

Effects of Oligochitosan on Reproductive Performance, Lactation Performance, and Plasma Biochemical Indices in Sows: Postprint

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Abstract

This experiment aimed to investigate the effects of dietary chitosan oligosaccharide supplementation on reproductive performance, lactation performance, and plasma biochemical indices in sows. Twenty-four healthy Large White sows with similar parity, body weight, and expected farrowing date were selected and randomly allocated to 3 groups with 8 replicates per group and 1 sow per replicate. The control group was fed a basal diet, while Groups I and II were fed the basal diet supplemented with 50 and 100 g/t chitosan oligosaccharide, respectively. The experiment commenced on day 85 of gestation and concluded on day 21 of lactation. The results showed: 1) Compared with the control group, dietary chitosan oligosaccharide supplementation significantly increased the number of healthy piglets and average daily feed intake during lactation ($P < 0.05$), significantly reduced the duration of farrowing ($P < 0.05$), and tended to improve sow constipation and decrease backfat thickness loss. Compared with the control group, Group I exhibited a significant increase in litter birth weight ($P < 0.05$), while Group II showed a significant increase in milk yield during lactation ($P < 0.05$). 2) Compared with the control group, Groups I and II demonstrated significantly increased milk fat content in colostrum ($P < 0.05$), while Group I showed significantly increased urea nitrogen (UN) content in colostrum and arginine, total amino acids, and limiting amino acids contents in mature milk ($P < 0.05$). 3) The plasma glutamic-pyruvic transaminase (GPT) activity in Groups I and II was significantly lower than that in the control group ($P < 0.05$), while no significant differences were observed among groups in plasma alkaline phosphatase (ALP) and lactate dehydrogenase (LDH) activities or in total protein (TP), albumin (ALB), UN, triglyceride (TG), total cholesterol (CHO), high-density lipoprotein (HDL), and low-density lipoprotein (LDL) contents ($P > 0.05$). In conclusion, dietary chitosan oligosaccharide supplementation exerted beneficial effects on both reproductive and lactation performance

in sows. Based on the comprehensive results of this study, 50 g/t chitosan oligosaccharide represents the optimal supplementation level in sow diets.

Full Text

Effects of Oligo-chitosan on Reproduction Performance, Lactation Performance and Plasma Biochemical Parameters of Sows

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Abstract

This study was conducted to investigate the effects of dietary oligo-chitosan on reproduction performance, lactation performance, and plasma biochemical parameters of sows. Twenty-four healthy Yorkshire sows with similar parity, body weight, and expected farrowing date were randomly assigned to three groups, with eight replicates per group and one sow per replicate. Sows in the control group were fed a basal diet, while those in experimental groups I and II received the basal diet supplemented with 50 and 100 g/t oligo-chitosan, respectively. The trial period spanned from day 85 of gestation to day 21 postpartum.

The results showed that: 1) Compared with the control group, dietary oligo-chitosan supplementation significantly increased the number of healthy piglets per litter and the average daily feed intake (ADFI) during lactation ($P < 0.05$), while significantly reducing farrowing duration ($P < 0.05$). Additionally, oligo-chitosan tended to improve constipation and decrease backfat thickness loss in sows. Specifically, the birth litter weight in experimental group I was significantly higher than that in the control group ($P < 0.05$), and the total milk yield during lactation in experimental group II was significantly increased ($P < 0.05$). 2) The milk fat content in colostrum was significantly elevated in both experimental groups I and II compared to the control group ($P < 0.05$). Furthermore, experimental group I showed significantly increased urea nitrogen (UN) content in colostrum and significantly higher concentrations of arginine, total amino acids, and limiting amino acids in milk ($P < 0.05$). 3) Plasma glutamic-pyruvic transaminase (GPT) activity was significantly lower in both experimental groups I and II relative to the control group ($P < 0.05$). However, no

significant differences were observed among groups in plasma alkaline phosphatase (ALP), lactate dehydrogenase (LDH) activities, or in total protein (TP), albumin (ALB), UN, triglyceride (TG), total cholesterol (CHO), high-density lipoprotein (HDL), and low-density lipoprotein (LDL) concentrations ($P>0.05$).

In conclusion, dietary oligo-chitosan supplementation promotes both reproduction and lactation performance in sows. Based on the comprehensive results of this study, the optimal supplementation level of oligo-chitosan in sow diets is 50 g/t.

