

## Characteristics of Plant Extracts and Their Application in Sow Production: Postprint

**Authors:** Wang Hao, 印遇龙' 邓百川' 邓小平, Tan Chengquan

**Date:** 2017-11-08T00:00:00+00:00

### Abstract

Sow reproductive performance constitutes a critical determinant of production levels and economic efficiency in pig farming operations, yet a significant gap persists between the annual productivity of sows in China and that achieved in developed European and American countries. Although antibiotics employed as feed additives have improved sow reproductive performance to a certain degree, their excessive utilization in feed engenders issues such as the proliferation of drug-resistant bacteria and residues in meat products, thereby jeopardizing human health and the ecological environment. Plant extracts are plant-derived additive products obtained from botanical sources through physical or chemical methods, containing multiple natural bioactive substances. In recent years, the application of plant extracts as antibiotic alternatives in sow production has garnered extensive attention. This paper synthesizes domestic and international research progress to review the characteristics of plant extracts and their effects on sow production performance, aiming to provide a reference for further in-depth research on the application of plant extracts in sow production.

### Full Text

#### Plant Extracts: Characteristics and Application in Sow Production

**WANG Hao, YIN Yulong, DENG Baichuan, DENG Jinping, TAN Chengquan**

College of Animal Science, South China Agricultural University, Guangzhou 510642, China

### Abstract

The reproductive performance of sows is a primary factor that determines production level and profitability in swine enterprises, while the number of piglets

per sow per year in China remains lower than in the USA and EU. Although antibiotics can improve sow performance, their overuse in diets has caused problems such as bacterial resistance and drug residues in pork products, which are harmful to human health and the ecological environment. Plant extracts are phytogetic products consisting of various natural active components, produced through physical or chemical extraction from natural plants. In recent years, plant extracts have attracted increased attention as alternatives to antibiotics in the sow industry. Referring to domestic and overseas research, this paper reviews the characteristics of plant extracts and their effects on sow performance, as well as practical application effects, to provide a reference for further study on the application of plant extracts in sow feeding.

**Keywords:** plant extracts; characteristics; sows; application

\*Corresponding authors: TAN Chengquan, lecturer, E-mail: tanchengquan@scau.edu.cn; DENG Jinping, professor, E-mail: 360437040@qq.com

---

## 1 Introduction

Sow reproductive performance is a critical factor determining production efficiency and economic returns in pig farming. However, the current piglets-per-sow-per-year figure in China still lags behind that of the United States and European Union. While antibiotics have been used to enhance sow performance, their excessive use in feed has led to serious issues including bacterial resistance and drug residues in pork products, posing risks to human health and the environment. Plant extracts, derived from natural plants through physical or chemical extraction processes, contain diverse bioactive compounds and have gained significant attention as potential antibiotic alternatives in sow production. This review synthesizes domestic and international research on the characteristics of plant extracts, their effects on sow performance, and practical application outcomes to provide a foundation for future research and application in sow nutrition.

### 1.1 Antioxidant Properties

Oxidative stress is a major concern during late gestation and lactation in sows, characterized by increased production of reactive oxygen species (ROS) and lipid peroxidation. Plant extracts rich in polyphenolic compounds can effectively scavenge free radicals and enhance antioxidant capacity. Studies have shown that dietary supplementation with plant extracts significantly reduces oxidative stress markers. For instance, supplementation with 15 mg/kg of certain extracts decreased 8-hydroxydeoxyguanosine (8-OHdG) and thiobarbituric acid reactive substances (TBARS) levels in sows [8]. Similarly, administration of 45 mg/kg reduced malondialdehyde (MDA) while increasing superoxide dismutase (SOD), total antioxidant capacity (T-AOC), and catalase (CAT) activities [9-11]. At 200 mg/kg, extracts have been shown to lower MDA and elevate glutathione

peroxidase (GSH-Px) activity [12]. These findings demonstrate dose-dependent antioxidant benefits that improve sow health and performance.

### 1.2 Effects on Production Performance

Plant extracts improve sow performance through multiple pathways, including enhanced lactation feed intake, reduced weight loss, and improved milk quality. Supplementation during gestation and lactation increases weaning weights and average daily gain (ADG) of piglets. Table 1 summarizes the effects of various plant extracts on sow performance. Key findings include: star anise supplementation increased weaning weight by 10.3-11.6% [31]; ginger extracts improved ADG by 11.6% [33]; and oregano essential oil enhanced piglet weight gain by 12.2-14.8% [8, 35]. Compound plant extracts containing multiple active components have shown even greater efficacy, with some formulations improving weaning weight by 13.4-19.4% [39-41]. These improvements are attributed to enhanced milk production, better nutrient digestibility, and reduced pathogen load.

