

Advances in Research on the Biological Activity and Mechanism of Action of Chlorogenic Acid (Postprint)

Authors: Wang Wenlong, Wen Chaoyue, Guo Qiuping, Duan Yehui, Li Yinghui, He Shanping, Li Fengna

Date: 2017-10-10T00:00:00+00:00

Abstract

Chlorogenic acid (CGA) is a widely distributed polyphenolic compound possessing multiple biological functions, including antioxidant, anti-inflammatory, antimicrobial, and hypoglycemic and hypolipidemic activities, and has broad application prospects in animal production and human health. This article briefly reviews the sources and distribution, physicochemical properties, biological activities, and main mechanisms of action of CGA, providing a reference basis for in-depth research and further development and application of CGA.

Full Text

Advances in Research on the Biological Activities and Mechanisms of Action of Chlorogenic Acid

WANG Wenlong^{1,2,3}, WEN Chaoyue^{1,2}, GUO Qiuping^{2,3}, DUAN Yehui^{2,3}, LI Yinghui^{2,3}, HE Shanping¹, LI Fengna^{2,*}

¹Laboratory of Animal Nutrition and Human Health, School of Life Sciences, Hunan Normal University, Changsha 410006, China

²Key Laboratory of Agro-Ecological Processes in Subtropical Region, National Engineering Laboratory for Pollution Control and Waste Utilization in Livestock and Poultry Production, Institute of Subtropical Agriculture, Chinese Academy of Sciences, Changsha 410125, China

³University of Chinese Academy of Sciences, Beijing 100049, China
Hunan Co-Innovation Center of Animal Production Safety, CICAPS, Changsha 410128, China

Abstract: Chlorogenic acid (CGA) is a widely distributed polyphenolic compound with diverse biological functions including antioxidant, anti-

inflammatory, antibacterial, hypoglycemic, and hypolipidemic activities. These properties give it broad application prospects in animal production and human health. This paper briefly reviews the sources, distribution, physicochemical properties, biological activities, and primary mechanisms of action of CGA to provide a reference basis for further research and development.

Keywords: chlorogenic acid; polyphenol; antioxidant; lipid metabolism; mechanism of action; animal product quality

Chlorogenic acid (CGA) is a phenylpropanoid compound produced through the shikimic acid pathway during aerobic respiration in plants and has attracted considerable scholarly attention since its first successful extraction from apples in the 1950s. CGA has been widely applied in food, cosmetics, pharmaceutical, and chemical industries due to its antioxidant, anti-inflammatory, antibacterial, and anti-radiation properties. Recent studies have also revealed its positive roles in anticancer activity, glucose and lipid regulation, immune modulation, and central nervous system stimulation, though the specific mechanisms remain unclear. Against the backdrop of renewed research enthusiasm for natural plant extracts like artemisinin, coupled with growing public concerns about antibiotics and food safety and increasing demand for high-quality animal products, the development and application of CGA has become a research hotspot. This review summarizes the sources, distribution, physicochemical properties, and biological activities of CGA, discusses its primary mechanisms of action, and explores its application prospects in animal production and human health.

1. Main Sources and Distribution of CGA

CGA is widely distributed throughout the plant kingdom, ranging from dicotyledons to ferns, and is primarily found in plants from the Caprifoliaceae (*Lonicera* L.) and Asteraceae (*Artemisia* L.) families, including *Eucommia ulmoides*, honeysuckle (*Lonicera japonica*), sunflower, wild chrysanthemum, capillary artemisia, and burdock. It is also present in coffee, apples, carrots, green tea, wheat, soybeans, potatoes, and kuding tea. Honeysuckle and *Eucommia ulmoides* contain particularly high CGA concentrations. The content of CGA varies depending on geographical origin, soil conditions, climate, developmental stage, genotype, and plant organ.