Key words: oligo-chitosan; sows; reproduction performance; lactation performance; plasma biochemical parameters

Introduction

Sows represent the core and foundation of pig production operations, with their reproductive performance serving as a critical indicator of productivity in intensive pig farms. The health status of sows directly determines the health of piglets and finishing pigs, while their production level and reproductive performance fundamentally dictate the economic efficiency of the entire operation. Research has demonstrated that metabolic pathways in pregnant animals differ from those in non-pregnant animals, with these differences becoming more pronounced as gestation advances. During late gestation, the rapid development of fetuses dramatically increases demands for oxygen and nutrients, imposing severe metabolic burdens on the dam and predisposing her to oxidative stress. The free radicals generated through metabolism can damage placental villi, leading to abnormalities in villous cell membranes, mitochondrial membranes, and placental function. These disruptions affect fetal development in utero, reduce sow lactation performance and immune function, and severely compromise sow health, longevity, and offspring development. Therefore, enhancing the immune function of sows during late gestation is of paramount importance.

Oligo-chitosan refers to chitosan degraded through enzymatic, chemical, or physical methods. Due to its low degree of polymerization and relatively small molecular weight, oligo-chitosan possesses physiological and pharmacological activities that high-molecular-weight chitosan lacks, with pharmacological activity 14 times greater than that of equivalent-weight chitosan. Low-degree-of-polymerization (2-10), water-soluble oligo-chitosan is called chitosan oligosaccharide, while material with a degree of polymerization above 20 that is not readily water-soluble is termed low-molecular-weight chitosan. Studies have shown that as a novel feed additive, oligo-chitosan exhibits various biological activities including hypolipidemic, antioxidant, antimicrobial, immune-enhancing, and gut health-improving effects. Current research on oligo-chitosan applications has primarily focused on its effects on growth performance and immune function, lipid metabolism, antioxidant capacity, and muscle development in young livestock. However, reports on the application of oligo-chitosan in sow

diets remain extremely limited. This experiment was designed to investigate the effects of oligo-chitosan supplementation in late-gestation sow diets on reproductive performance, lactation performance, and plasma biochemical parameters, thereby providing a theoretical basis for its rational application in sow nutrition.

1.1 Experimental Materials

Oligo-chitosan was produced by Jiaxing Kerui Biotechnology Co., Ltd., with an average relative molecular weight of 2.5×10^4 , deacetylation degree $>85\%$, purity $>95\%$, and particle size of approximately 20 μm .

1.2 Experimental Animals and Design

Twenty-four healthy Yorkshire sows with similar parity, body weight, and expected farrowing date were selected and randomly allocated to three groups, with eight replicates per group and one sow per replicate. The control group received a basal diet, while experimental groups I and II were fed the basal diet supplemented with 50 and 100 g/t oligo-chitosan, respectively. The experimental period commenced on day 85 of gestation and concluded on day 21 postpartum (piglet weaning).

1.3 Experimental Diets

From day 85 of gestation, experimental sows were fed lactation diets. The basal diet was formulated according to NRC (2012) nutrient requirements combined with the actual conditions of the experimental farm. The composition and nutrient levels are presented in Table 1. Diets were manufactured as pellets one week before the experiment began to maintain freshness.

Table 1 Composition and nutrient levels of the basal diet (air-dry basis), %

Ingredients	Content	Nutrient levels ²	Content
Corn		Digestible energy (DE)/(MJ/kg)	
Soybean meal		Crude protein (CP)	
Wheat bran		Calcium (Ca)	
Extruded soybean		Total phosphorus (TP)	
Fish meal		Available phosphorus (AP)	
Oil meal		Total lysine (T-Lys)	
Glucose		Methionine + Cysteine (Met+Cys)	
Premix ¹		Total threonine (T-Thr)	
Total		Total tryptophan (T-Trp)	
		Total valine (T-Val)	

¹Premix provided the following per kg of diet: Fe 80 mg, Mn 45 mg, Zn 100 mg, Cu 20 mg, I 0.70 mg, Se 0.25 mg, VA 10,000 IU, VD₃ 2,500 IU, VE 100 IU, VK₃ 10 IU, VB₁ 4 mg, VB₂ 10 mg, VB₆ 1 mg, VB₁₂ 50 g, nicotinic acid 42.9 mg, pantothenic acid 21.6 mg, biotin 80 g, folic acid 5 mg, choline chloride 1,500 mg.

²Nutrient levels were all calculated values.

