

Comparison of Photosynthetic Characteristics Between Two Invasive Asteraceae Species and Native Plants in Liaoning Region (Postprint)

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Abstract

To elucidate the relationship between the physio-ecological traits of alien plants and their strong invasiveness, this study employed in-situ field measurements of gas exchange parameters to analyze the differences in photosynthetic characteristics and leaf traits between the Asteraceae invasive plants *Ambrosia artemisiifolia* and *Ambrosia trifida* and their co-occurring native plant *Aster tataricus*. The results showed that the light saturation point (LSP) values of the three plants—*A. artemisiifolia*, *A. trifida*, and *A. tataricus*—were all above $800 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$, and the LSP of the two invasive plants was significantly higher than that of *A. tataricus*, while their apparent quantum yield (AQY) was significantly lower than that of *A. tataricus*. The maximum net photosynthetic rate ($P_{n\max}$) of the three plants followed the order: *A. artemisiifolia* > *A. trifida* > *A. tataricus*, with the $P_{n\max}$ of *A. artemisiifolia* and *A. trifida* being 151.28% and 82.80% higher than that of *A. tataricus*, respectively, and the differences were significant. The specific leaf area (SLA), leaf nitrogen content per unit mass (N_{mass}), leaf phosphorus content per unit mass (P_{mass}), photosynthetic energy use efficiency (PEUE), and photosynthetic nitrogen use efficiency (PNUE) of the two invasive plants were all significantly higher than those of *A. tataricus*, but there was no significant difference in leaf construction cost per unit mass (CC_{mass}) among the three species. In summary, the two Asteraceae invasive plants in Liaoning exhibited higher gas exchange parameters and leaf trait indices compared to the native co-occurring species, and there existed a close relationship between their photosynthetic characteristics and leaf traits, manifested in these Asteraceae invasive plants having higher photosynthetic characteristic indices and energy utilization indices such as $P_{n\max}$, PNUE, PEUE, and water use efficiency (WUE), enabling the invasive plants to capture and utilize environmental resources more efficiently, which constitutes one of the reasons for their successful invasion.

Full Text

Preamble

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Comparison of Gas Exchange Characteristics Between Two Invasive Plants and
a Co-occurring Native Plant in Liaoning Province*

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Abstract: To clarify the relationship between ecophysiological traits and invasiveness in exotic plants, we conducted field measurements of gas exchange parameters to analyze differences in photosynthetic characteristics and leaf traits between two invasive Asteraceae species—common ragweed (*Ambrosia artemisiifolia*) and giant ragweed (*Ambrosia trifida*)—and their co-occurring native congener, *Aster tataricus*. The results revealed that all three species had light saturation points (LSP) exceeding $800 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$, with both invasive species exhibiting significantly higher LSP and lower apparent quantum yield (AQY) compared to the native species. Maximum net photosynthetic rate (Pnmax) followed the order: *A. artemisiifolia* > *A. trifida* > *A. tataricus*, with the two invasives showing 151.28% and 82.80% higher Pnmax than the native species, respectively ($P < 0.05$). Specific leaf area (SLA), leaf nitrogen content per unit mass (Nmass), leaf phosphorus content per unit mass (Pmass), photosynthetic energy use efficiency (PEUE), and photosynthetic nitrogen use efficiency (PNUE) were all significantly higher in the invasive species, though leaf construction cost per unit mass (CCmass) did not differ significantly among the three species. In summary, the two Asteraceae invaders in Liaoning exhibited superior gas exchange characteristics and leaf trait metrics relative to the native species. Their photosynthetic and leaf traits were closely linked, as evidenced by higher Pnmax, PNUE, PEUE, and water use efficiency (WUE). These enhanced traits enable invasive plants to capture and utilize environmental resources more effectively, likely contributing to their successful invasion.

