

Effects of AM Fungi on Photosynthetic and Chlorophyll Fluorescence Characteristics of *Elaeagnus angustifolia* Seedlings under Salt Stress (Postprint)

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Abstract

To investigate the effects of arbuscular mycorrhizal (AM) fungi on the photosynthetic and physiological characteristics of seedlings under salt stress, a pot experiment was conducted using *Elaeagnus angustifolia* seedlings inoculated with the AM fungus *Glomus intraradices* (GI) and non-inoculated controls (CK), which were subjected to NaCl treatments at concentrations of 0, 100, 200, and 300 mmol/L. Indices including net photosynthetic rate (P_n), gas exchange parameters (transpiration rate T_r , stomatal conductance G_s , intercellular CO_2 concentration C_i), pigment contents (chlorophyll a, b, total chlorophyll, carotenoids), and chlorophyll fluorescence parameters (maximum photochemical efficiency F_v/F_m , photosystem II efficiency Φ_{PSII} , photochemical quenching coefficient q_P , non-photochemical quenching coefficient NPQ, apparent electron transport rate ETR, potential activity of PSII reaction centers F_v/F_o , heat dissipation rate HDR) were measured in leaves of *E. angustifolia* seedlings under different treatments. The results demonstrated that: (1) With increasing salt concentration, the changing trends of P_n , T_r , G_s , and C_i in leaves of *E. angustifolia* seedlings under GI and CK treatments were fundamentally consistent, all decreasing significantly. However, at identical salt concentrations, these indices in GI-inoculated leaves were significantly higher than those in the CK treatment group ($P < 0.05$). Moreover, compared with the salt-free control treatment, the magnitude of change in these parameters was significantly lower in the GI group than in the CK group. (2) For pigment content parameters in leaves of *E. angustifolia* seedlings, the changing trends with increasing salt concentration were fundamentally consistent between GI-inoculated and CK groups, with all parameters either decreasing or increasing. However, compared with the salt-free control treatment, the magnitude of change in the CK treatment group was significantly greater than that in the GI treatment. (3) With increasing salt

concentration across treatments, F_v/F_m , $\Phi PSII$, qP , ETR , and F_v/F_o in the GI-inoculated treatment exhibited a trend of initial increase followed by subsequent decrease, while NPQ and HDR showed a trend of initial decrease followed by subsequent increase. In contrast, the CK treatment group exhibited a significant decreasing trend for these values, whereas NPQ and HDR displayed trends of initial decrease followed by increase as well as gradual increase. Compared with the salt-free control treatment, the magnitude of change in the GI treatment group was significantly lower than that in the CK group. These findings further reveal that AM fungi play an important role in saline habitats by enhancing plant photosynthetic and chlorophyll fluorescence characteristics, and that salt stress intensity is also a factor influencing this functional role of AM fungi. The symbiosis between halophytes and AM fungi holds promising application prospects for saline-alkali soil remediation.

Full Text

Preamble

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Effects of Arbuscular Mycorrhizal Fungi on Photosynthetic and Chlorophyll Fluorescence Characteristics in *Elaeagnus angustifolia* Seedlings under Salt Stress

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Abstract

To elucidate the effects of arbuscular mycorrhizal (AM) fungi on the photosynthetic physiology of seedlings under salt stress, a pot experiment was conducted with *Elaeagnus angustifolia* seedlings inoculated with *Glomus intraradices* (GI) and non-inoculated controls (CK) subjected to NaCl concentrations of 0, 100, 200, and 300 mmol/L. Gas exchange parameters (net photosynthetic rate P_n , transpiration rate Tr , stomatal conductance G_s , intercellular CO₂ concentration C_i), chlorophyll fluorescence parameters (maximum fluorescence efficiency F_v/F_m , actual PSII efficiency $\Phi PSII$, photochemical quenching coefficient qP , non-photochemical quenching coefficient NPQ , apparent electron transfer rate ETR , PSII reaction center potential activity F_v/F_o , heat dissipation rate HDR), and pigment contents (chlorophyll a, chlorophyll b, carotenoids) were measured in leaf tissues.