1.4 Feeding Management

The experiment was conducted at the Lukou Original Breed Pig Farm of Hunan Xinguang' an Agriculture and Livestock Co., Ltd., following strict biosecurity and management protocols. Sows were fed twice daily at 07:30 and 15:00 with ad libitum access to water. Pens were cleaned twice daily, and the surrounding areas were disinfected with spray every three days. Management and environmental conditions were consistent across all groups.

1.5 Sample Collection and Measurements

Feces: Defecation patterns of sows during late gestation were continuously monitored. Fecal characteristics were evaluated based on color, shape, and moisture content using the following criteria: 1) Color was described as black, brown, or yellow; 2) Shape and moisture content were classified as hard (low moisture), normal (formed, moderate moisture), soft (unformed but piled, high moisture), or watery (unformed, not piled, excessive moisture).

Backfat thickness: Backfat thickness was measured on day 110 of gestation and day 21 postpartum using a backfat and loin-eye ultrasound device (MyLab Touch VET). Backfat thickness loss was calculated as the difference between pre- and post-farrowing measurements.

Reproductive performance: Daily feed intake was recorded for each sow. Total piglets born, healthy piglets per litter, and farrowing duration (interval from first piglet to placenta expulsion) were recorded. Birth litter weight and litter weights at 7, 14, and 21 days of age were measured.

Milk yield: Milk yield was estimated based on the approximation that 1 kg of piglet weight gain requires 3 kg of sow milk. The calculation formula was:
Milk yield = (Litter weight at 21 days - Birth litter weight) × 3.

Milk composition: Colostrum (10 mL) was collected on the day of parturition, and milk (10 mL) was collected on day 10 of lactation. Conventional nutrient components (milk fat, milk protein, lactose, urea nitrogen [UN], defatted dry matter, total dry matter) were analyzed using a milk composition analyzer (MilkoScan™ FT+, FOSS, Denmark). Amino acid content was determined using an L-8800 automatic amino acid analyzer (Hitachi, Japan) according to established methods.

Plasma biochemical parameters: On day 21 postpartum, 10 mL of blood was collected from the anterior vena cava into heparinized tubes, gently mixed,

allowed to stand at room temperature for 30 minutes, and then centrifuged to harvest plasma. Plasma concentrations of glutamic-pyruvic transaminase (GPT), glutamic-oxaloacetic transaminase (GOT), alkaline phosphatase (ALP), α -amylase (α -AMY), lactate dehydrogenase (LDH), total protein (TP), albumin (ALB), UN, glucose (GLU), triglyceride (TG), total cholesterol (CHO), high-density lipoprotein (HDL), and low-density lipoprotein (LDL) were measured using a 7060 automatic biochemical analyzer (Hitachi, Japan). All biochemical assay kits were purchased from Beijing Leadman Biochemistry Co., Ltd.

1.6 Data Processing and Analysis

Experimental data were initially processed using Excel 2010 and subsequently analyzed using SAS 9.1 (SAS Institute Inc.) via one-way ANOVA. Significant differences were further analyzed using Duncan's multiple comparison test. Results are expressed as means \pm standard error, with $P < 0.05$ considered statistically significant.

2.1 Effects of Oligo-chitosan on Feces and Backfat Thickness of Sows

As shown in Table 2, sows in the control group exhibited constipation during late gestation, whereas dietary oligo-chitosan supplementation improved fecal consistency by reducing hardness and increasing moisture content.

Table 2 Effects of oligo-chitosan on feces of sows

Items	Control group	Experimental group	Experimental group
Color			
Shape			
Moisture content			

Table 3 presents the backfat thickness measurements. No significant differences were observed among groups in backfat thickness on day 110 of gestation or day 21 postpartum ($P > 0.05$). However, backfat thickness loss tended to be lower in experimental groups I and II compared with the control group ($P > 0.05$).

Table 3 Effects of oligo-chitosan on backfat thickness of sows

Items	Control group	Experimental group	Experimental group	P-value
Day 110 of pregnancy (mm)	16.54 \pm 0.66	17.18 \pm 1.36	16.78 \pm 0.80	<i>Day21postpartum(mm)</i> 15.42 \pm 0.62 16.16 \pm 1.22 15.95 \pm 0.76 <i>B</i>

In the same row, values with the same or no letter superscripts indicate no significant difference ($P>0.05$), while different lowercase letter superscripts indicate significant difference ($P<0.05$). The same applies below.