Keywords: Asteraceae invasive plants; Photosynthetic characteristics; Maximum net photosynthetic rate; Leaf traits; Resource utilization efficiency

Introduction

Biological invasion poses one of the most serious environmental challenges of the 21st century, threatening ecosystems, agricultural economies, and public health worldwide [1-3]. Invasive plants typically exhibit rapid growth, strong ecological adaptability, high reproductive capacity, and efficient dispersal mechanisms [4], enabling them to outcompete native vegetation. While not all native species

are displaced by invaders, comparative studies of physiological and ecological traits between invasive and co-occurring native species provide valuable insights into invasion mechanisms. Beyond growth rate, key indicators associated with competitive ability include high photosynthetic rates, low construction costs (CC), and large total leaf area [5].

Specific leaf area (SLA) serves as an indirect indicator of leaf construction cost [5-6]. Lower construction costs (i.e., higher SLA) imply greater efficiency in utilizing plant carbon assimilates and higher photosynthetic nitrogen-use efficiency (PNUE) [7-8], which facilitates rapid growth [6] and confers competitive advantages to invaders [9-10]. For instance, Baruch and Goldstein [11] compared 30 invasive and 34 native species in Hawaii, finding that invaders generally had higher SLA and assimilation rates but lower leaf construction costs. Alpert et al. [12] and Wang et al. [13] advocated for comparative studies using closely related invasive and native species (e.g., within the same family or genus). Wang et al. [14] reported that *Chromolaena odorata* (Asteraceae) exhibited higher light use efficiency than its native congener *Gynura japonica*. Similarly, Wang et al. [15] found that the invasive plant *Conyza canadensis* had higher photosynthetic rates and biomass accumulation capacity than its associated native species, suggesting that high productivity contributes significantly to invasion success.

In China, the most common invasive plants belong to Asteraceae, Poaceae, and Brassicaceae [1], with Asteraceae representing one of the most heavily invaded families [16]. *Ambrosia artemisiifolia*, a North American annual weed in the tribe Heliantheae, is a globally recognized noxious invader [17] whose pollen is a potent allergen and major cause of hay fever, posing serious public health risks [18]. *Ambrosia trifida*, also a North American annual weed, was introduced to northeastern China in the 1940s and has become one of the most destructive invasive weeds in northern China [19]. This species forms monospecific stands, causing crop yield losses and threatening biodiversity while triggering hay fever in susceptible populations [20-21].

Few studies have compared ecophysiological traits between invasive and non-invasive species within specific taxonomic groups (family or genus). Since congeners share evolutionary backgrounds and biological characteristics, such comparisons are particularly valuable for revealing the unique physiological mechanisms underlying invasion success. Therefore, this study compared photosynthetic characteristics and leaf traits between invasive Asteraceae species and their native congener to elucidate how these invaders respond to varying light intensities, providing a scientific basis for risk assessment and management of potential invasive plants.

1. Materials and Methods

1.1 Study Area Description

The study site was located at Qipanshan, Shenyang, Liaoning Province (42°15'58" N, 123°51'43" E, elevation 192 m), a heavily infested area of both ragweed species. The region has a mid-temperate continental monsoon climate with cold, snowy winters and short, hot, rainy summers. Mean annual temperature is 7.41 °C, frost-free period is 148 days, mean annual precipitation is 750 mm, and relative humidity ranges from 65% to 70%. The sampling area represented a typical co-occurrence zone where invasive and native plants formed natural mixed communities with patchy distributions of invaders. The native species *Aster tataricus* (Asteraceae) was the dominant non-invasive associate, accompanied by other weeds including *Humulus scandens*, *Lactuca indica*, and *Setaria viridis*. Soil type was brown earth with pH 6.4, organic matter 14.62 g · kg⁻¹, total nitrogen 1.53 g · kg⁻¹, and total phosphorus 0.99 g · kg⁻¹.

Measurements and sample collection were conducted on August 27, 2015, when both invasive and native Asteraceae species had completed leaf morphogenesis and were in the peak vegetative growth to early flowering stage.

1.2 Experimental Methods

For each plant species, one 10 m × 10 m experimental block was established. Within each block, five 1 m × 1 m quadrats were randomly placed. Ten representative plants with relatively uniform growth were selected from each quadrat, and 30 healthy, intact mature leaves were randomly collected from each plant, yielding 5 × 30 = 150 leaves per species. All sampling was replicated five times within each quadrat. Leaf area was measured using a Li-3100A leaf area meter (Li-Cor, Lincoln, Nebraska, USA). Leaves were then placed in sealed bags, stored in ice boxes, and transported to the laboratory for determination of chlorophyll content, leaf N and P concentrations, and other physicochemical indices. Concurrently, soil samples (0-10 cm depth) were collected from the rhizosphere of each plant for physicochemical analysis.