China possesses abundant CGA resources, providing a strong foundation for its development and utilization. Honeysuckle, one of the most important CGA sources, is produced in most regions of China, with major production areas including Pingyi in Shandong Province and Longhui in Hunan Province, both designated as “Hometown of Chinese Honeysuckle.” Pingyi represents the traditional source and main production area for *Lonicera japonica*, while Longhui has traditionally cultivated *Lonicera macranthoides* since the Qing Dynasty, as recorded in the *Xinhua County Annals* of 1871. Originally a wild species distributed across approximately 5,000 mu (1 mu = 666.67 m²), *Lonicera macranthoides* was

domesticated in the early 1960s with systematic cultivation techniques. The CGA content in *L. macranthoides* flower buds is significantly higher than in traditional honeysuckle varieties, reaching over 4% in Longhui and Xinning counties. The current cultivation area exceeds 200,000 mu with annual production of over 10,000 tons (dry flowers), accounting for more than 50% of national output and ranking first in China. Additionally, *Eucommia ulmoides*, another important CGA source, is abundant in China. As a species endemic to central and southwestern China, Hunan represents one of its central production areas. Cili County in Zhangjiajie, home to China's largest wild *Eucommia* forest base, was renowned as "China's *Eucommia* Hometown at the Mysterious 30°N Latitude" as early as 1996, with preserved areas exceeding 150,000 mu. Therefore, investigating the biological activities of CGA and maximizing its application potential in animal production and human health is crucial for fully utilizing China's natural resources and improving food safety and public health.

2. Physicochemical Properties of CGA

CGA, also known as caffeotannic acid or caffeoylquinic acid, is a depside composed of caffeic acid and quinic acid formed through the shikimic acid pathway during plant aerobic respiration. Based on the esterification position on quinic acid, CGA can be classified into 3-*O*-caffeoylquinic acid (chlorogenic acid), 4-*O*-caffeoylquinic acid (cryptochlorogenic acid), 5-*O*-caffeoylquinic acid (neochlorogenic acid), 3,4-dicaffeoylquinic acid (isochlorogenic acid B), 3,5-dicaffeoylquinic acid (isochlorogenic acid A), and 4,5-dicaffeoylquinic acid (isochlorogenic acid C).

The molecular formula of CGA is $C_{16}H_{16}O_9$ with a relative molecular mass of 354.30. Its chemical structure is shown in [Figure 1: see original paper]. The hemihydrate forms white or yellow needle-like crystals that become anhydrous at 110°C, with a melting point of 206–208°C. CGA is widely distributed in traditional Chinese medicinal herbs and foods. Natural CGA in plants often coexists with its isomers, so plant-extracted CGA is typically a mixture, with 5-*O*-caffeoylquinic acid (neochlorogenic acid) being the main component. At room temperature, CGA has low water solubility (approximately 4%) that increases in hot water; it is readily soluble in ethanol, acetone, and methanol, slightly soluble in ethyl acetate, and poorly soluble in lipophilic organic solvents such as chloroform, ether, and benzene. As a polar organic acid, CGA is relatively unstable and prone to isomerization during extraction. The ortho-dihydroxyphenyl group in its molecular structure serves as an optimal substrate for phenolase catalysis and is susceptible to oxidation when exposed to heat or light.

3.1 Antioxidant Activity

Studies have demonstrated that CGA is an effective phenolic antioxidant with stronger antioxidant capacity than common antioxidants such as caffeic acid, *p*-hydroxybenzoic acid, ferulic acid, syringic acid, vitamin C, and vitamin E. The

natural antioxidant properties of CGA depend on its unique molecular structure, which contains five active hydroxyl groups and one carboxyl group that can donate hydrogen radicals to scavenge reactive oxygen species and hydroxyl radicals. The ortho-dihydroxyphenyl group is unstable and readily oxidized, thereby protecting tissues from oxidative damage.

