 13.0 software (SPSS, Chicago, Illinois, USA) with two-way ANOVA. Homogeneity tests were performed first, followed by Duncan's test for comparing means between groups with equal variances. Tamhane's T2 test was used for groups with unequal variances.

3.1 Effects of low soil CaCO₃ content on *A. ordosica* seedling growth

In the low soil CaCO₃ content orthogonal experiment, *A. ordosica* seedlings treated with 50 g/kg soil CaCO₃ showed reductions of 69.29%, 46.18%, 87.88%, and 66.75% in plant height, root length, shoot-leaf biomass, and root biomass, respectively, compared to the control. As shown in Figure 1 [Figure 1: see original paper], the RGR values for all four growth response variables were lowest under the highest soil CaCO₃ content and highest under the lowest content.

3.2 Effects of high soil CaCO₃ content on *A. ordosica* seedling growth

In the high soil CaCO₃ content two-factor randomized block experiment, RGR values for plant height and shoot-leaf biomass decreased significantly with increasing soil CaCO₃ content, while RGR values for root length and root biomass also declined but not significantly (Fig. 2a [Figure 2: see original paper]). In this experiment, watering level treatments showed no significant effects on any growth response variable ($P > 0.05$) (data not shown).

In the high soil CaCO₃ content orthogonal experiment, soil available P and watering amount treatments had no significant effects on any growth response variable ($P > 0.05$) (data not shown). Soil CaCO₃ content did not significantly influence RGR values for plant height, shoot-leaf biomass, or root biomass, but it did significantly and negatively impact root length RGR (Fig. 2b).

3.3 Relationships of RGR with root N/P ratio and root P concentration in *A. ordosica* seedlings

Regression analyses revealed that under high soil CaCO₃ content, *A. ordosica* seedlings with higher root N/P ratios exhibited lower RGR values for plant height (Fig. 3a [Figure 3: see original paper]) and shoot-leaf biomass (Fig. 3b)

in the two-factor randomized block experiment, and lower root length RGR in the orthogonal experiment (Fig. 3c). A significant positive logarithmic relationship was observed between root length RGR and root P concentration in the orthogonal experiment (Fig. 3d). In other words, root length RGR in *A. ordosica* seedlings decreased significantly with increasing root N/P ratio but increased significantly with rising root P concentration.

4 Discussion

Soil CaCO₃ can exert complex influences on plant phenotypic plasticity. In this study, CaCO₃ additions significantly affected RGR values for plant height, shoot-leaf biomass, and root length in *A. ordosica* seedlings, as did soil available P concentrations under high CaCO₃ conditions. Higher soil CaCO₃ contents reduced growth in plant height, root length, shoot-leaf biomass, and root biomass of *A. ordosica* seedlings, demonstrating that soil CaCO₃ can affect phenotypic plasticity. This is an intriguing phenomenon, as CaCO₃ molecules are neutral. How might they influence seedling growth rates? One plausible explanation is that CaCO₃ made the soil “tighter,” increasing water retention and hindering seedling water uptake, thereby adversely affecting growth. Another mechanistic explanation is that added CaCO₃ strongly reduced soil P availability, further limiting growth of water-stressed *A. ordosica* seedlings. Regardless of the mechanism, soil CaCO₃ additions reduced the growth potential of *A. ordosica* seedlings.

Notably, root length growth in *A. ordosica* seedlings gradually declined under higher soil CaCO₃ contents. Root length was constrained rather than promoted by high soil CaCO₃, likely because the anterior root extremities decayed during early seedling growth. When *A. ordosica* seedlings were transferred from seedling-raising pans (filled with 1:1 turf:perlite) to PVC pots (filled with sandy loam containing CaCO₃), some decayed roots probably died, with higher soil CaCO₃ content causing more root death. Thus, CaCO₃ additions prevented root length expansion in *A. ordosica* seedlings.