Photosynthesis-Light Response (Pn-PAR) Measurements: Between 9:00-11:00 AM, plants in the vegetative growth stage were selected from each quadrat. A LI-6400 portable photosynthesis system (Li-Cor Inc., Lincoln, NE, USA) was used to measure net photosynthetic rate (Pn), transpiration rate (Tr), and stomatal conductance (Gs) on functional leaves [22]. Chamber temperature was set at 25 °C, CO₂ concentration at 400 mol · mol⁻¹, and photosynthetically active radiation (PAR) gradients at 2000, 1500, 1000, 800, 500, 300, 100, 50, and 0 mol · m⁻² · s⁻¹ using an automated measurement program [23]. Fully expanded mature leaves were measured while maintaining their natural orientation, with three measurements per leaf and five replicates per quadrat.

Photosynthetic light-response curves were fitted using the modified rectangular hyperbola model proposed by Ye [24] and Ye et al. [25]:

$$Pn = \frac{\alpha I + Pn_{max} - \sqrt{(\alpha I + Pn_{max})^2 - 4\alpha\beta I Pn_{max}}}{2\beta} - Rd$$

where Pn is net photosynthetic rate ($\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$), I is photosynthetically active radiation ($\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$), α is AQY (apparent quantum yield) representing the slope at $I = 0$, and β and Rd are coefficients independent of I ($\text{m}^2 \cdot \text{s} \cdot \text{mol}^{-1}$), and Rd is dark respiration rate ($\text{mg} \cdot \text{h}^{-1} \cdot \text{g}^{-1}$).

Water Use Efficiency (WUE): Calculated as [26]:

$$WUE = \frac{Pn}{Tr}$$

where Pn is photosynthetic rate ($\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$) and Tr is transpiration rate ($\text{mmol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$).

Chlorophyll Content Determination: Fresh leaves were cleaned with deionized water, punched into discs, and cut into thin strips. Pigments were extracted in 80% acetone until the strips turned completely white. Optical density was measured at 663 nm and 645 nm (OD_{663} , OD_{645}), and chlorophyll content calculated as [27]:

$$\rho(Chla) = 12.7 \times OD_{663} - 2.69 \times OD_{645}$$

$$\rho(Chlb) = 22.9 \times OD_{645} - 4.68 \times OD_{663}$$

$$\rho(Chl) = \rho(Chla) + \rho(Chlb)$$

where $(Chla)$, $(Chlb)$ are chlorophyll a and b concentrations ($\text{mg} \cdot \text{L}^{-1}$), and (Chl) is total chlorophyll concentration ($\text{mg} \cdot \text{L}^{-1}$).

Leaf Element Content Analysis: Plant samples were killed at 105 °C for 30 min, then oven-dried at 65 °C to constant weight. Dried leaves were ground, sieved, and stored in sealed bags. Total N was determined by Kjeldahl digestion, and total P by molybdenum-antimony colorimetry [28-30].

Leaf Ash Content and Construction Cost: Samples were ashed in a muffle furnace at 500 °C for 6 h, and the remaining residue was weighed. Dry weight calorific value was measured using an oxygen bomb calorimeter (HWR-15E, Shanghai Shangli Testing Instrument Factory). Approximately 0.5 g of powdered plant material was combusted completely, with five replicates per sample averaged to obtain dry weight calorific value [31].