Furthermore, CGA has been reported to inhibit xanthine oxidase (XO) activity and reduce oxygen radical production *in vivo*, while also upregulating antioxidant enzyme activity and decreasing lipid peroxidation levels. The potential mechanisms of CGA's antioxidant activity ([Figure 2: see original paper]) include: (1) promoting amino acid metabolism and glutathione metabolic pathways to improve lipid metabolism; (2) upregulating the nuclear factor erythroid 2-related factor 2/antioxidant response element (Nrf2-ARE) signaling pathway by inhibiting ubiquitin-mediated degradation of Nrf2 protein, stabilizing cytoplasmic Nrf2 concentration, enhancing Nrf2 transcriptional activity under stress conditions, and promoting expression of protective genes such as antioxidant proteins and phase II detoxification enzymes; (3) upregulating the phosphatidylinositol 3-kinase/protein kinase B (PI3K-Akt) signaling pathway to promote Akt phosphorylation and expression of forkhead box O transcription factor (FoxO), tumor suppressor gene p53, and anti-apoptotic protein B-cell leukemia/lymphoma 2 (Bcl-2), thereby inhibiting apoptosis; and (4) modulating the mitogen-activated protein kinase (MAPK) signaling pathway by inhibiting phosphorylation of extracellular signal-regulated protein kinase 1/2 (ERK1/2), c-jun N-terminal kinase (JNK), and p38 MAPK.

Abbreviations: CGA, chlorogenic acid; Nrf2-ARE, nuclear factor erythroid 2-related factor 2/antioxidant response element pathway; PI3K-Akt, phosphatidylinositol 3-kinase/protein kinase B pathway; MAPK, mitogen-activated protein kinases pathway; Nrf2, nuclear factor erythroid 2-related factor 2; p-Akt, phosphorylated protein kinase B; FoxO, forkhead box O transcription factor; p53, tumor suppressor gene p53; Bcl-2, B-cell leukemia/lymphoma 2; p-p38, phosphorylated p38 mitogen-activated protein kinase; p-JNK, phosphorylated c-jun N-terminal kinase; p-ERK, phosphorylated extracellular signal-regulated protein kinase.

3.2 Anti-inflammatory Activity

Anti-inflammatory activity represents another important biological function of CGA and a key characteristic for its widespread application beyond antioxidant properties. CGA can participate in the Janus kinase/signal transducer and activator of transcription 3 (JAK-STAT3) signaling pathway by inhibiting expression of interleukin-6 (IL-6) receptor subunit (gp130), Janus kinase 1 (JAK1), and phosphorylated STAT3 (p-STAT3) under oxidative stress conditions, thereby negatively regulating inflammatory factor expression and secretion. Inflammatory responses are crucial manifestations of autoimmunity, and the nuclear factor kappa B (NF- κ B) pathway is closely associated with autoimmunity, playing a key regulatory role in secretion of pro-inflammatory

cytokines, chemokines, and adhesion molecules. Studies have shown that CGA can inhibit carbon tetrachloride (CCl₄)- and lipopolysaccharide (LPS)-induced expression of interleukin-1 (IL-1), IL-6, and tumor necrosis factor- α (TNF- α) by suppressing NF- κ B, and can also alleviate inflammation by inhibiting NF- κ B activation. Additionally, Toll-like receptor (TLR) signaling pathways represent important regulatory mechanisms for CGA's anti-inflammatory effects. CGA can inhibit TLR4 pathway activation, downregulate expression of TLR4, myeloid differentiation factor 88 (MyD88), inducible nitric oxide synthase (iNOS), and cyclooxygenase-2 (COX-2), and upregulate BMP and activin membrane-bound inhibitor (BAMBI) expression, thereby inhibiting secretion of TNF- α , IL-6, and IL-1 and alleviating hepatic inflammatory injury and fibrosis. Subsequent studies have found that CGA can also improve inflammatory responses by inhibiting TLR2, TLR3, and TLR9 signaling pathways.