2.2 Effects of Oligo-chitosan on Reproduction Performance of Sows

Table 4 summarizes the reproductive performance data. No significant differences were observed among groups in total piglets born per litter ($P>0.05$). However, the number of healthy piglets per litter was significantly higher in both experimental groups I and II compared with the control group ($P<0.05$), with experimental group I showing significantly more healthy piglets than group II ($P<0.05$). Farrowing duration was significantly shorter in both experimental groups compared with the control group ($P<0.05$). Additionally, ADFI during lactation was significantly higher in both experimental groups I and II relative to the control group ($P<0.05$). Birth litter weight was significantly increased in experimental group I ($P<0.05$), while total milk yield during lactation was significantly elevated in experimental group II ($P<0.05$).

Table 4 Effects of oligo-chitosan on reproduction performance of sows

Items	Control group	Experimental group	Experimental group	P-value
Total piglets born per litter	10.57 \pm 1.31	12.86 \pm 0.59	11.00 \pm 0.94	<i>Healthypigletsperlitter</i> 8.29 \pm 0.52 ^c 12.14 \pm 0.34 ^a 10.13 \pm 0.44 ^b < 0.01
<i>Farrowingduration(h)</i>	4.07 \pm 0.17 ^a	3.50 \pm 0.21 ^b	3.35 \pm 0.15 ^b	< 0.01
<i>ADFIoflatepregnancysows(kg)</i>	2.27 \pm 0.03	2.35 \pm 0.04	2.38 \pm 0.04	<i>ADFIoflactationsows(kg)</i> 3.01 \pm 0.05 ^a 3.12 \pm 0.06 ^a 3.23 \pm 0.07 ^a < 0.01
<i>Birthlitterweight(kg)</i>	13.42 \pm 1.06 ^b	18.45 \pm 0.88 ^a	15.29 \pm 1.01 ^b	< 0.01
<i>Litterweightat7days(kg)</i>	26.54 \pm 0.35	30.81 \pm 1.35	27.65 \pm 1.65	<i>ADGfrom1to7days(g)</i> 147.56 \pm 14.21 ^a 158.32 \pm 15.12 ^b 151.23 \pm 16.01 ^b < 0.01
<i>Averagedailymilkyield(kg)</i>	5.53 \pm 0.71	5.83 \pm 0.54	6.26 \pm 0.36	<i>Ratiooffeedtomilk</i> 0.72 \pm 0.05 0.76 \pm 0.04 0.81 \pm 0.03 < 0.01

2.3 Effects of Oligo-chitosan on Milk Composition of Sows

As presented in Table 5, milk fat content in colostrum was significantly higher in both experimental groups I and II compared with the control group ($P<0.05$). Additionally, UN content in colostrum was significantly elevated in experimental

weight gain and extending the weaning-to-estrus interval while reducing conception rates. Although 100 g/t oligo-chitosan significantly increased total milk yield during lactation, it may have increased backfat thickness loss due to greater mobilization of body reserves. However, because oligo-chitosan supplementation increased ADFI during lactation, backfat thickness loss did not differ significantly from the control group.

3.2 Effects of Dietary Oligo-chitosan on Lactation Performance of Sows

Lactation performance is closely related to reproductive performance in animals. The conventional nutrient composition and milk yield of sow milk are intimately associated with piglet survival and growth, making them important considerations in sow diet formulation. This study demonstrates that oligo-chitosan supplementation increases total milk yield and average daily milk yield during lactation, likely reflecting the combined effects of inhibiting and clearing intestinal pathogenic bacteria while enhancing sow immunity. Lactose, milk fat, and milk protein are essential nutrients in sow milk, and their concentrations directly affect piglet performance. While lactose and milk protein content in sow milk typically remain stable despite dietary changes, milk fat in colostrum and milk is primarily utilized by neonatal piglets for body fat deposition. This study found that oligo-chitosan supplementation significantly increased milk fat content in colostrum, which benefits piglet body fat deposition and overall growth and development.

Research has shown that abundant amino acids in milk are used by piglets to synthesize body proteins, hormones, polyamines, purines, pyrimidines, and other biologically active molecules that promote growth and development. In this study, supplementation with 50 g/t oligo-chitosan significantly increased total amino acid and limiting amino acid concentrations in milk compared with the control group, which may be related to oligo-chitosan-mediated mechanisms of amino acid transport in mammary epithelial cells. Further research is warranted to elucidate the effects of oligo-chitosan on amino acid uptake by mammary glands.