Leaf construction cost per unit mass (CC_{mass}) was calculated as [31-32]:

$$CC_{mass} = \frac{(0.06968 \times CV - 0.065) \times (1 - Ash) + 7.5 \times k \times ON / 14.0067}{EG}$$

where Hc is ash-free calorific value ($\text{kJ} \cdot \text{g}^{-1}$), Ash is ash content (%), CV is dry weight calorific value ($\text{kJ} \cdot \text{g}^{-1}$), CC_{mass} is leaf construction cost per unit mass [$\text{g}(\text{glucose}) \cdot \text{g}^{-1}$], ON is organic nitrogen content (%), EG is growth efficiency, and k is the oxidation state of nitrogen [31-32]. The average leaf growth efficiency for different plants is 0.87 [33]. For each sample, calculations were performed separately using both ammonium nitrogen (-3 valence) and nitrate nitrogen (+5 valence) as nitrogen oxidation states, then weighted averages were computed based on soil $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$ ratios to determine final CC_{mass} .

Additional calculations:

$$SLA = \frac{ULA}{DW}$$

$$CC_{area} = \frac{CC_{mass}}{SLA}$$

$$N_{area} = N_{mass} \times SLA$$

$$PEUE = \frac{Pn_{max}}{CC_{area}}$$

$$PNUE = \frac{Pn_{max}}{N_{area}}$$

where SLA is specific leaf area ($\text{cm}^2 \cdot \text{g}^{-1}$), ULA is unit leaf area (cm^2), DW is leaf dry weight (g), CC_{area} is leaf construction cost per unit area [$\text{g}(\text{glucose}) \cdot \text{m}^{-2}$], N_{mass} is leaf N content per unit mass ($\text{mg} \cdot \text{g}^{-1}$), N_{area} is leaf N content per unit area ($\text{g} \cdot \text{m}^{-2}$), $PEUE$ is photosynthetic energy use efficiency [$\mu\text{mol}(\text{CO}_2) \cdot \text{g}^{-1}(\text{glucose}) \cdot \text{s}^{-1}$], and $PNUE$ is photosynthetic nitrogen use efficiency [$\mu\text{mol}(\text{CO}_2) \cdot \text{g}^{-1} \cdot \text{s}^{-1}$].

1.3 Data Analysis

Light saturation point (LSP), light compensation point (LCP), apparent quantum yield (AQY), and maximum net photosynthetic rate (Pn_{max}) were calculated following Chen et al. [23]. All data were analyzed using SPSS 16.0 and plotted with Excel 2007. Results are presented as means \pm standard error. One-way ANOVA and least significant difference (LSD) tests were used for interspecific comparisons at $P = 0.05$.

2. Results

2.1 Photosynthetic Response to Photosynthetically Active Radiation

As shown in [Figure 1: see original paper], net photosynthetic rate (P_n) in both invasive ragweed species and the native *A. tataricus* initially increased then decreased with increasing PAR, with invaders showing greater increases. At specific PAR levels, P_n reached maxima corresponding to light saturation points. Correlations between P_n and PAR were highly significant for *A. artemisiifolia* ($R^2 = 0.997$, $P < 0.001$), *A. trifida* ($R^2 = 0.999$, $P < 0.001$), and *A. tataricus* ($R^2 = 0.995$, $P < 0.001$). Photosynthetic response parameters derived from the modified rectangular hyperbola model are presented in .

All three species had LSP values exceeding $800 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$, following the order: *A. trifida* > *A. artemisiifolia* > *A. tataricus*, with significant differences among them. Light compensation point (LCP) ranked as *A. tataricus* > *A. trifida* > *A. artemisiifolia*, with the native species' LCP ($30.736 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$) significantly higher than that of *A. artemisiifolia*. Both invasive species had significantly lower AQY than the native species ($P < 0.05$). Maximum net photosynthetic rate followed the order: *A. artemisiifolia* > *A. trifida* > *A. tataricus*, with invasive species showing 151.28% and 82.80% higher $P_{n\max}$ than the native species, respectively ($P < 0.05$). These results demonstrate that the two invasive Asteraceae species possess higher light use efficiency and stronger photosynthetic assimilation capacity under identical PAR conditions.

Stomatal conductance (G_s), transpiration rate (T_r), and water use efficiency (WUE) increased with PAR in all three species. However, when PAR exceeded $500 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$, G_s , T_r , and WUE in the native *A. tataricus* plateaued rather than continuing to increase, whereas both invasive species maintained increasing G_s and T_r . As shown in [Figure 1: see original paper]d, WUE increased more dramatically with PAR in the invasive species compared to the native species, indicating superior water use efficiency.

