

Allelopathic Effects of *Artemisia halodendron* Leaf Aqueous Extract on Seed Germination of Four Sandy Land Herbaceous Plants: Postprint

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Abstract

Artemisia halodendron is an important species in vegetation succession during sand dune restoration and one of the dominant species in semi-mobile dunes. This study investigated, from an allelopathic perspective, the effects of different concentrations of *Artemisia halodendron* leaf extract (0, 10, 20, 40, 60, 80 g · L⁻¹ and 100 g · L⁻¹) on seed germination of *Setaria viridis*, *Lespedeza davurica*, *Corispermum macrocarpum*, and *Agropyron cristatum*. The results demonstrated that *Artemisia halodendron* leaf extract had minimal effects on seed germination of *Setaria viridis*, with low-concentration treatments exhibiting a certain stimulatory effect on its germination, whereas significant inhibitory effects were observed on the other three species, and the inhibitory effect intensified with increasing treatment concentration. *Artemisia halodendron* exhibits strong allelopathic effects on seed germination of other species, and allelopathy resulting from the release of substances following leaf fall may represent one of the important mechanisms for maintaining its competitive advantage.

Full Text

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Abstract

Artemisia halodendron is one of the dominant species in vegetation succession during sandland restoration. At the initial stage of vegetation regeneration over sandland, *A. halodendron* dominates semi-fixed dunes, but is subsequently replaced by other annual or perennial species such as *Lespedeza bicolor*, *Corispermum macrocarpum*, and *Agropyron cristatum* as restoration progresses. However, the mechanism underlying this succession remains unclear. This study investigated the allelopathic effects of *A. halodendron* leaf water extracts at different concentrations (0, 10, 20, 40, 60, 80, and 100 g · L⁻¹) on seed germination of one symbiotic species, *Setaria viridis*, and three competitive species: *L. bicolor*, *C. macrocarpum*, and *A. cristatum*. Results showed that *A. halodendron* leaf extracts had minimal effects on *S. viridis* germination, with slight promotion observed at low concentrations. However, germination of the three target species was significantly inhibited by the extracts, and this allelopathic inhibition intensified with increasing concentration. These findings demonstrate that *A. halodendron* exerts strong allelopathic effects on other species during vegetation regeneration in sandland restoration. The allelopathic substances released from decomposing leaves may represent a key competitive strategy maintaining its dominance during early successional stages.

Keywords: *Artemisia halodendron*; allelopathic effect; seed germination; allelopathic index; vegetation succession; Tongliao

1 Introduction

Artemisia halodendron plays a crucial role in vegetation succession during sandland restoration. As a pioneer species, it initially colonizes semi-fixed dunes and dominates the early stages of vegetation regeneration. However, as restoration proceeds, it is gradually replaced by other annual and perennial species including *Lespedeza bicolor*, *Corispermum macrocarpum*, and *Agropyron cristatum*. The ecological mechanisms driving this species replacement remain poorly understood. Allelopathy, the chemical inhibition of one plant by another through

the release of secondary metabolites into the environment, may contribute significantly to this successional pattern. Previous studies have documented allelopathic effects in various plant species, including *Melilotus officinalis*, *Sorghum sudanense*, *Lolium multiflorum*, *Stellera chamaejasme*, *Onobrychis viciifolia*, *Gentiana straminea*, *Medicago sativa*, and *Trifolium* species. However, the allelopathic potential of *A. halodendron* and its role in sandland vegetation dynamics requires further investigation.

2 Materials and Methods

2.1 Study Site and Climate Conditions

The study was conducted at the Horqin Sandy Land Experimental Station (42°58 N, 120°43 E) at an elevation of 345 m. The region experiences a semi-arid continental climate with an average annual precipitation of 343 mm (1961–2017), with approximately 80% occurring during June–September. The mean annual temperature is 6.7°C. Soils are classified as sandy chestnut soil with low organic matter content.

2.2 Plant Material and Extract Preparation

Artemisia halodendron leaves were collected from the experimental site in April 2017. Fresh leaves were washed, air-dried, and ground into powder. Leaf water extracts were prepared by soaking 100 g of powder in 1000 mL distilled water at 25°C for 24 h, then filtered through filter paper. The stock solution (100 g · L⁻¹) was diluted to obtain concentrations of 0, 10, 20, 40, 60, 80, and 100 g · L⁻¹. The control treatment (0 g · L⁻¹) used distilled water only.

2.3 Germination Bioassay

Seeds of *Setaria viridis*, *Lespedeza bicolor*, *Corispermum macrocarpum*, and *Agropyron cristatum* were collected from the same region in September 2016. Prior to germination, seeds were surface-sterilized with 0.5% sodium hypochlorite for 10 minutes, then rinsed thoroughly with distilled water. For each treatment, 50 seeds were placed in sterile Petri dishes lined with two layers of filter paper moistened with 5 mL of the respective extract solution. Dishes were sealed with parafilm and incubated in a growth chamber at 25°C with a 16 h light/8 h dark photoperiod (light intensity 9000 lx). Germination was recorded daily for 21 days, with seeds considered germinated when radicles emerged 2 mm. Each treatment was replicated four times.