The primary findings were threefold. First, P_n , Tr , G_s , and C_i decreased significantly with increasing salt concentration in both CK and GI treatments. However, at the same salt concentration, these parameters were significantly higher in GI-treated seedlings compared to CK ($P < 0.05$), and the magnitude of decline was smaller in GI treatments. Second, chlorophyll and carotenoid

contents showed consistent decreasing trends with increasing salt stress in both treatments, but the reduction amplitude was significantly less pronounced in GI treatments. Third, GI-treated seedlings exhibited an initial increase followed by decrease in F_v/F_m , $\Phi PSII$, qP , ETR , and F_v/F_o with rising salt levels, whereas CK treatments showed consistent declines. NPQ and HDR displayed initial decreases followed by increases in GI treatments, while CK treatments showed gradual increases in NPQ and continuous decreases in HDR. Across all salt concentrations, the magnitude of change in these parameters was significantly lower in GI treatments compared to CK.

These results demonstrate that AM fungi play an important role in improving photosynthetic and chlorophyll fluorescence characteristics under saline conditions, thereby promoting plant growth in salt-affected habitats. The study further reveals that salt stress intensity is a key factor influencing this symbiotic effect. Inoculation of halophytes with mycorrhizal symbionts shows potential application for saline-alkali land improvement.

Keywords: Arbuscular mycorrhizal fungi; salt stress; *Elaeagnus angustifolia*; chlorophyll content; chlorophyll fluorescence parameters

1. Materials and Methods

1.1 Experimental Materials

Elaeagnus angustifolia seeds were provided by Heilongjiang Jinxiu Dadi Bioengineering Co., Ltd. The AM fungus *Glomus intraradices* was propagated by the Restoration Ecology Laboratory of Heilongjiang University. The inoculum consisted of a rhizosphere mixture containing spores, mycelium, and mycorrhizal root fragments. The cultivation substrate was a mixture of forest soil, peat soil, and vermiculite (V:V:V = 6:2:2) sterilized at 120°C for 2 hours.

1.2 Experimental Design and Sampling

The experiment employed a two-factor design: AM fungal inoculation (GI vs. CK) and salt stress (0, 100, 200, and 300 mmol/L NaCl). The inoculation rate was 1% (g/g). For CK treatments, sterilized inoculum was applied. Surface-sterilized seeds were sown in pots (30 cm × 15 cm × 15 cm) containing the substrate. All pots were placed in a plastic greenhouse at Heilongjiang Botanical Garden for cultivation. Salt treatments were applied using distilled water solutions at the specified concentrations.

For physiological measurements, 10 uniform seedlings per treatment were randomly selected, and the 3rd to 5th functional leaves from the top were sampled between 9:00–11:00. For pigment analysis, leaf samples were immediately frozen in liquid nitrogen and stored at -80°C.

1.3 Measurement Methods

Mycorrhizal colonization was determined using the acid fuchsin staining method [19]. Gas exchange parameters (P_n , Tr , G_s , C_i) were measured using a Li-6400 portable photosynthesis system. Chlorophyll fluorescence was measured with a Li-6400-40 fluorometer after 30 minutes of dark adaptation, followed by light adaptation under $500 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ irradiance to determine F_v/F_m , Φ_{PSII} , qP , NPQ , ETR , F_v/F_o , and HDR . Pigment contents were determined by ethanol extraction [20]. All measurements were performed in triplicate.

1.4 Data Analysis

Data were processed using Microsoft Excel 2003 and OriginPro 8.5. Two-way ANOVA and S-N-K tests were used to analyze significant differences among treatments. Change amplitude (A) was calculated following Wang et al. [21] as:

$$A = (P - P_0)/P_0$$

where P represents the parameter value under salt stress (100, 200, or 300 mmol/L) and P_0 represents the control value (0 mmol/L). Positive A values indicate increases relative to control, while negative values indicate decreases. The absolute value of A reflects the magnitude of change.

2. Results

2.1 Mycorrhizal Colonization of *E. angustifolia* Roots

Acid fuchsin staining revealed extensive mycorrhizal structures in GI-treated roots, including abundant hyphae and vesicles with strong colonization intensity. Mycorrhizal infection rates exceeded a certain threshold across all salt concentrations, while no colonization was observed in CK treatments. These results confirm that *G. intraradices* forms effective symbiosis with *E. angustifolia* roots.

[Figure 1: see original paper] The hyphae and vesicles of AM fungi inside *E. angustifolia* roots

2.2 Effects on Net Photosynthetic Rate and Gas Exchange Parameters

Under salt stress, P_n , Tr , G_s , and C_i in both treatments decreased significantly with increasing NaCl concentration. However, at identical salt concentrations, GI-treated seedlings maintained significantly higher P_n , Tr , G_s , and C_i values compared to CK ($P < 0.05$). The decline patterns of G_s and C_i were consistent across treatments.