Kerley (2000) observed growth plasticity in *Lupinus albus* when exposed to limed soil profiles. In our study, increased soil CaCO₃ contents progressively decreased plant height, root length, shoot-leaf biomass, and root biomass in *A. ordosica* seedlings. During the 40-day growth period, seedlings treated with 50 g/kg soil CaCO₃ showed reductions of 69.29%, 46.18%, 87.88%, and 66.75% in plant height, root length, shoot-leaf biomass, and root biomass, respectively, compared to controls, with corresponding RGR reductions of 89.99%, 145.41%, 111.73%, and 78.90%. Clearly, soil CaCO₃ can inhibit growth plasticity in *A. ordosica* seedlings. While soil CaCO₃ is a soil formation matrix, its influence on plant growth differs from other ecological factors such as light, temperature, water, heat, wind, soil nutrients, and salinity (Fan et al., 2012; Zheng et al., 2012). Our results suggest that soil CaCO₃ represents an abiotic stress factor for *A. ordosica* in arid and semi-arid Northwest China. As Ghalambor et al. (2007) noted, adaptive phenotypic plasticity favors plant establishment in new habitats,

while non-adaptive plasticity corresponds with stressful environments. In this context, soil CaCO₃ appears to generate non-adaptive phenotypic plasticity.

The inhibition of growth plasticity in *A. ordosica* seedlings by soil CaCO₃ was essentially due to restricted P absorption. In calcareous soils of arid and semi-arid regions, approximately 70% of total inorganic P exists as Ca-P complexes (Ma et al., 2009). Among soil inorganic P pools, only Ca-P was used here to simplify the more complex 0.5 M NaHCO₃ (pH 7.5)-soluble P form, which significantly affects soil available P levels (Ma et al., 2009). Soil CaCO₃ accumulation maintains soil pH at 7.0–8.0, reducing P availability and restricting plant P utilization rates (Reinbott and Blevins, 1999; Li et al., 2004; Wojewodzic et al., 2011). P restriction inhibits plant growth primarily by altering plant C/N/P stoichiometry (Ehlers et al., 2010; Persson et al., 2011; Cai et al., 2012). In arid and semi-arid regions where P is crucial for plant growth (Pajaei et al., 2018), altered P concentrations can change plant N/P ratios (He et al., 2016; Martinez-Oro et al., 2017). The growth rate hypothesis proposed by Elser et al. (2010) states that plant specific growth rate is negatively related to N/P ratio (Matzek and Vitousek, 2009). Our results showing decreased RGR values for growth response variables (plant height, root-leaf biomass, and root length) with higher N/P ratios in *A. ordosica* seedlings are consistent with this hypothesis.

Plant roots are particularly sensitive to environmental change (Qiu et al., 2017). In this study, root length in *A. ordosica* seedlings showed a negative response to higher soil CaCO₃ contents. Our follow-up analysis suggested that root length growth increased with higher root P concentrations, implying that the influence of root N/P ratio on root length RGR occurred via CaCO₃ effects on P absorption. Thus, higher soil CaCO₃ contents resulted in lower P concentrations in *A. ordosica* seedlings, consequently affecting their phenotypic plasticity.

5 Conclusions

Soil CaCO₃ is clearly a stress factor that adversely affects *A. ordosica* seedling growth. Regardless of whether CaCO₃ contents are lower (<50 g/kg) or higher (>50 g/kg), soil CaCO₃ additions slow *A. ordosica* seedling growth. Therefore, soil CaCO₃ induces non-adaptive phenotypic plasticity in *A. ordosica* seedlings. Under high soil CaCO₃ contents, RGR values for plant height, shoot-leaf biomass, and root length all decreased significantly with increasing root N/P ratio, while root length RGR increased significantly with rising root P concentration. Thus, altered root P concentration is the primary driver of phenotypic plasticity in *A. ordosica* seedlings.