2.4 Measurement Indices

Germination Rate (GR) was calculated as:

$$\text{GR} = (\text{Number of germinated seeds} / \text{Total seeds}) \times 100\%$$

Germination Index (GI) was calculated following Williamson and Richardson (1988):

$$GI = \sum(G / D)$$

where G is the number of germinated seeds on day t , and D is the corresponding germination day.

Allelopathic Index (AI) was calculated as:

$$AI = (\text{Treated GR} - \text{Control GR}) / \text{Control GR}$$

Data were analyzed using SPSS 13.0 software. One-way ANOVA was performed to test for significant differences among treatments, followed by LSD post-hoc tests at $P < 0.05$ significance level.

3 Results

3.1 Effects on Germination Rate

The leaf water extracts of *Artemisia halodendron* had concentration-dependent effects on seed germination rates (Table 1). Germination rates across all species ranged from 44.7% to 58.7% under different extract concentrations, with no significant differences among treatments for individual species ($F = 1.86$, $P > 0.05$). However, significant differences were observed in germination indices.

3.2 Effects on Germination Index

The germination index varied significantly among extract concentrations ($F = 6.33$, $P < 0.01$). The lowest germination index ($17.1 \pm 3.1\%$) occurred at $60 \text{ g} \cdot \text{L}^{-1}$ concentration, which was significantly lower than the control ($12.9 \pm 3.2\%$). These results indicate that while germination rates were only moderately affected, the speed and synchrony of germination were substantially inhibited by higher extract concentrations.

4 Discussion

4.1 Allelopathic Mechanisms

Our results demonstrate that *Artemisia halodendron* leaf extracts exert significant allelopathic effects on co-occurring species during sandland vegetation succession. The inhibition of germination index, rather than final germination percentage, suggests that allelochemicals primarily affect germination speed and seedling vigor. This finding aligns with previous studies on allelopathic effects in arid ecosystems, where water-soluble compounds from decomposing litter influence seed bank dynamics and community composition.

The concentration-dependent response observed in this study is consistent with the hormesis hypothesis, where low concentrations of allelochemicals may stimulate germination while higher concentrations cause inhibition. This pattern has been documented in other *Artemisia* species and may reflect evolutionary adaptations to resource competition in nutrient-poor sandy soils.

4.2 Ecological Implications

The strong allelopathic effect of *A. halodendron* on *Lespedeza bicolor*, *Corispermum macrocarpum*, and *Agropyron cristatum* may explain its initial dominance in early successional stages. By suppressing the germination and establishment of potential competitors, *A. halodendron* can maintain its population advantage during critical establishment phases. However, as litter accumulates and allelochemicals leach into the soil, the selective pressure may shift, allowing more tolerant species to establish and eventually replace *A. halodendron*.

This allelopathic-mediated succession represents an important mechanism in sandland restoration. Understanding these chemical interactions can inform management practices, such as optimizing planting densities and timing to facilitate natural succession processes. Future research should identify specific allelochemical compounds and their persistence in sandy soils to better predict long-term community dynamics.

5 Conclusion

Artemisia halodendron exhibits significant allelopathic effects on symbiotic and competitive species in sandland ecosystems. The leaf water extracts inhibit germination index in a concentration-dependent manner, with stronger inhibition observed at concentrations $40 \text{ g} \cdot \text{L}^{-1}$. These allelopathic interactions likely contribute to species replacement patterns during vegetation succession in restored sandlands. The release of allelochemicals from decomposing leaves represents a key competitive strategy that influences community assembly and restoration trajectories.

References

- [1] JABRAN K, MAHAJAN G, SARDANA V, et al. Allelopathy for weed control in agricultural systems[J]. *Crop Protection*, 2015, 72: 57-65.
- [2] RAWAT L S, MAIKHURI R K, BAHUGUNA Y M, et al. Sunflower allelopathy for weed control in agriculture systems[J]. *Journal of Crop Science and Biotechnology*, 2017, 20(1): 45-60.
- [3-4] WANG Jing-yi, YAO Dan-dan, ZHAO Guo-qi, et al. Allelopathic effects of aqueous extract of *Melilotus officinalis* on *Lolium multiflorum* and *Sorghum sudanense*[J]. *Acta Prataculturae Sinica*, 2017, 26(8): 85-92.
- [5] ZHOU Shu-qing, HUANG Zu-jie, WANG Hui, et al. Allelopathic effect of *Stellera chamaejasme* decomposing in soil on *Onobrychis viciifolia*[J]. *Pratacultural Science*, 2009, 26(3): 91-94.
- [6-7] DING Chun-fa, WEI Xiao-hong, WANG Fang-lin. Effects of active allelochemicals from *Gentiana straminea* on seed germination and seedling physiolog-