### 1.3 Immune Function Enhancement

Plant extracts modulate immune function by increasing immunoglobulin levels and regulating cytokine production. Studies demonstrate that 200 mg/kg ginseng polysaccharide supplementation elevated IgG concentrations in sow colostrum and milk, improving passive immunity transfer to piglets [13]. Similarly, 200 mg/kg herbal supplementation increased IgG by 15% and IgA levels while reducing interleukin-6 (IL-6) [27]. At 90 mg/kg, extracts enhanced IL-2 production by 28% and increased IgG while decreasing IL-6 [13]. These immunomodulatory effects strengthen both innate and adaptive immunity, reducing sow susceptibility to mastitis-metritis-agalactia (MMA) syndrome and improving overall herd health.

### 1.4 Weaning-to-Estrus Interval (WEI)

Shortening the WEI is crucial for improving sow reproductive efficiency. Plant extracts can reduce WEI by alleviating oxidative stress and improving uterine recovery post-weaning. Research indicates that phytogetic supplementation decreases WEI by 7-21 days compared to control groups [20-22, 30]. The mechanism involves reduced inflammation and enhanced endometrial repair through antioxidant and antimicrobial actions. Oregano essential oil, for example, has been shown to improve farrowing rates and reduce culling due to reproductive failure [20, 21].

## 2 Mechanism of Action

The beneficial effects of plant extracts on sow performance are mediated through three primary mechanisms (Fig. 1):

**Antioxidant Activity:** Polyphenols and flavonoids scavenge ROS, upregulate antioxidant enzymes (SOD, CAT, GSH-Px), and reduce lipid peroxidation. This protects mammary tissue from oxidative damage during peak lactation [44, 45].

**Antimicrobial Effects:** Essential oils containing thymol, carvacrol, and eugenol disrupt bacterial cell membranes, inhibit pathogenic proliferation, and reduce endotoxin production. This decreases the incidence of swine urogenital disease (SUGD) and MMA syndrome [8, 21-22, 35-36, 38-39].

**Immunomodulation:** Bioactive compounds activate nuclear factor erythroid 2-related factor 2 (Nrf2) pathways, suppress NF- $\kappa$ B signaling, and modulate cytokine expression (IL-2, IL-6, TNF- $\alpha$ ), thereby reducing inflammatory responses [46-49].

### 3 Gut Microbiota Modulation

Plant extracts positively influence gut microbiota composition by promoting beneficial bacteria (Lactobacillus, Bifidobacterium) and inhibiting pathogens (E. coli, Salmonella). This modulation increases short-chain fatty acid (SCFA) production, enhances gut barrier integrity, and reduces noxious gas emissions [50-53]. The prebiotic-like effects of certain polysaccharides support microbial balance, which is critical for nutrient absorption and immune development in both sows and piglets [54, 55]. Improved gut health translates to better feed efficiency and reduced post-weaning diarrhea in piglets.

### 4 Conclusion and Future Directions

Plant extracts represent viable alternatives to antibiotics in sow production, offering multifaceted benefits including antioxidant protection, immune enhancement, and microbiota modulation. While current research demonstrates positive effects on performance, immune status, and reproductive efficiency, several knowledge gaps remain. Future studies should focus on: (1) optimizing dosage and delivery methods for different production stages; (2) investigating synergistic effects of compound extracts; (3) elucidating precise molecular mechanisms; and (4) evaluating long-term impacts on sow longevity and piglet development. Standardized extraction protocols and quality control measures are essential for consistent field application. As the industry moves toward antibiotic-free production, plant extracts will play an increasingly important role in sustainable swine nutrition.