[Figure 2: see original paper] Effects of GI and CK treatments on net photosynthetic rate and gas exchange parameters in *E. angustifolia* leaves under salt stress

The amplitude of change analysis revealed that at 100 mmol/L NaCl, GI-treated seedlings showed reductions of 34%, 56%, 55%, and 26% in P_n , Tr , G_s , and C_i

respectively, compared to non-salt controls, whereas CK seedlings exhibited greater reductions of 44%, 63%, 63%, and 34%. At 200 mmol/L, GI reductions were 50%, 71%, 76%, and 44%, while CK reductions reached 73%, 85%, 86%, and 66%. At 300 mmol/L, the differences in change amplitude between treatments were statistically significant ($P < 0.05$), with GI-treated seedlings showing significantly smaller declines.

2.3 Effects on Pigment Content

Pigment contents in both treatments decreased progressively with increasing salt concentration. However, chlorophyll a, chlorophyll b, and carotenoid contents were significantly higher in GI-treated leaves at all salt levels ($P < 0.05$). The chlorophyll a/b ratio in GI treatments remained lower than CK throughout the experiment.

[Figure 3: see original paper] Effects of GI and CK treatments on chlorophyll content in *E. angustifolia* seedlings under salt stress

Change amplitude analysis (Table 2) showed that at 100 mmol/L NaCl, GI-treated seedlings exhibited smaller reductions in chlorophyll a (2%) and total chlorophyll (1%) compared to CK (34% and 26% respectively), with differences reaching significant levels ($P < 0.05$). At 200 and 300 mmol/L, the divergence between treatments became more pronounced, with CK showing 56–61% reductions while GI showed only 9–18% reductions ($P < 0.01$).

2.4 Effects on Chlorophyll Fluorescence Parameters

Chlorophyll fluorescence parameters responded differently to salt stress between treatments. In GI-treated seedlings, F_v/F_m , $\Phi PSII$, qP , ETR, and F_v/F_o initially increased slightly at low salt concentrations, peaking at 100 mmol/L before declining. In contrast, these parameters decreased consistently in CK treatments with increasing salt stress. NPQ and HDR showed initial decreases followed by increases in GI treatments, while HDR decreased gradually in CK treatments.

[Figure 4: see original paper] Effects of GI and CK treatments on chlorophyll fluorescence parameters in *E. angustifolia* leaves under salt stress

Change amplitude analysis (Table 3) demonstrated that GI-treated seedlings maintained significantly smaller changes in fluorescence parameters across all salt concentrations compared to CK ($P < 0.05$). For example, at 200 mmol/L NaCl, the reduction amplitude for $\Phi PSII$ was 28% in GI versus 56% in CK.

2.5 Two-Way ANOVA Analysis

Two-way ANOVA revealed that both salt level and mycorrhizal inoculation significantly affected all measured parameters ($P < 0.001$). Significant interactions between the two factors were detected for P_n , C_i , chlorophyll a, chlorophyll b,

total chlorophyll content, and F_v/F_o ($P < 0.05$), indicating that the effect of AM fungi depended on salt concentration.

Two-way ANOVA for the effects and interactions of different treatments on *E. angustifolia* leaf parameters

3. Discussion

3.1 Photosynthetic Response of Mycorrhizal *E. angustifolia* to Salt Stress

Photosynthesis, the foundation of plant growth, is highly sensitive to environmental changes. Under salt stress, stomatal conductance decreases, limiting CO uptake and water loss. This study showed that P_n , T_r , G_s , and C_i declined significantly with increasing salinity, consistent with previous research [1]. However, GI-treated seedlings maintained significantly higher values and experienced smaller reduction amplitudes, demonstrating that AM fungi can enhance photosynthetic capacity and salt tolerance in *E. angustifolia*.

3.2 Chlorophyll Content Response of Mycorrhizal *E. angustifolia* to Salt Stress

Chlorophyll content reflects photosynthetic capacity and is crucial for light energy absorption, transfer, and conversion. Salt stress typically enhances chlorophyllase activity, accelerating chlorophyll degradation [24–26]. Our results align with these findings, but GI-treated seedlings showed significantly higher pigment contents and smaller reduction amplitudes. This suggests AM fungi can mitigate salt-induced chlorophyll degradation, particularly in young seedlings which are more salt-sensitive.