- ical properties of forages[J]. *Acta Prataculturae Sinica*, 2017, 26(4): 150-161.
- [8-9] YE Xiao-qi, WU Ming, SHAO Xue-xin, et al. Effects of water extracts from *Solidago canadensis* on the growth of maize seedlings and underlying photosynthetic mechanisms[J]. *Acta Prataculturae Sinica*, 2014, 23(6): 217-224.
- [10] ZUO Xiao-an, ZHAO Halin, ZHAO Xue-yong, et al. Species diversity of degraded vegetation in different age restorations in Horqin Sandy Land, Northern China[J]. *Acta Prataculturae Sinica*, 2009, 18(4): 9-16.
- [11] ZHANG Jing, ZUO Xiao-an, LÜ Peng, et al. Plant functional traits and interrelationships of dominant species on typical grassland in Horqin Sandy Land, China[J]. *Arid Zone Research*, 2018, 35(1): 137-143.
- [12] ZHAO Halin, LI Jin, ZHOU Rui-lian, et al. Effect of wind blowing time and intensity on membrane permeability and protection system of corn seedling[J]. *Arid Zone Research*, 2016, 33(3): 519-524.
- [13] ABHILASHA D, QUINTANA N, VIVANCO J, et al. Do allelopathic compounds in invasive *Solidago canadensis* s.l. restrain the native European flora?[J]. *Journal of Ecology*, 2008, 96(5): 993-1001.
- [14] ZHANG Q, YAO L J, YANG R Y, et al. Potential allelopathic effects of an invasive species *Solidago canadensis* on the mycorrhizae of native plant species[J]. *Allelopathy Journal*, 2007, 20(1): 71-77.
- [15] ZHAO Halin, ZHAO Xue-yong, ZHANG Tong-hui, et al. *Desertification Processes and Its Restoration Mechanisms in the Horqin Sandy Land*[M]. Beijing: China Ocean Press, 2003.
- [16] WANG Tao, WU Wei, ZHAO Halin, et al. Analyses on driving factors to sandy desertification process in Horqin Region, China[J]. *Journal of Desert Research*, 2004, 24(5): 519-528.
- [17] WANG Jun, WANG Zhuo-han, YANG Long, et al. Effects of litter coverage and watering frequency on seed germination and seedling survival of *Castanopsis fissa*[J]. *Chinese Journal of Applied Ecology*, 2008, 19(10): 2097-2102.
- [18] LUO Yong-qing, ZHAO Xue-yong, ZHU Yang-chun, et al. Germination and seeding growth of *Artemisia halodendron* under different incubation environment[J]. *Chinese Journal of Applied Ecology*, 2014, 25(1): 31-36.
- [19] LUO Yong-qing, ZHAO Xue-yong, ZHOU Xin, et al. Relationship between growth character and belowground biomass distribution of *Artemisia halodendron*[J]. *Journal of Desert Research*, 2015, 35(1): 152-159.
- [20] QU H, ZHAO X, ZHAO H, et al. Litter decomposition rates in Horqin Sandy Land, Northern China: Effects of habitat and litter quality[J]. *Fresenius Environmental Bulletin*, 2012, 20(12): 3304-3312.
- [21] ZUO X, KNOP J M H, ZHAO X, et al. Indirect drivers of plant diversity-productivity relationship in semiarid sandy grasslands[J]. *Biogeosciences*, 2012,

9(4): 1277-1289.

[22] ZUO X, ZHAO X, WANG S, et al. Influence of dune stabilization on relationship between plant diversity and productivity in Horqin Sandy Land, Northern China[J]. *Environmental Earth Sciences*, 2012, 67(5): 1547-1556.

[23] ZHAO X, ZHAO H, ZUO X, et al. Effects of water extracts from *Solidago canadensis* on the growth of maize seedlings and underlying photosynthetic mechanisms[J]. *Acta Prataculturae Sinica*, 2014, 23(6): 217-224.

[24] ZHANG Jin, ZUO Xiao-an, LÜ Peng, et al. Plant functional traits and interrelationships of dominant species on typical grassland in Horqin Sandy Land, China[J]. *Arid Zone Research*, 2018, 35(1): 137-143.

[25] ZHAO Halin, LI Jin, ZHOU Rui-lian, et al. Effect of wind blowing time and intensity on membrane permeability and protection system of corn seedling[J]. *Arid Zone Research*, 2016, 33(3): 519-524.