---

**Table 1** Effects of plant extracts on performance of sows

Plant Name	Main Active Components	Duration of Supplement/d	Supplemental Dose/(mg/kg)	Treatment Effects (vs. Control)/%
Star anise	6-anethole, 6-shikimic acid	G85-L21	5.0	+10.3 (weaning weight)
Star anise	6-anethole, 6-shikimic acid	G85-L21	7.0	+11.6 (weaning weight)
Dan-shen root	Tanshinone IIA	G85-L28	3.0	+11.6 (ADG)
Ginger	Gingerol	G107-L26	2.5	+11.6 (ADG)
Soy	Isoflavones	L1-L18	3.2	+14.8 (weaning weight)
Fenugreek seed	Saponins	G90-L28	2.5	+12.2 (ADG)
Seaweed	Polysaccharides	G1-L21	3.6	+13.9 (weaning weight)
Milk thistle	Silymarin	G1-L19	2.4	+10.5 (weaning weight)
Ginseng	Ginsenosides	G109-L22	4.3	+11.1 (ADG)
Oregano	Carvacrol, thymol	G110-L21	2.1	+10.4 (ADG)
Yucca shidig-era	Saponins	L1-E1	5 g/d	+11.5 (weaning weight)
Quillaja saponaria	Saponins	G107-L23	10 g/d	+16.4 (weaning weight)
Plant ex-tracts compound	Mixed	G90-L28	3.4	+13.4 (weaning weight)
Plant ex-tracts compound	Mixed	G108-L25	2.6	+12.4 (ADG)
Plant ex-tracts compound	Mixed	G107-L28	2.1	+10.5 (weaning weight)

Note: “G” = gestation, “L” = lactation, “E” = estrus. For example, “G85-L21” indicates supplementation from day 85 of gestation to day 21 of lactation.

---

**Fig. 1** Potential mechanism of plant extracts for affecting performance of sows

The figure illustrates three main pathways: (1) Antioxidant pathway -ROS scavenging and Nrf2 activation; (2) Antimicrobial pathway -inhibition of SUGD/MMA pathogens; (3) Immunomodulatory pathway -NF- $\kappa$ B suppression and cytokine regulation.

---

## References

- [1] EISSEN J J, KANIS E, KEMP B. Sow factors affecting voluntary feed intake during lactation[J]. *Livestock Production Science*, 2000, 64(2/3): 147-165.
- [2] ENGBLOM L, LUNDEHEIM N, STRANDBERG E, et al. Factors affecting length of productive life in Swedish commercial sows[J]. *Journal of Animal Science*, 2008, 86(2): 432-441.
- [3] KIM S W, WEAVER A C, SHEN Y B, et al. Improving efficiency of sow productivity: nutrition and health[J]. *Journal of Animal Science and Biotechnology*, 2013, 4(1): 26.
- [4] MAXWELL C V, COMBS G E, KNABE D A, et al. Effect of dietary chlortetracycline during breeding and/or farrowing and lactation on reproductive performance of sows: a cooperative study. S-145 committee on nutritional systems for swine to increase reproductive efficiency[J]. *Journal of Animal Science*, 1994, 72(12): 3169-3176.
- [5] ZENG Z K, ZHANG S, WANG H L, et al. Essential oil and aromatic plants as feed additives in non-ruminant nutrition: a review[J]. *Journal of Animal Science and Biotechnology*, 2015, 6(1): 7.
- [6] SI W, GONG J, TSAO R, et al. Antimicrobial activity of essential oils and structurally related synthetic food additives towards selected pathogenic beneficial bacteria[J]. *Journal of Applied Microbiology*, 2006, 100(2): 296-305.
- [7] CAPECKA E, MARECZEK A, LEJA M. Antioxidant activity of fresh and dry herbs of some Lamiaceae species[J]. *Food Chemistry*, 2005, 93(2): 223-226.
- [8] TAN C Q, WEI H K, SUN H Q, et al. Effects of dietary supplementation of oregano essential oil to sows on oxidative stress status, lactation intake of sows, and piglet performance[J]. *BioMed Research International*, 2015, 2015(2): 525218.
- [9] BREWER M S. Natural antioxidants: sources, compounds, mechanisms of action, and potential applications[J]. *Comprehensive Reviews in Food Science and Food Safety*, 2011, 10(4): 221-247.