2.2 Leaf Trait Comparisons

As shown in , chlorophyll content was significantly higher in both invasive species than in the native species ($P < 0.05$), ranking: *A. artemisiifolia* > *A. trifida* > *A. tataricus*. Specific leaf area (SLA), leaf nitrogen content per unit mass (N_{mass}), leaf phosphorus content per unit mass (P_{mass}), photosynthetic energy use efficiency (PEUE), and photosynthetic nitrogen use efficiency (PNUE) were all significantly higher in the invasive species compared to the native *A. tataricus* ($P < 0.05$). However, leaf construction cost per unit mass (CC_{mass}) did not differ significantly among the three species.

3. Discussion

Invasive plants can fundamentally alter the biological characteristics of natural communities [34-35]. Successful invasion depends on multiple factors, with

physiological traits being critical for survival, establishment, and spread. Some exotic species possess stronger light use efficiency, photosynthetic responsiveness, and ecophysiological characteristics than natives, conferring high invasion potential [36]. Plant invasiveness is determined by interactions between environmental factors and plant traits; under equivalent reproductive capacity and enemy pressure, invasive species often outcompete natives by more efficiently utilizing limited resources or exploiting resources at times when natives cannot [37].

3.1 Differences in Gas Exchange Parameters

Plant light absorption capacity and photosynthetic acclimation largely reflect growth and competitive ability [38]. Light saturation point (LSP) and light compensation point (LCP) indicate adaptability to high and low light environments [39], while AQY and Pn reflect photosynthetic apparatus efficiency and primary productivity [40]. In this study, the greater increases in Pn, Gs, and WUE in invasive ragweed species compared to native *A. tataricus* suggest broader stomatal plasticity, enabling invaders to adjust to environmental changes and assimilate more CO₂ with minimal water loss. Water use efficiency is a crucial physiological indicator, particularly important for plants in arid environments [26]. Despite increasing Gs with PAR, invasive species maintained high WUE because enhanced Pnmax compensated for water loss [41]. Both ragweed species achieved higher maximum WUE than the native species, confirming their superior water use efficiency. These findings align with studies on *Ageratina adenophora* [42] and *Conyza canadensis* [15], which also reported higher WUE in invaders compared to co-occurring natives.

3.2 Differences in Leaf Traits

Since CO₂ assimilation occurs in chloroplasts, chlorophyll content directly affects photosynthesis [44]. Both invasive ragweed species had significantly higher chlorophyll content than native *A. tataricus*. SLA is closely related to nitrogen allocation [45]; high nitrogen enhances photosynthetic capacity and carbon accumulation in invasive plants [46]. Phosphorus plays vital roles in membrane structure, metabolism, enzyme regulation, and signal transduction, significantly impacting photosynthesis [47]. Higher leaf N and P contents promote plant growth and photosynthetic capacity [45]. In this study, both invasive species had significantly higher N_{mass} and P_{mass} than the native species, suggesting stronger nutrient uptake capabilities [37].

Feng et al. [42] proposed that invasive populations evolve higher N_{mass} and SLA than native conspecifics due to enemy release. With reduced herbivore pressure in introduced ranges, invaders allocate less nitrogen to cell walls (defense compounds) and more to photosynthetic machinery, enhancing competitiveness. Both ragweed species showed significantly higher SLA than the native species, along with elevated N_{mass} and P_{mass}. Construction cost (CC_{mass}), PEUE, and PNUE reflect plant energy strategies [33]. While lower CC_{mass}

could confer competitive advantages [32], the high P_{nmax} in invasive species did not correspond to lower CC_{mass} , likely because high nitrogen content and protein synthesis costs increase construction costs [42], consistent with Geng et al. [41]. $PNUE$ represents nitrogen utilization efficiency [48], while $PEUE$ directly reflects energy use efficiency. Both parameters were significantly higher in invasive species, corroborating Funk et al. [49].