3.3 Antimicrobial and Antiviral Activity

CGA is one of the primary active antibacterial components in traditional Chinese medicinal herbs such as honeysuckle and *Eucommia ulmoides*. Research demonstrates that CGA exhibits broad-spectrum antimicrobial activity with strong inhibitory and bactericidal effects against various pathogenic bacteria and fungi, showing greater efficacy against fungi than bacteria with dose-dependent effects. The antimicrobial mechanisms of purified CGA compounds remain unclear, and systematic studies are scarce. Based on available literature, potential mechanisms may include: (1) CGA's phenolic hydroxyl groups serve as optimal substrates for phenolase catalysis, affecting metabolic enzyme activity and reducing material and energy metabolism levels, thereby blocking metabolic processes and inhibiting bacterial activity. Evidence suggests CGA may be an effective non-competitive inhibitor of arylamine acetyltransferase in gastrointestinal bacteria such as *Escherichia coli*, affecting bacterial metabolism and inhibiting sugar metabolism to cause energy deficiency and impair growth and reproduction. (2) The strong polarity of CGA molecules enables high affinity for macromolecules like lipids, allowing binding to bacterial surfaces and altering membrane structure, increasing cell membrane permeability, and causing leakage of DNA, RNA, electrolytes, enzymes, and nutrients, thereby affecting protein synthesis. (3) Beyond biofilm stability, bacterial drug resistance and infectivity are closely related to swarming ability. CGA may exert antimicrobial effects by inhibiting bacterial flagella synthesis, reducing flagella numbers, and decreasing swarming motility.

Research on CGA's antiviral effects is relatively limited, and the mechanisms remain unclear. Reports indicate that CGA significantly blocks and inhibits influenza A virus FM1 strain, herpes simplex virus (HSV), porcine parvovirus (PPV) in vitro, and porcine reproductive and respiratory syndrome virus (PRRSV). The mechanism may involve inhibiting expression of certain proteins required for virus-host binding, thereby preventing viral entry and release of progeny virions, though further investigation is needed. Additionally,

chlorogenic acid ethyl ester exhibits auxin-like activity, suggesting CGA may serve as a potential lead compound for anti-HIV drug development.

3.4 Hypoglycemic and Hypolipidemic Effects

Metabolic diseases such as type II diabetes, triggered by obesity, have become major threats to human health with rapidly increasing prevalence. Reducing excessive fat deposition represents an important strategy for addressing these issues, while regulating fat deposition also improves meat quality in animal production. CGA has been identified as the first novel specific inhibitor of glucose-6-phosphate translocase in rat liver microsomes. Since this enzyme plays a crucial role in endogenous glucose formation through gluconeogenesis and glycogenolysis, CGA intake may help reduce excessive hepatic glucose output in type II diabetes, making it a potential therapeutic option. CGA can also reduce fat deposition and decrease lipid content in blood and liver by inhibiting expression of peroxisome proliferator-activated receptor-2 (PPAR-2) and CCAAT enhancer binding protein (C/EBP), which are transcription factors associated with adipocyte differentiation. Moreover, CGA participates in the AMP-activated protein kinase (AMPK) signaling pathway, which plays vital biological roles in metabolism and cell development. AMPK serves as a key sensor of cellular energy status and a major regulator of lipid homeostasis in the liver and whole body, representing an important target for diabetes and related metabolic diseases.

Studies have shown that CGA can activate AMPK, upregulate glucose transporter 4 (GLUT4) expression, stimulate glucose uptake in skeletal muscle, downregulate glucose-6-phosphate translocase expression, inhibit gluconeogenesis, and suppress fatty acid synthesis. CGA has also been demonstrated to reduce fatty acid synthesis and decrease fat deposition in animals by downregulating expression of aconitase (ACO), catalase (CAT), fatty acid synthetase (FAS), and peroxisome proliferator-activated receptor-2 (PPAR-2). Pancreatic lipase (PL), secreted by the pancreas, is responsible for 50–70% of dietary fat digestion. CGA and other plant extracts are natural PL inhibitors that can be used for obesity and diabetes intervention.

Furthermore, CGA's hypoglycemic effects have been confirmed, with potential mechanisms including: (1) stimulating glucagon-like peptide-1 (GLP-1)-mediated insulin secretion, and (2) activating AMPK to promote GLUT4 translocation to the cell membrane, thereby enhancing glucose uptake.