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References

- Abbas J A, Mohammed S A, Saleh M A. 1991. Edaphic factors and plant species distribution in a protected area in the desert of Bahrain Island. *Vegetation*, 95(1): 87-93.
- Bao S D. 2007. *Analytical Methods for Soil and Agro-Chemistry* (3rd ed.). Beijing: Chinese Agriculture Science and Technology Press, 268-270, 389-391. (in Chinese)
- Bossdorf O, Pigliucci M. 2009. Plasticity to wind is modular and genetically variable in *Arabidopsis thaliana*. *Evolutionary Ecology*, 23(5): 669-685.
- Bradshaw A D. 1965. Evolutionary significance of phenotypic plasticity in plants. *Advances in Genetics*, 13: 115-155.
- Bradshaw A D, Hardwick K. 1989. Evolution and stress—genotypic and phenotypic components. *Biological Journal of the Linnean Society*, 37(1-2): 137-155.
- Cai H M, Xie W B, Zhu T, et al. 2012. Transcriptome response to phosphorus starvation in rice. *Acta Physiologiae Plantarum*, 34(1): 327-341.
- Chen M M, Yin H B, O' Connor P, et al. 2010. C: N: P stoichiometry and specific growth rate of clover colonized by arbuscular mycorrhizal fungi. *Plant and Soil*, 326(1-2): 21-29.
- D' Ambrosio P, Colagè I. 2017. Extending epigenesis: from phenotypic plasticity to the bio-cultural feedback. *Biology & Philosophy*, 32(5): 705-728.
- Ehlers K, Bakken L R, Frostegård Å, et al. 2010. Phosphorus limitation in a Ferralsol: impact on microbial activity and cell internal P pools. *Soil Biology and Biochemistry*, 42(4): 558-566.
- Elser J J, Fagan W F, Kerkhoff A J, et al. 2010. Biological stoichiometry of plant production: metabolism, scaling and ecological response to global change. *New Phytologist*, 186(3): 593-608.
- Fan Y, Li P F, Hou Z A, et al. 2012. Water adaptive traits of deep-rooted C halophyte (*Karelinia caspica* (Pall.) Less.) and shallow-rooted C halophyte (*Atriplex tatarica* L.) in an arid region, Northwest China. *Journal of Arid Land*, 4(4): 469-478.
- Fusco G, Minelli A. 2010. Phenotypic plasticity in development and evolution: facts and concepts. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365: 547-556.
- Geng Y P, Zhang W J, Li B, et al. 2004. Phenotypic plasticity and invasiveness of alien plants. *Biodiversity Science*, 12(4): 447-455. (in Chinese)
- Ghalambor C K, McKay J K, Carroll S P, et al. 2007. Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Functional Ecology*, 21(3): 394-407.

- He X D, You W X, Yu D. 2016. *Ecological Restoration Theory and Vegetation Reconstruction Technique in Yanchi County of the Ningxia Hui Autonomous Region*. Tianjin: Nankai University Press, 32-58. (in Chinese)
- Jain S K, Bradshaw A D. 1966. Evolutionary divergence among adjacent plant populations. *Heredity*, 21(3): 407-441.
- Kerley S J. 2000. Changes in root morphology of white lupin (*Lupinus albus* L.) and its adaptation to soils with heterogeneous alkaline/acid profiles. *Plant and Soil*, 218(1-2): 197-205.
- Lauri P É, Barigah T S, Lopez G, et al. 2016. Erratum to: Genetic variability and phenotypic plasticity of apple morphological responses to soil water restriction in relation with leaf functions and stem xylem conductivity. *Trees*, 30(5): 1909-1910.
- Lee J A, Woolhouse H W. 1971. The relationship of compartmentation of organic acid metabolism to bicarbonate ion sensitivity of root growth in calcicoles and calcifuges. *New Phytologist*, 70: 103-111.
- Li J M, Du L S, Guan W B, et al. 2016. Latitudinal and longitudinal clines of phenotypic plasticity in the invasive herb *Solidago canadensis* in China. *Oecologia*, 182(3): 755-764.
- Li L, Tang C, Renge Z, et al. 2004. Calcium, magnesium and microelement uptake as affected by phosphorus sources and interspecific root interactions between wheat and chickpea. *Plant and Soil*, 261(1-2): 29-37.
- Ma B, Zhou Z Y, Zhang C P, et al. 2009. Inorganic phosphorus fractions in the rhizosphere of xerophytic shrubs in the Alxa Desert. *Journal of Arid Environments*, 73(1): 55-61.
- Martinez-Oro D, Parraga-Aguado I, Querejeta J I, et al. 2017. Importance of intra- and interspecific plant interactions for the phytomanagement of semiarid mine tailings using the tree species *Pinus halepensis*. *Chemosphere*, 186: 405-413.
- Matzek V, Vitousek P M. 2009. N: P stoichiometry and protein: RNA ratios in vascular plants: an evaluation of the growth-rate hypothesis. *Ecology Letters*, 12(8): 765-771.
- Pedersen J, Fransson A M, Olsson P A. 2011. Performance of *Anisantha* (*Bromus*) *tectorum* and *Rumex acetosella* in sandy calcareous soil. *Flora*, 206(3): 276-281.
- Persson J, Wojewodzic M W, Hessen D O, et al. 2011. Increased risk of phosphorus limitation at higher temperatures for *Daphnia magna*. *Oecologia*, 165: 123-129.
- Pigliucci M, Diiorio P, Schlichting C. 1997. Phenotypic plasticity of growth trajectories in two species of *Lobelia* in response to nutrient availability. *Journal of Ecology*, 85(3): 265-276.