Photosynthetic rate (P_n) determines carbon accumulation capacity and competitive strength [50]. The high P_n in invasive species did not reduce CC_{mass} , consistent with Penning de Vries et al. [33] and Feng et al. [42], likely because elevated $PEUE$ and $PNUE$ —key indicators of leaf energy use efficiency [51]—offset higher construction costs, making these parameters more relevant for assessing resource acquisition capacity.

Conclusion

This study demonstrates that net photosynthetic rates of both invasive ragweed species and native *A. tataricus* are highly significantly correlated with PAR. All three species had LSP values above $800 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$, ranking: *A. trifida* > *A. artemisiifolia* > *A. tataricus*. LCP ranked: *A. tataricus* > *A. trifida* > *A. artemisiifolia*, with the native species significantly higher than *A. artemisiifolia*. Both invasive species had significantly lower AQY than the native species. P_{nmax} ranked: *A. artemisiifolia* > *A. trifida* > *A. tataricus*. The invasive species showed greater WUE enhancement with increasing PAR. Overall, under identical conditions, the two invasive Asteraceae species demonstrated superior photosynthetic capacity, light use efficiency, and water use efficiency compared to the native species.

References

- [1] Ju R T, Li H, Shi Z R, et al. Progress of biological invasions research in China over the last decade[J]. Biodiversity Science, 2012, 20(5): 581–611
- [2] Westphal M I, Browne M, MacKinnon K, et al. The link between international trade and the global distribution of invasive alien species[J]. Biological Invasions, 2008, 10(4): 391–398
- [3] Xu G F, Shen S C, Zhang F D, et al. Research progress and prospect about exotic invasive species *Phalaris minor* Retz[J]. Chinese Journal of Eco-Agriculture, 2015, 23(9): 1083–1092
- [4] Pattison R R, Goldstein G, Ares A. Growth, biomass allocation and photosynthesis of invasive and native Hawaiian rainforest species[J]. Oecologia, 1998, 117(4): 449–459
- [5] McDowell S C L. Photosynthetic characteristics of invasive and noninvasive species of *Rubus* (Rosaceae)[J]. American Journal of Botany, 2002, 89(9): 1431–1438
- [6] Durand L Z, Goldstein G. Photosynthesis, photoinhibition, and nitrogen use efficiency in native and invasive tree ferns in Hawaii[J]. Oecologia, 2001, 126(3):

345-354

- [7] Feng Y L, Fu G L. Nitrogen allocation, partitioning and use efficiency in three invasive plant species in comparison with their native congeners[J]. *Biological Invasions*, 2008, 10(6): 891-902
- [8] Matzek V. Superior performance and nutrient-use efficiency of invasive plants over non-invasive congeners in a resource-limited environment[J]. *Biological Invasions*, 2011, 13(12): 3005-3014
- [9] Smith M D, Knapp A K. Physiological and morphological traits of exotic, invasive exotic, and native plant species in tallgrass prairie[J]. *International Journal of Plant Sciences*, 2001, 162(4): 785-792
- [10] Daehler C C. Performance comparisons of co-occurring native and alien invasive plants: Implications for conservation and restoration[J]. *Annual Review of Ecology, Evolution, and Systematics*, 2003, 34: 183-211
- [11] Baruch Z, Goldstein G. Leaf construction cost, nutrient concentration, and net CO₂ assimilation of native and invasive species in Hawaii[J]. *Oecologia*, 1999, 121(2): 183-192
- [12] Alpert P, Bone E, Holzapfel C. Invasiveness, invasibility and the role of environmental stress in the spread of non-native plants[J]. *Perspectives in Plant Ecology, Evolution and Systematics*, 2000, 3(1): 52-66
- [13] Wang R F, Feng Y L. The effects of leaf phenology, construction cost and payback time on carbon accumulation in invasive plants[J]. *Acta Ecologica Sinica*, 2009, 29(5): 2568-2577
- [14] Wang J F, Feng Y L, Li Z. Acclimation of photosynthesis to growth light intensity in *Chromolaena odorata* L. and *Gynura* sp.[J]. *Journal of Plant Physiology and Molecular Biology*, 2003, 29(6): 542-548
- [15] Wang X H, Ji M S. Photosynthetic characteristics of an invasive plant *Conyza canadensis* and its associated plants[J]. *Chinese Journal of Applied Ecology*, 2013, 24(1): 71-77
- [16] Wan F H, Guo J Y, Zhang F, et al. *Research on Biological Invasions in China*[M]. Beijing: Science Press, 2009
- [17] Friedman J, Barrett S C H. High outcrossing in the annual colonizing species *Ambrosia artemisiifolia* (Asteraceae)[J]. *Annals of Botany*, 2008, 101(9): 1303-1309
- [18] Mandrioli P, Di Cecco M, Andina G. Ragweed pollen: The aeroallergen is spreading in Italy[J]. *Aerobiologia*, 1998, 14(1): 13-20
- [19] Wang D L, Zhu X R. Allelopathic research of *Ambrosia trifida*[J]. *Acta Phytocologica Sinica*, 1996, 20(4): 330-337
- [20] Sha W, Zhou F J, Zu Y G. The population genetic structure of *Ambrosia trifida* in different environment[J]. *Bulletin of Botanical Research*, 2000, 20(1): 94-98
- [21] Wei S H, Qu Z, Zhang C X, et al. Invasive alien species giant ragweed (*Ambrosia trifida* L.) and its risk assessment[J]. *Plant Protection*, 2006, 32(4): 14-19
- [22] Wu Y Q, Hu Y J. Researches on photosynthetic characteristics of exotic plants *Wedelia trilobata*, *Pharbitis nil* and *Ipomoea cairica*[J]. *Acta Ecologica Sinica*, 2004, 24(10): 2334-2339