4.1 Broad Application Prospects of CGA in Animal Production

As a widely sourced natural plant extract with multiple biological functions and no pollution or toxic side effects, CGA can improve intestinal microflora structure and redox balance, alleviate adverse effects of various stress factors during animal production, regulate fat deposition, and enhance immunity and

disease resistance. It represents an ideal feed additive to replace antibiotics and hormones, holding significant importance for reducing antibiotic use and improving production efficiency and product quality. Adding 300 mg/kg CGA (85% purity) to weaned piglet diets significantly increases plasma glutathione peroxidase (GSH-Px) and CAT activities, reduces malondialdehyde (MDA) content, enhances plasma hydroxyl radical scavenging capacity, and improves antioxidant capacity to alleviate oxidative stress damage, likely due to CGA's phenolic hydroxyl groups forming stable semiquinone structures with free radicals. Similar results were obtained in sow studies, where 300 mg/kg dietary CGA (90% purity) improved antioxidant capacity in sows and newborn piglets, significantly increased piglet birth weight, litter size, and 21-day litter weight, and reduced sow backfat loss during lactation, effectively alleviating farrowing stress and improving sow health and production efficiency. Adding 0.08% *Eucommia* leaf polyphenol extract (containing 33.7% CGA) to finishing pig diets significantly increased final body weight and average daily gain, reduced feed-to-gain ratio, improved growth performance, enhanced dietary protein utilization, regulated lipid metabolism, and improved meat quality. Beyond swine production, studies have shown that dietary CGA supplementation can improve growth performance, alleviate stress, enhance immunity, and improve product quality in poultry and aquatic animals, providing a solid foundation for CGA application in animal production.

4.2 Promising Breakthroughs for CGA in Human Health

Numerous studies and applications have demonstrated CGA's free radical scavenging, antibacterial, and anti-inflammatory effects. According to incomplete statistics, over 170 traditional Chinese patent medicines recorded in the Ministry of Health's Drug Standards for heat-clearing, detoxifying, antibacterial, and anti-inflammatory purposes contain CGA as a major component, making it an important quality control indicator in products such as Shuanghuanglian preparations. CGA is also widely used as a primary active ingredient in preservatives, cosmetics, and daily chemical products. With advancing research and medical technology, more application values of CGA are being discovered, and future breakthroughs are anticipated in curbing the spread of nutritional metabolic diseases (obesity, diabetes), treating immunodeficiency diseases (HIV), and enhancing natural immunity.

In summary, CGA possesses multiple biological functions with broad application foundations in animal production and human health. However, several issues currently limit its deeper development and utilization: (1) immature preparation technology and high costs for isolating pure CGA from plants; (2) unclear metabolic pathways and low bioavailability requiring improvement; (3) unclear mechanisms of action; and (4) limited application in animal production with need for further research on optimal utilization. Therefore, systematic investigation of CGA biosynthesis methods and elucidation of its metabolic pathways and molecular mechanisms are urgently needed to maximize the development

potential of CGA-rich plant resources.