- [10] MUCHUWETI M, KATIVU E, MUPURE C H, et al. Phenolic composition and antioxidant properties of some spices[J]. *American Journal of Food Technology*, 2007, 2(5): 414-420.
- [11] KÄHKÖNEN M P, HOPIA A I, VUORELA H J, et al. Antioxidant activity of plant extracts containing phenolic compounds[J]. *Journal of Agricultural and Food Chemistry*, 1999, 47(10): 3954-3962.
- [12] HU Y J, GAO K G, ZHENG C T, et al. Effect of dietary supplementation with glycitein during late pregnancy and lactation on antioxidative indices and performance of primiparous sows[J]. *Journal of Animal Science*, 2015, 93(5): 2246-2254.
- [13] XI Q Y, JIANG Y, ZHAO S, et al. Effect of ginseng polysaccharides on the immunity and growth of piglets by dietary supplementation during late pregnancy and lactating sows[J]. *Animal Science Journal*, 2016, 88(6): 863-872.
- [14] BURT S. Essential oils: their antibacterial properties and potential applications in foods—a review[J]. *International Journal of Food Microbiology*, 2004, 94(3): 223-253.
- [15] FRIEDMAN M, HENIKA P R, MANDRELL R E. Bactericidal activities of plant essential oils and some of their isolated constituents against *Campylobacter jejuni*, *Escherichia coli*, *Listeria monocytogenes*, and *Salmonella enterica*[J]. *Journal of Food Protection*, 2002, 65(10): 1545-1560.
- [16] BOGAVAC M, KARAMAN M, JANJUŠEVIĆ L, et al. Alternative treatment of vaginal infections—in vitro antimicrobial and toxic effects of *Coriandrum sativum* L. and *Thymus vulgaris* L. essential oils[J]. *Journal of Applied Microbiology*, 2015, 119(3): 697-710.
- [17] KARAMAN M, BOGAVAC M, RADOVANOVIĆ B, et al. *Origanum vulgare* essential oil affects pathogens causing vaginal infections[J]. *Journal of Applied Microbiology*, 2017, 122(5): 1177-1185.
- [18] FRATINI F, CASELLA S, LEONARDI M, et al. Antibacterial activity of essential oils, their blends and mixtures of their main constituents against some strains supporting livestock mastitis[J]. *Fitoterapia*, 2014, 96: 1-7.
- [19] SARKAR P, KUMAR H, RAWAT M, et al. Effect of administration of garlic extract and PGF<sub>2</sub> on hormonal changes and recovery in endometritis cows[J]. *Asian-Australasian Journal of Animal Sciences*, 2006, 19(7): 964-969.
- [20] MAUCH C, BILKEI G. Strategic application of oregano feed supplements reduces sow mortality and improves reproductive performance—a case study[J]. *Journal of Veterinary Pharmacology and Therapeutics*, 2004, 27(1): 61-63.
- [21] AMRIK B, BILKEI G. Influence of farm application of oregano on performances of sows[J]. *Canadian Veterinary Journal*, 2004, 45(8): 674-677.
- [22] ALLAN P, BILKEI G. Oregano improves reproductive performance of sows[J]. *Theriogenology*, 2005, 63(3): 716-721.

- [23] HOSSAIN M M, BEGUM M, NYACHOTI C M, et al. Dietary fenugreek seed extract improves performance and reduces fecal *E. coli* counts and fecal gas emission in lactating sows and suckling piglets[J]. *Canadian Journal of Animal Science*, 2015, 95(4): 561-568.
- [24] LEONARD S G, SWEENEY T, BAHAR B, et al. Effect of dietary seaweed extracts and fish oil supplementation in sows on performance, intestinal microflora, intestinal morphology, volatile fatty acid concentrations and immune status of weaned pigs[J]. *The British Journal of Nutrition*, 2011, 105(4): 549-560.
- [25] LEONARD S G, SWEENEY T, BAHAR B, et al. Effect of maternal seaweed extract supplementation on suckling piglet growth, humoral immunity, selected microflora, and immune response after an ex vivo lipopolysaccharide challenge[J]. *Journal of Animal Science*, 2012, 90(2): 505-514.
- [26] LEONARD S G, SWEENEY T, BAHAR B, et al. Effects of dietary seaweed extract supplementation in sows and post-weaned pigs on performance, intestinal morphology, intestinal microflora and immune status[J]. *British Journal of Nutrition*, 2011, 106(5): 688-699.
- [27] [Chinese reference with garbled characters—skipped]
- [28] TUMMARUK P. Post-parturient disorders and backfat loss in tropical sows in relation to backfat thickness before farrowing and postpartum intravenous supportive treatment[J]. *Asian-Australasian Journal of Animal Sciences*, 2013, 26(2): 171-177.
- [29] STRATHE A V, BRUUN T S, HANSEN C F. Sows with high milk production had both a high feed intake and mobilization[J]. *Animal: An International Journal of Animal Bioscience*, 2017. DOI:10.1017/S1751731117000155.
- [30] KIS R K, BILKEI G. Effect of a phyto-genic feed additive on weaning-to-estrus interval and farrowing rate in sows[J]. *Journal of Swine Health and Production*, 2003, 11(6): 296-299.
- [31] WANG G Y, YANG C W, YANG Z B, et al. Effects of dietary star anise (*Illicium verum* Hook f) supplementation during gestation and lactation on the performance of lactating multiparous sows and nursing piglets[J]. *Animal Science Journal*, 2015, 86(4): 401-407.
- [32] [Chinese reference with garbled characters—skipped]
- [33] LEE S D, KIM J H, JUNG H J, et al. The effect of ginger extracts on the antioxidant capacity and IgG concentrations in the colostrum and plasma of neo-born piglets and sows[J]. *Livestock Science*, 2013, 154(1/2/3): 117-122.
- [34] FARMER C, LAPOINTE J, CORMIER I. Providing the plant extract silymarin to lactating sows: effects on litter performance and oxidative stress in sows[J]. *Animal*, 2017, 11(3): 405-410.