3.3 Chlorophyll Fluorescence Response of Mycorrhizal *E. angustifolia* to Salt Stress

Chlorophyll fluorescence analysis serves as an effective tool for diagnosing photosynthetic performance and plant stress tolerance [28]. AM fungi can alleviate damage to PSII reaction centers and enhance photochemical and non-photochemical quenching coefficients [32–33]. In this study, GI-treated seedlings showed initial increases in F_v/F_m , Φ_{PSII} , qP , ETR , and F_v/F_o at moderate salt stress (100 mmol/L), suggesting enhanced photosynthetic efficiency. The subsequent decline at higher salinity indicates that the protective effect has limits. Nevertheless, the change amplitudes were consistently smaller in GI treatments, confirming that mycorrhizal symbiosis can protect the photosynthetic apparatus from salt damage.

The dual role of AM fungi—improving both chlorophyll content and fluorescence efficiency—helps maintain photosynthetic electron transport and reduces PSII damage under salt stress [34–35]. This synergistic effect significantly enhances net photosynthetic rate and overall salt tolerance.

4. Conclusion

Through pot experiments investigating the effects of *Glomus intraradices* inoculation on *Elaeagnus angustifolia* seedlings under 0, 100, 200, and 300 mmol/L NaCl stress, this study demonstrated that:

1. AM fungi significantly improved net photosynthetic rate, gas exchange parameters, chlorophyll content, and chlorophyll fluorescence characteristics under salt stress.
2. The magnitude of salt-induced changes was significantly smaller in mycorrhizal seedlings, indicating enhanced salt tolerance.
3. Salt concentration and mycorrhizal inoculation showed significant interactive effects on most photosynthetic parameters.

These findings reveal that AM fungi can alleviate salt stress damage by improving plant photosynthetic performance, suggesting potential applications for saline-alkali land improvement through halophyte-mycorrhizal symbiosis.

References

- [1] 孙玉芳等. 盐碱胁迫下真菌对沙枣苗木生长和生理的影响, 2016, 52(6): 18-27.
- [2] Porcel R, Aroca R, Ruiz-Lozano JM. Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. *Agronomy for Sustainable Development*, 2012, 32(1): 181-200.
- [3] Kapoor R, Evelin H, Mathur P, Giri B. Arbuscular mycorrhiza: approaches for abiotic stress tolerance in crop plants. In: Tuteja N, Gill SS, eds. *Plant Acclimation to Environmental Stress*. New York: Springer, 2013: 359-401.
- [4] Bethke PC, Drew MC. Stomatal and nonstomatal components to inhibition of photosynthesis in leaves of *Capsicum annuum* during progressive exposure to NaCl salinity. *Plant Physiology*, 1992, 99(1): 219-226.
- [5] He Y, Chen Y, Yu CL, Jiang QS, Fu JL, Wang GM, Jiang DA. Photosynthesis and yield traits in different soybean lines in response to salt stress. *Photosynthetica*, 2016, 54(4): 630-635.
- [6] Mao PL, Zhang YJ, Cao BH, Guo LM, Shao HB, Cao ZY, Jiang QK, Wang X. Effects of salt stress on eco-physiological characteristics in *Robinia pseudoacacia* based on salt-soil rhizosphere. *Science of the Total Environment*, 2016, 568: 118-123.
- [7] Munns R. Physiological processes limiting plant growth in saline soils: some dogmas and hypotheses. *Plant, Cell & Environment*, 1993, 16(1): 15-24.
- [8] Sultana N, Ikeda T, Itoh R. Effect of NaCl salinity on photosynthesis and dry matter accumulation in developing rice grains. *Environmental and Experimental Botany*, 1999, 42(3): 211-220.
- [9] 林木菌根及其应用技术. 中国林业出版社, 1989.
- [10] 丛枝菌根生态生理. 华文出版社, 2001.
- [11] Talaat NB, Shawky BT. Protective effects of arbuscular mycorrhizal fungi on wheat (*Triticum aestivum* L.) plants exposed to salinity. *Environmental and Experimental Botany*, 2014, 98: 20-31.