References

- PCHELKIN V P. Natural phenolic and lipophilic complexes of chlorogenic acid[J]. Pharmaceutical Chemistry Journal, 2003, 37(1): 25-27.
- BASSOLI B K, CASSOLLA P, BORBA-MURAD G R, et al. Chlorogenic acid reduces the plasma glucose peak in the oral glucose tolerance test: effects on hepatic glucose release and glycaemia[J]. Cell Biochemistry and Function, 2008, 26(3): 320-328.
- DOS SANTOS M D, ALMEIDA M C, LOPES N P, et al. Evaluation of anti-inflammatory, analgesic and antipyretic activities of the natural polyphenol chlorogenic acid[J]. Biological and Pharmaceutical Bulletin, 2006, 29(11): 2236-2240.
- DE SOTILLO D V R, HADLEY M, SOTILLO J E. Insulin receptor exon 11+/- is expressed in Zucker (fa/fa) rats, and chlorogenic acid modifies their plasma insulin and liver protein and DNA[J]. The Journal of Nutritional Biochemistry, 2006, 17(1): 63-71.
- NICASIO P, AGUILAR-SANTAMARÍA L, ARANDA E, et al. Hypoglycemic effect and chlorogenic content of *Cecropia* species[J]. Phytotherapy Research, 2005, 19(8): 661-664.
- DAAYF F, LATTANZIO V. Recent advances in polyphenol research Vol. 1[M]. New Jersey: Wiley-Blackwell, 2009: 311-351.
- KIMURA Y, OKUDA H, OKUDA T, et al. Studies on the activities of tannins and related compounds from medicinal plants and drugs. VII. Effects of extracts of leaves of *Artemisia* species, and caffeic acid and chlorogenic acid on lipid metabolic injury in rats fed peroxidized oil[J]. Chemical and Pharmaceutical Bulletin, 1985, 33(5): 2028-2034.
- 周荣汉. 中药资源学 [M]. 北京: 中国医药科技出版社, 1993: 240.
- MATTILA P, HELLSTRÖM J. Phenolic acids in potatoes, vegetables, and some of their products[J]. Journal of Food Composition and Analysis, 2007, 20(3/4): 152-160.
- 周志娥, 罗秋水, 熊建华, 等. 绿原酸、异绿原酸 A 对大肠杆菌的抑菌机制 [J]. 食品科技, 2014, 39(3): 228-232.
- 李淑媛, 常翠青. 氯原酸的生物活性与人体健康 [J]. 卫生研究, 2005, 34(6): 762-764.
- CLIFFORD M N. Chlorogenic acids and other cinnamates—nature, occurrence, dietary burden, absorption and metabolism[J]. Journal of the Science of Food and Agriculture, 2000, 80(7): 1033-1043.
- DORRELL D G. Chlorogenic acid content of meal from cultivated and wild sunflowers[J]. Crop Science, 1976, 16(3): 422-424.

宁正祥. 食品生物化学 [M]. 2 版. 广州: 华南理工大学出版社, 2006: 356.

JI L L, JIANG P, LU B, et al. Chlorogenic acid, a dietary polyphenol, protects acetaminophen-induced liver injury and its mechanism[J]. The Journal of Nutritional Biochemistry, 2013, 24(11): 1911-1919.

FENG R T, LU Y J, BOWMAN L L, et al. Inhibition of activator protein-1, NF- κ B, and MAPKs and induction of phase 2 detoxifying enzyme activity by chlorogenic acid[J]. Journal of Biological Chemistry, 2005, 280(30): 27888-27895.

李文娜, 肖苑, 陈阳, 等. 杜仲叶绿原酸提取物与绿原酸、维生素 C 体外抗氧化比较 [J]. 食品工业科技, 2012, 33(11): 137-140.

BOETTLER U, VOLZ N, PAHLKE G, et al. Coffees rich in chlorogenic acid or N-methylpyridinium induce chemopreventive phase II-enzymes via the Nrf2/ARE pathway in vitro and in vivo[J]. Molecular Nutrition & Food Research, 2011, 55(5): 798-802.

LOU L X, ZHOU J W, LIU Y J, et al. Chlorogenic acid induces apoptosis to inhibit inflammatory proliferation of IL-6-induced fibroblast-like synoviocytes through modulating the activation of JAK/STAT and NF- κ B signaling pathways[J]. Experimental and Therapeutic Medicine, 2016, 11(5): 2054-2060.

LAWRENCE T. The nuclear factor NF- κ B pathway in inflammation[J]. Cold Spring Harbor Perspectives in Biology, 2009, 1(6): a001651.

SHAN J H, FU J, ZHAO Z H, et al. Chlorogenic acid inhibits lipopolysaccharide-induced cyclooxygenase-2 expression in RAW264.7 cells through suppressing NF- κ B and JNK/AP-1 activation[J]. International Immunopharmacology, 2009, 9(9): 1042-1048.

HWANG S J, KIM Y W, PARK Y, et al. Anti-inflammatory effects of chlorogenic acid in lipopolysaccharide-stimulated cells[J]. Inflammation Research, 2014, 63(1): 81-90.

YOON T, CHEON M S, LEE A Y, et al. Anti-inflammatory activity of methylene chloride fraction from *Glehnia littoralis* extract via suppression of NF- κ B and mitogen-activated protein kinase activity[J]. Journal of Pharmacological Sciences, 2010, 112(1): 46-55.