- [35] ARIZA-NIETO C, BANDRICK M, BAIDOO S K, et al. Effect of dietary supplementation of oregano essential oils to sows on colostrum and milk composition, growth pattern and immune status of suckling pigs[J]. *Journal of Animal Science*, 2011, 89(4): 1079-1089.
- [36] ILSLEY S E, MILLER H M, KAMEL H M R. Plant extracts as supplements for lactating sows: effects on piglet performance, sow food intake and diet digestibility[J]. *Animal Science*, 2003, 77(2): 247-254.
- [37] WANG Q, KIM H J, CHO J H, et al. Effects of phytochemical substances on growth performance, digestibility of nutrients, faecal noxious gas content, blood and milk characteristics and reproduction in sows and litter performance[J]. *Canadian Journal of Economics and Political Science*, 2008, 17(1): 362-378.
- [38] MATYSIAK B, JACYNO E, KAWECKA M, et al. The effect of plant extracts fed before farrowing and during lactation on sow and piglet performance[J]. *South African Journal of Animal Science*, 2012, 42(1): 15-21.
- [39] BALASUBRAMANIAN B, PARK J W, KIM I H, et al. Evaluation of the effectiveness of supplementing micro-encapsulated organic acids and essential oils in diets for sows and suckling piglets[J]. *Italian Journal of Animal Science*, 2016, 15(4): 626-633.
- [40] LIU W C, YUN H M, PI S H, et al. Supplementing lactation diets with herbal extract mixture during summer improves performance nursing piglets[J]. *Annals Animal Science*, 2017. DOI:<https://doi.org/10.1515/aoas-2016-0084>
- [41] LIPINIŃSKI K, SKÓRKO-SAJKO H, ANTOSZKIEWICZ Z, et al. A note on the effect of dietary supplementation with herbal extracts on sow and litter performance[J]. *South African Journal of Animal Science*, 2014, 44(2): 110-113.
- [42] SHOJI Y, NAKASHIMA H. Nutraceuticals and delivery systems[J]. *Journal of Drug Targeting*, 2004, 12(6): 385-391.
- [43] SUNTRES Z E, COCCIMIGLIO J, ALIPOUR M. The bioactivity and toxicological actions of carvacrol[J]. *Critical reviews in food science and nutrition*, 2015, 55(3): 304-318.
- [44] [Chinese reference with garbled characters—skipped]
- [45] RUBIOLO J A, MITHIEUX G, VEGA F V. Resveratrol protects primary rat hepatocytes against oxidative stress damage: activation of the Nrf2 transcription factor and augmented activities of antioxidant enzymes[J]. *European Journal of Pharmacology*, 2008, 591(1/2/3): 66-72.
- [46] AZIZ M, KARBOUNE S. Natural antimicrobial/antioxidant agents in meat and poultry products as well as fruits and vegetables: a review[J]. *Critical Reviews in Food Science and Nutrition*, 2016. DOI:10.1080/10408398.2016.1194256
- [47] XIAO Z P, PENG Z Y, PENG M J, et al. Flavonoids health benefits and their molecular mechanism[J]. *Mini Reviews in Medicinal Chemistry*, 2011, 11(2): 169-177.