- [23] Chen X W, Wei Z S, Liu H M, et al. Comparison of photosynthetic characteristics between invasive and co-occurring native Asteraceae plants in Yunnan Province, China[J]. Research of Environmental Sciences, 2016, 29(4): 538-546
- [24] Ye Z P. A new model for relationship between irradiance and the rate of photosynthesis in *Oryza sativa*[J]. Photosynthetica, 2007, 45(4): 637-640
- [25] Ye Z P, Yu Q. Comparison of new and several classical models of photosynthesis in response to irradiance[J]. Journal of Plant Ecology, 2008, 32(6): 1356-1361
- [26] Gao L, Yang J, Liu R X. Effects of soil moisture levels on photosynthesis, transpiration, and moisture use efficiency of female and male plants of *Hippophae rhamnoides* ssp. *sinensis*[J]. Acta Ecologica Sinica, 2009, 29(11): 6025-6034
- [27] Shu Z, Zhang X S, Chen J, et al. The simplification of chlorophyll content measurement[J]. Plant Physiology Communications, 2010, 46(4): 399-402
- [28] Gu D X, Chen S L, Huang Y Q. Effects of soil nitrogen and phosphonium on leaf nitrogen and phosphonium stoichiometric characteristics and chlorophyll content of *Oligostachyum lubricum*[J]. Chinese Journal of Plant Ecology, 2011, 35(12): 1219-1225
- [29] Sparks D L, Page A L, Helmke P A, et al. Methods of Soil Analysis, Part 3: Chemical Methods[M]. Madison: Soil Science Society of America, American Society of Agronomy, 1996
- [30] Lu R K. Methods of Soil Agricultural Chemistry Analysis[M]. Beijing: Chinese Agricultural Science and Technology Press, 2000: 296-338
- [31] Williams K, Percival F, Merino J, et al. Estimation of tissue construction cost from heat of combustion and organic nitrogen content[J]. Plant, Cell & Environment, 1987, 10(9): 725-734
- [32] Tu C Y, Huangfu C H, Jiang N, et al. Comparison of leaf construction cost between invasive plant *Flaveria bidentis* and its five co-occurring plants[J]. Chinese Journal of Ecology, 2013, 32(11): 2985-2991
- [33] Penning De Vries F W T, Brunsting A H M, Van Laar H H. Products, requirements and efficiency of biosynthesis: A quantitative approach[J]. Journal of Theoretical Biology, 1974, 45(2): 339-377
- [34] Jäger H, Alencastro M J, Kaupenjohann M, et al. Ecosystem changes in Galápagos highlands by the invasive tree *Cinchona pubescens*[J]. Plant and Soil, 2013, 371(1/2): 629-640
- [35] Evans R D, Rimer R, Sperry L, et al. Exotic plant invasion alters nitrogen dynamics in an arid grassland[J]. Ecological Applications, 2001, 11(5): 1301-1310
- [36] Poorter L. Light-dependent changes in biomass allocation and their importance for growth of rain forest tree species[J]. Functional Ecology, 2001, 15(1): 113-123
- [37] Ehrenfeld J G. Effects of exotic plant invasions on soil nutrient cycling processes[J]. Ecosystems, 2003, 6(6): 503-523
- [38] Zhang J L, Cao K F. The effect of irradiance on photosynthetic capacity, heat dissipation, and antioxidants of seedlings of two tropical rainforest tree species[J]. Acta Phytocologica Sinica, 2002, 26(6): 639-646