MITJAVILA M T, MORENO J J. The effects of polyphenols on oxidative stress and the arachidonic acid cascade. Implications for the prevention/treatment of high prevalence diseases[J]. Biochemical Pharmacology, 2012, 84(9): 1113-1122.

SHI H T, DONG L, JIANG J, et al. Chlorogenic acid reduces liver inflammation and fibrosis through inhibition of toll-like receptor signaling pathway[J]. Toxicology, 2013, 303: 107-114.

GAO R F, FU Y H, WEI Z K, et al. Chlorogenic acid attenuates lipopolysaccharide-induced mice mastitis by suppressing TLR4-mediated

NF- B signaling pathway[J]. *European Journal of Pharmacology*, 2014, 729: 54-58.

GUO Y J, LUO T, WU F, et al. Involvement of TLR2 and TLR9 in the anti-inflammatory effects of chlorogenic acid in HSV-1-infected microglia[J]. *Life Sciences*, 2015, 127: 12-18.

ZHENG Z Y, SHENG Y C, LU B, et al. The therapeutic detoxification of chlorogenic acid against acetaminophen-induced liver injury ameliorating hepatic inflammation[J]. *Chemico-Biological Interactions*, 2015, 238: 93-101.

ZHU X F, ZHANG H X, LO R. Phenolic compounds from the leaf extract of artichoke (*Cynara scolymus* L.) and their antimicrobial activities[J]. *Journal of Agricultural and Food Chemistry*, 2004, 52(24): 7272-7278.

KONO Y, KOBAYASHI K, TAGAWA S, et al. Antioxidant activity of polyphenolics in diets: rate constants of reactions of chlorogenic acid and caffeic acid with reactive species of oxygen and nitrogen[J]. *Biochimica et Biophysica Acta (BBA) - General Subjects*, 1997, 1335(3): 335-342.

LO H H, CHUNG J G. The effects of plant phenolics, caffeic acid, chlorogenic acid and ferulic acid on arylamine N-acetyltransferase activities in human gastrointestinal microflora[J]. *Anticancer Research*, 1999, 19(1A): 133-139.

罗艺晨, 黄利明, 杨颖, 等. 绿原酸抑制金黄色葡萄球菌机理研究 [J]. *西南大学学报: 自然科学版*, 2016, 38(3): 15-19.

FRANCISCO V, COSTA G, FIGUEIRINHA A, et al. Anti-inflammatory activity of *Cymbopogon citratus* leaves infusion via proteasome and nuclear factor- B pathway inhibition: contribution of chlorogenic acid[J]. *Journal of Ethnopharmacology*, 2013, 148(1): 126-134.

LOU Z X, WANG H X, ZHU S, et al. Antibacterial activity and mechanism of action of chlorogenic acid[J]. *Journal of Food Science*, 2011, 76(6): M398-M403.

REN S, WU M, GUO J Y, et al. Sterilization of polydimethylsiloxane surface with Chinese extract: antibiotic mechanism of chlorogenic acid[J]. *Scientific Reports*, 2015, 5: 10464.

OJHA D, MUKHERJEE H, GHOSH S, et al. Evaluation of anti-infective potential of a tribal folklore *Odina wodier* Roxb against some selected microbes and herpes simplex virus associated with skin infection[J]. *Journal of Applied Microbiology*, 2013, 115(6): 1317-1328.

潘翌翌, 王雪峰, 闫丽娟, 等. 金银花提取物体外抗甲型流感病毒 FM1 株的研究 [J]. *中国中医药信息杂志*, 2007, 14(6): 37-38, 51.

王学兵, 崔保安, 魏战勇, 等. 绿原酸对猪繁殖与呼吸综合征病毒体外作用的研究 [J]. *中国农业科技导报*, 2008, 10(3): 107-110.

王学兵, 魏战勇, 崔保安, 等. 绿原酸的提取及其对猪细小病毒的体外作用研究 [J]. *中国畜牧兽医*, 2008, 35(12): 123-125.

相婷, 吴立军, 董梅, 等. 西南忍冬化学成分的研究 () [J]. 中国药物化学杂志, 1998, 8(1): 44-45, 48.