- Pigliucci M. 2002. Touchy and bushy: phenotypic plasticity and integration in response to wind stimulation in *Arabidopsis thaliana*. *International Journal of Plant Sciences*, 163(3): 399–408.
- Pigliucci M, Kolodynska A. 2002. Phenotypic plasticity to light intensity in *Arabidopsis thaliana*: invariance of reaction norms and phenotypic integration. *Evolutionary Ecology*, 16(1): 27–47.
- Pigliucci M. 2005. Evolution of phenotypic plasticity: where are we going now? *Trends in Ecology & Evolution*, 20(9): 481–486.
- Qiu R J, Du T S, Kang S Z. 2017. Root length density distribution and associated soil water dynamics for tomato plants under furrow irrigation in a solar greenhouse. *Journal of Arid Land*, 9(5): 637–650.
- Reinbott T M, Blevins D G. 1999. Phosphorus nutritional effects on root hydraulic conductance, xylem water flow and flux of magnesium and calcium in squash plants. *Plant and Soil*, 209(2): 263–273.
- Rivas-Ubacha A, Sardansa J, Pérez-Trujillo M, et al. 2012. Strong relationship between elemental stoichiometry and metabolome in plants. *Proceedings of the National Academy of Sciences of the United States of America*, 109(11): 4181–4186.
- Schinas S, Rowell D L. 1977. Lime-induced chlorosis. *European Journal of Soil Science*, 28(2): 351–368.
- Schlichting C D. 1986. The evolution of phenotypic plasticity in plants. *Annual Review of Ecology and Systematics*, 17: 667–693.
- Stearns S C. 1989. The evolutionary significance of phenotypic plasticity. *Bio-science*, 39(7): 436–445.
- Storz J F, Scott G R, Cheviron Z A. 2010. Phenotypic plasticity and genetic adaptation to high-altitude hypoxia in vertebrates. *The Journal of Experimental Biology*, 213: 4125–4136.
- Sultan S E. 2001a. Phenotypic plasticity for fitness components in *Polygonum* species of contrasting ecological breadth. *Ecology*, 82(2): 328–343.
- Sultan S E. 2001b. Phenotypic plasticity for plant development, function and life history. *Trends in Plant Science*, 5(12): 537–542.
- Via S, Gomulkiewicz R, De Jong G, et al. 1995. Adaptive phenotypic plasticity: consensus and controversy. *Trends in Ecology & Evolution*, 10(5): 212–217.
- Wang X, Song N P, Yang X G, et al. 2018. Grazing exclusion-induced shifts, the relative importance of environmental filtering, biotic interactions and dispersal limitation in shaping desert steppe communities, northern China. *Journal of Arid Land*, 10(3): 423–436.
- Wojewodzic M W, Kyle M, Elser J J, et al. 2011. Joint effect of phosphorus limitation and temperature on alkaline phosphatase activity and somatic growth

in *Daphnia magna*. *Oecologia*, 165(4): 837–846.

Zhao X L, He X D, Xue P P, et al. 2012. Effects of soil stoichiometry of the CaCO₃/available phosphorus ratio on plant density in *Artemisia ordosica* communities. *Chinese Science Bulletin*, 57(5): 492–499.

Zheng M, Lai L, Jiang L, et al. 2012. Moderate water supply and partial sand burial increase relative growth rate of two *Artemisia* species in an inland sandy land. *Journal of Arid Environments*, 85: 105–113.

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