[26] WILLIAMSON G B, RICHARDSON D. Bioassays for allelopathy: Measuring treatment responses with independent controls[J]. *Journal of Chemical Ecology*, 1988, 14: 181-187.

[27] HU Rong, LIN Bo, LIU Qing, et al. Effects of forest gaps and litter on the early regeneration of *Picea asperata* plantations[J]. *Scientia Silvae Sinicae*, 2011, 47(6): 23-29.

[28] WANG Jun, WANG Zhuo-han, YANG Long, et al. Effects of litter coverage and watering frequency on seed germination and seedling survival of *Castanopsis fissa*[J]. *Chinese Journal of Applied Ecology*, 2008, 19(10): 2097-2102.

[29] SHI Yan, CHEN Fang-qing. Effects of *Larix kaempferi* litter on seed germination and seedling growth of *Cynodon dactylon* and *Lespedeza formosa*[J]. *Jiangsu Agricultural Sciences*, 2018, 46(6): 267-270.

[30] ZHOU Yan, CHEN Xu, WEI Xiao-li, et al. Effects of litter on the seedling regeneration and seed germination of *Rhododendron agastum*[J]. *Scientia Silvae Sinicae*, 2015, 51(3): 65-74.

[31] HAN C M, PAN K W, WU N, et al. Allelopathic effect of ginger on seed germination and seedling growth of soybean and chive[J]. *Scientia Horticulturae*, 2008, 116(3): 330-336.

[32] JEFFERSON L V, PENNACCCHIO M. Allelopathic effects of foliage extracts from four *Chenopodiaceae* species on germination[J]. *Journal of Arid Environments*, 2003, 55(2): 275-285.

[33] ZHANG Y, TANG S, LIU K, et al. The allelopathic effect of *Potentilla acaulis* on the changes of plant community in grassland, northern China[J]. *Ecological Research*, 2015, 30(1): 41-47.

[34] LUO Y, ZHAO X, LI Y, et al. Effects of foliage litter of a pioneer shrub (*Artemisia halodendron*) on germination from the soil seedbank in a semi-arid

- sandy grassland in China[J]. Journal of Plant Research, 2017, 130(3): 1013-1021.
- [35] BRUCE W G, RICHARDSON D. Bioassays for allelopathy: Measuring treatment responses with independent controls[J]. Journal of Chemical Ecology, 1988, 14(1): 181-187.
- [36] RABOTNOV T A. Importance of the evolutionary approach to the study of allelopathy[J]. Soviet Journal of Ecology, 1982, 3: 5-8.
- [37] PUTNAM A R, TANG C S. The Science of Allelopathy[M]. New York: Wiley, 1986.
- [38] PRATI D, BOSSDORF O. Allelopathic inhibition of germination by *Alliaria petiolata* (Brassicaceae)[J]. American Journal of Botany, 2004, 91(2): 285-288.
- [39] TURK M A, TAWAHA A M. Allelopathic effect of black mustard (*Brassica nigra* L.) on germination and growth of wild oat (*Avena fatua* L.)[J]. Crop Protection, 2003, 22(4): 673-677.
- [40] RODRÍGUEZ A A, GRUNBERG K A, TALEISNIK E L. Reactive oxygen species in the elongation zone of maize leaves are necessary for leaf extension[J]. Plant Physiology, 2002, 129(4): 1627-1632.
- [41] PRITHIVIRAJ B, PERRY L G, BADRI D V, et al. Chemical facilitation and induced pathogen resistance mediated by a root-secreted phytotoxin[J]. New Phytologist, 2007, 173(4): 852-860.
- [42] ZHANG S S, WANG B, ZHANG L, et al. Hormetic-like dose response relationships of allelochemicals of invasive *Solidago canadensis* L.[J]. Allelopathy Journal, 2012, 29(1): 151-160.
- [43] ZHONG Yu, ZHANG Jian, YANG Wan-qin, et al. Allelopathic effects of *Eucalyptus grandis* on *Medicago sativa* growing under different soil water conditions[J]. Acta Prataculturae Sinica, 2009, 18(4): 81-86.
- [44] ZHANG Jing, ZUO Xiao-an, LÜ Peng, et al. Plant functional traits and interrelationships of dominant species on typical grassland in Horqin Sandy Land, China[J]. Arid Zone Research, 2018, 35(1): 137-143.
- [45] HAN Juan-juan, LI Yu-qiang, WANG Shao-kun, et al. Characteristics of soil organic carbon and total nitrogen under different land use types in Naiman banner[J]. Journal of Arid Land Resources and Environment, 2014, 28(1): 37-42.
- [46] LI Yu-qiang, ZHAO Halin, ZHAO Xue-yong, et al. Characteristics of soil carbon and nitrogen during desertification process in Horqin Sandy Land[J]. Journal of Soil and Water Conservation, 2005, 19(5): 73-76.

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