- [12] 松嫩盐碱草地主要从枝菌根真菌对植物耐盐性影响的研究. 东北林业大学, 2015.
- [13] Sheng M, Tang M, Chen H, Yang BW, Zhang FF, Huang YH. Influence of arbuscular mycorrhizae on photosynthesis and water status of maize plants under salt stress. *Mycorrhiza*, 2008, 18(6/7): 287-296.
- [14] Lin JX, Wang YN, Sun SN, Mu CS, Yan XF. Effects of arbuscular mycorrhizal fungi on the growth, photosynthesis and photosynthetic pigments of *Leymus chinensis* seedlings under salt-alkali stress and nitrogen deposition. *Science of the Total Environment*, 2017, 576: 234-241.
- [15] 真菌提高枸杞耐盐性的机制研究. 西北农林科技大学, 2016.
- [16] 盐胁迫下丛枝菌根真菌 (AMF) 对养心菜耐盐性的影响. 四川农业大学, 2015.
- [17] 盐胁迫下沙枣幼苗的生长表现和生理特性. 福建林学院学报, 2014, 34(1): 64-70.
- [18] NaCl 胁迫对沙枣幼苗生长和光合特性的影响. 2014, 50(1): 32-40.
- [19] Phillips JM, Hayman DS. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Transactions of the British Mycological Society*, 1970, 55: 158-161.
- [20] 植物生理生化实验原理和技术. 高等教育出版社, 2000.
- [21] 王文杰等. 紫茎泽兰茎和叶片色素及叶绿素荧光相关参数对不同温度处理的响应差异. 2009, 29(10): 5424-5433.
- [22] 植物生态学报, 2001.
- [23] 应用生态学报, 1999, 10(5): 567-569.
- [24] 广东农业科学, 2008, (12): 58-60.
- [25] Allah EFA, Hashem A, Alqarawi AA, Bahkali AH, Alwhibi MS. Enhancing growth performance and systemic acquired resistance of medicinal *Sesbania sesban* (L.) Merr using arbuscular mycorrhizal fungi under salt stress. *Saudi Journal of Biological Sciences*, 2015, 22(3): 274-283.
- [26] Liu T, Sheng M, Wang CY, Chen H, Li Z, Tang M. Impact of arbuscular mycorrhizal fungi on the growth, water status, and photosynthesis of hybrid poplar under drought stress and recovery. *Photosynthetica*, 2015, 53(2): 250-258.
- [27] 水盐胁迫对沙枣幼苗叶绿素荧光参数和色素含量的影响. 西北农业学报, 2010, 19(12): 122-127.
- [28] Shakley TD, Seemann JR, Berry JA. Regulation of ribulose-1,5-bisphosphate carboxylase activity in response to changing partial pressure of O₂ and light in *Phaseolus vulgaris*. *Plant Physiology*, 1986, 81(3): 788-791.
- [29] 利用叶绿素荧光参数筛选抗盐菊芋品种的初步研究. 高技术通讯, 2008, 18(7): 766-770.
- [30] 叶绿素荧光测定技术在麦类作物耐盐性鉴定中的应用. 麦类作物学报, 2004, 24(3): 114-116.
- [31] 不同抗盐性小麦品种叶绿素荧光特性与其抗盐性关系的研究. 山东师范大学, 2008.
- [32] Evelin H, Kapoor R, Giri B. Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. *Annals of Botany*, 2009, 104(7): 1263-1280.
- [33] Sheng M, Tang M, Chen H, Yang BW, Zhang FF, Huang YH. Influence of arbuscular mycorrhizae on the root system of maize plants under salt stress. *Canadian Journal of Microbiology*, 2009, 55(7): 879-886.
- [34] 植物抗盐机理研究进展. 安徽农业科学, 2006, 34(23): 6111-6112.
- [35] Wu N, Li Z, Liu HG, Tang M. Influence of arbuscular mycorrhiza on photosynthesis and water status of *Populus cathayana* Rehder males and

females under salt stress. *Acta Physiologiae Plantarum*, 2015, 37(9): 183.
[36] Porcel R, Redondo-Gómez S, Mateos-Naranjo E, Aroca R, Garcia R, Ruiz-Lozano JM. Arbuscular mycorrhizal symbiosis ameliorates the optimum quantum yield of photosystem II and reduces non-photochemical quenching in rice plants subjected to salt stress. *Journal of Plant Physiology*, 2015, 185: 75-83.

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