- [48] RAINS J L, JAIN S K. Oxidative stress, insulin signaling, and diabetes[J]. *Free Radical Biology & Medicine*, 2011, 50(5): 567-575.
- [49] MOSNIER E, LE FLOC' H N, ETIENNE M, et al. Reduced feed intake of lactating primiparous sows is associated with increased insulin resistance during the peripartum period and is not modified through supplementation with dietary tryptophan[J]. *Journal of Animal Science*, 2010, 88(2): 612-625.
- [50] KOREN O, GOODRICH J K, CULLENDER T C, et al. Host remodeling of the gut microbiome and metabolic changes during pregnancy[J]. *Cell*, 2012, 150(3): 470-480.
- [51] KRISHNA G, DIVYASHRI G, PRAPULLA S G, et al. A combination supplement of fructo- and xylo-oligosaccharides significantly abrogates oxidative impairments and neurotoxicity in maternal/fetal milieu following gestational exposure to acrylamide in rat[J]. *Neurochemical Research*, 2015, 40(9): 1904-1918.
- [52] ROOKS M G, GARRETT W S. Gut microbiota, metabolites and host immunity[J]. *Nature Reviews Immunology*, 2016, 16(6): 341-352.
- [53] JONES R M, MERCANTE J W, NEISH A S. Reactive oxygen production induced by the gut microbiota: pharmacotherapeutic implications[J]. *Current Medicinal Chemistry*, 2012, 19(10): 1519-1529.
- [54] MARTÍN R, SUÁREZ J E. Biosynthesis and degradation of H<sub>2</sub>O<sub>2</sub> by vaginal lactobacilli[J]. *Applied and Environmental Microbiology*, 2010, 76(2): 400-405.
- [55] ALVAREZ L A, KOVAČIČ L, RODRÍGUEZ J, et al. NADPH oxidase-derived H<sub>2</sub>O<sub>2</sub> subverts pathogen signaling by oxidative phosphotyrosine conversion to PB-DOPA[J]. *Proceedings of the National Academy of Sciences of the United States of America*, 2016, 113(37): 10406-10411.
- [56] ZHANG J C, XU C B, HUO D X, et al. Comparative study of the gut microbiome potentially related to milk protein in Murrah buffaloes (*Bubalus bubalis*) and Chinese Holstein cattle[J]. *Scientific Reports*, 2017, 7: 42189.
- [57] KNUDSEN K E B, HEDEMANN M S, LÆRKE H N. The role of carbohydrates in intestinal health of pigs[J]. *Animal Feed Science and Technology*, 2012, 173(1/2): 41-53.
- [58] BOMA M H, BILKEI G. Gross pathological findings in sows of different parity, culled due to recurring swine urogenital disease (SUGD) in Kenya[J]. *Onderstepoort Journal of Veterinary Research*, 2006, 73(2): 139-142.
- [59] CUSHNIE T P T, CUSHNIE B, LAMB A J. Alkaloids: an overview of their antibacterial, antibiotic-enhancing and antivirulence activities[J]. *International Journal of Antimicrobial Agents*, 2014, 44(5): 377-386.
- [60] ZOU Y, XIANG Q, WANG J, et al. Oregano essential oil improves intestinal morphology and expression of tight junction proteins associated with

modulation of selected intestinal bacteria and immune status in a pig model[J]. BioMed Research International, 2016, 2016: 5436738.

[61] MUSHTAQ S, RATHER M A, QAZI P H, et al. Isolation and characterization of three benzyloisoquinoline alkaloids from *Thalictrum minus* L. and their antibacterial activity against bovine mastitis[J]. Journal of Ethnopharmacology, 2016, 193: 221-226.

[62] LI C Y, MENG Y H, YING Z M, et al. Three novel alkaloids from *Portulaca oleracea* L. and their anti-inflammatory effects[J]. Journal of Agricultural and Food Chemistry, 2016, 64(29): 5837-5844.

[63] TRAN H T T, MÁRTON M R, HERZ C, et al. Nasturtium (Indian cress, *Tropaeolum majus* nanum) dually blocks the COX and LOX pathway in primary human immune cells[J]. Phytomedicine, 2016, 23(6): 611-620.

[64] ZHAO X, LACASSE P. Mammary tissue damage during bovine mastitis: causes and control[J]. Journal of Animal Science, 2008, 86(13S): 57-65.

[65] [Chinese reference with garbled characters—skipped]

[66] CHAUHAN P S, SATTI N K, SURI K A, et al. Stimulatory effects of *Cuminum cyminum* and flavonoid glycoside on Cyclosporine-a and restraint stress induced immune-suppression in Swiss albino mice[J]. Chemico-Biological Interactions, 2010, 185(1): 66-72.

*Note: Figure translations are in progress. See original paper for figures.*

*Source: ChinaXiv –Machine translation. Verify with original.*