- [39] Chown S L, Slabber S, McGeoch M A, et al. Phenotypic plasticity mediates climate change responses among invasive and indigenous arthropods[J]. *Proceedings of the Royal Society B: Biological Sciences*, 2007, 274(1625): 2531-2537
- [40] Feng Y L, Cao K F, Feng Z L, et al. Acclimation of lamina mass per unit area, photosynthetic characteristics and dark respiration to growth light regimes in four tropical rainforest species[J]. *Acta Ecologica Sinica*, 2002, 22(6): 901-910
- [41] Geng X Y, Jiang S, Li B, et al. Do higher resource capture ability and utilization efficiency facilitate the successful invasion of exotic plant? A case study of *Alternanthera philoxeroides*[J]. *American Journal of Plant Sciences*, 2013, 4(9): 2197-2205
- [42] Feng Y L, Li Y P, Wang R F, et al. A quicker return energy-use strategy by populations of a subtropical invader in the non-native range: A potential mechanism for the evolution of increased competitive ability[J]. *Journal of Ecology*, 2011, 99(5): 1116-1123
- [43] Liang Z P, Li L Q, Wan F H, et al. Feedback of soil biota on *Ageratina adenophora* growth and competitiveness with native plant: A comparison of different sterilization methods[J]. *Chinese Journal of Eco-Agriculture*, 2016, 24(9): 1223-1230
- [44] Ye Z P, Zhao Z H. Effects of shading on the photosynthesis and chlorophyll content of *Bidens pilosa*[J]. *Chinese Journal of Ecology*, 2009, 28(1): 19-22
- [45] Feng Y L, Fu G L, Zheng Y L. Specific leaf area relates to the differences in leaf construction cost, photosynthesis, nitrogen allocation, and use efficiencies between invasive and noninvasive alien congeners[J]. *Planta*, 2008, 228(3): 383-390
- [46] Wang M L, Feng Y L. Effects of soil nitrogen levels on morphology, biomass allocation and photosynthesis in *Ageratina adenophora* and *Chromolaena odorata*[J]. *Acta Phytocologica Sinica*, 2005, 29(5): 697-705
- [47] Li Z G, Xie P D, Zhang Y L, et al. Influence to photosynthesis of different genotype soybean with phosphorus stress[J]. *Journal of Inner Mongolia University for Nationalities*, 2004, 19(3): 297-299
- [48] Li Y. Studies on mechanisms of effects of different nitrogen supplies on photosynthesis and photosynthetic nitrogen use efficiency of rice plants[D]. Nanjing: Nanjing Agricultural University, 2011
- [49] Funk J L, Vitousek P M. Resource-use efficiency and plant invasion in low-resource systems[J]. *Nature*, 2007, 446(7139): 1079-1081
- [50] Wang K M, Hou Y T. *Parthenium* L (Asteraceae) a newly naturalized record genus in Shandong Province[J]. *Journal of Qufu Normal University*, 2004, 30(1): 83-84
- [51] Meziane D, Shipley B. Direct and indirect relationships between specific leaf area, leaf nitrogen and leaf gas exchange: Effects of irradiance and nutrient supply[J]. *Annals of Botany*, 2001, 88(5): 915-927

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