SCHINDLER P W, BELOW P, HEMMERLE H, et al. Identification of two new inhibitors of hepatic glucose-6-phosphatase system [J]. Drug Development Research, 1998, 44(1): 34-40.

邱阳阳. 咖啡碱和绿原酸对小鼠和 3T3-L1 细胞脂肪代谢的影响 [D]. 硕士学位论文. 南昌: 江西农业大学, 2013: 43-56.

吴铁梅, 闫素梅, 格日乐玛. 脂肪细胞因子对动物脂类代谢的调控机理 [J]. 动物营养学报, 2016, 28(10): 3034-3041.

ONG K W, HSU A, TAN B K H. Anti-diabetic and anti-lipidemic effects of chlorogenic acid mediated activation [J]. Biochemical Pharmacology, 2013, 85(9): 1341-1351.

ZHENG G D, QIU Y Y, ZHANG Q F, et al. Chlorogenic acid and caffeine in combination inhibit fat accumulation by regulating hepatic lipid metabolism-related enzymes in mice [J]. British Journal of Nutrition, 2014, 112(6): 1034-1040.

BIRARI R B, BHUTANI K K. Pancreatic lipase inhibitors from natural sources: unexplored potential [J]. Drug Discovery Today, 2007, 12(19/20): 879-889.

DE LA GARZA A L, MILAGRO F I, BOQUE N, et al. Natural inhibitors of pancreatic lipase as new players in obesity treatment [J]. Planta Medica, 2011, 77(8): 773-785.

JOHNSTON K L, CLIFFORD M N, MORGAN L M. Coffee acutely modifies gastrointestinal hormone secretion and glucose tolerance in humans: glycemic effects of chlorogenic acid and caffeine [J]. The American Journal of Clinical Nutrition, 2003, 78(4): 728-733.

JUNG U J, LEE M K, PARK Y B, et al. Antihyperglycemic and antioxidant properties of caffeic acid in db/db mice [J]. Journal of Pharmacology and Experimental Therapeutics, 2006, 318(2): 476-483.

ONG K W, HSU A, TAN B K H. Chlorogenic acid stimulates glucose transport in skeletal muscle via AMPK activation: a contributor to the beneficial effects of coffee on diabetes [J]. PLoS One, 2012, 7(3): e32718.

刘英, 王之盛, 周安国. 橙皮苷、绿原酸对断奶仔猪生长、抗氧化和免疫机能的影响 [J]. 中国兽医学报, 2009, 29(9): 1233-1236.

黄少文, 魏金涛, 赵娜, 等. 绿原酸和维生素 E 对母猪繁殖和抗氧化性能的影响 [J]. 中国畜牧杂志, 2015, 51(24): 79-83.

周艳. 杜仲叶多酚提取物对猪肉品质及绿原酸缓解肝-肠损伤研究 [D]. 博士学位论文. 南昌: 南昌大学, 2015: 17-34.

RUAN Z, LIU S Q, ZHOU Y, et al. Chlorogenic acid decreases intestinal permeability and increases expression of intestinal tight junction proteins in weaned

rats challenged with LPS[J]. PLoS One, 2014, 9(6): e97815.

陈玉敏, 黄涛, 宋小珍, 等. 饲料中添加杜仲叶提取物对爱拔益加肉鸡生长性能及免疫功能的影响 [J]. 动物营养学报, 2015, 27(7): 2224-2230.

李乃顺, 冷向军, 李小勤, 等. 绿原酸对草鱼鱼种生长、非特异性免疫和肉质的影响 [J]. 水生生物学报, 2014, 38(4): 619-626.

温安祥, 舒辉, 肖洋. 绿原酸对中华鳖生产性能及抗氧化能力的影响 [J]. 动物营养学报, 2010, 22(3): 729-733.

LIANG N J, KITTTS D D. Role of chlorogenic acids in controlling oxidative and inflammatory stress conditions[J]. Nutrients, 2015, 8(1): 16.

Note: Figure translations are in progress. See original paper for figures.

Source: ChinaXiv –Machine translation. Verify with original.