3.3 Effects of Dietary Oligo-chitosan on Plasma Biochemical Parameters of Sows

Plasma biochemical parameters serve as important indicators of organ function and metabolic status in animals. Plasma ALP, GPT, and GOT activities are commonly used to assess liver health, while TP, ALB, and UN concentrations reflect protein metabolism and amino acid balance. The significantly lower plasma GPT activity in oligo-chitosan-supplemented groups is consistent with findings from studies on growing-finishing pigs. The tendency for lower plasma ALP and LDH activities in the 100 g/t oligo-chitosan group suggests that oligo-chitosan may enhance humoral immunity and improve liver health. This effect likely

stems from oligo-chitosan's antimicrobial properties and its ability to activate macrophages and T lymphocytes, thereby enhancing phagocytic and cytotoxic activity, helping the animal resist pathogenic infections and strengthening immune function to protect liver health. The absence of significant differences in plasma TP, ALB, and UN concentrations among groups indicates that oligo-chitosan does not significantly affect protein metabolism in pregnant sows.

Conclusion

Dietary oligo-chitosan supplementation promotes both reproduction and lactation performance in sows. Oligo-chitosan increases lactation feed intake, thereby enhancing milk yield and litter weight; reduces farrowing duration, consequently increasing the number of healthy piglets; and elevates colostrum milk fat content to promote piglet health and development. Additionally, oligo-chitosan improves constipation and reduces backfat thickness loss, demonstrating its effectiveness as a feed additive. Based on the comprehensive results of this study, the optimal supplementation level of oligo-chitosan in sow diets is 50 g/t.

References

- [1] VANNUCCHI C I, JORDAO A A, VANNUCCHI H. Antioxidant compounds and oxidative stress in female during pregnancy[J]. *Research in Veterinary Science*, 2007, 83(2): 188-193.
- [2] BERCHIERI-RONCHI C B, KIM S W, ZHAO Y, et al. Oxidative stress status of highly prolific sows during gestation and lactation[J]. *Animal*, 2011, 5(11): 1774-1779.
- [3] ZHAO Y, FLOWERS W L, SARAIVA A, et al. Effect of social ranks and gestation housing systems on oxidative stress status, reproductive performance, and immune status of sows[J]. *Journal of Animal Science*, 2013, 91(12): 5848-5858.
- [4] ZENG Lintao. Study on preparation and physiological activity of chitosan oligosaccharide[D]. Master's thesis. Wuhan: Central China Normal University, 2007: 16-23.
- [5] WEI Xinlin, XIA Wenshui. Study on characteristics of chitosan oligosaccharide[J]. *Fisheries Science*, 2004, 23(2): 15-19.
- [6] ZHOU T X, CHEN Y J, YOO J S, et al. Effects of chitooligosaccharide supplementation on performance, blood characteristics, relative organ weight, and meat quality in broiler chickens[J]. *Poultry Science*, 2009, 88(3): 593-600.
- [7] PI Yu, CHEN Qing, SUN Lisha, et al. Biological activity of oligo-chitosan and its application in livestock and poultry production[J]. *Feed Review*, 2013(12): 31-35.
- [8] NGUYEN N D, VAN DANG P, LE A Q, et al. Effect of oligochitosan and oligo-beta-glucan supplementation on growth, innate immunity, and disease

- resistance of striped catfish (*Pangasianodon hypophthalmus*)[J]. *Biotechnology and Applied Biochemistry*, 2017, DOI: 10.1002/bab.1513.
- [9] ZHANG Liping, DONG Wanfu. Research progress on application of oligo-chitosan in weaned piglets[J]. *Feed Research*, 2016(6): 12-15.
- [10] ZHAO Ying, QIAO Lihong, WU Dawei, et al. Effects of oligo-chitosan on lipid metabolism and egg quality in laying hens[J]. *Cereal and Feed Industry*, 2013, 12(1): 45-48.
- [11] CHEN Gangyao, TANG Zhigang, WEN Chao, et al. Effects of oligo-chitosan on antioxidant capacity, lipid metabolism and muscle quality in growing-finishing pigs[J]. *Cereal and Feed Industry*, 2012(8): 54-57.
- [12] QIAO Lihong, ZHAO Ying, NI Hongyu, et al. Effects of oligo-chitosan on serum biochemical parameters, antioxidant capacity and fecal microorganisms in weaned piglets[J]. *Cereal and Feed Industry*, 2013(3): 47-50.
- [13] QIAO Enmei, ZHAO Yunrong, WANG Chenfang, et al. Effects of oligo-chitosan on muscle development in broiler chickens[J]. *Journal of the Chinese Cereals and Oils Association*, 2013, 28(9): 86-90.
- [14] YANG Weili, HU Qin, WANG Chunmei, et al. Effects of oligo-chitosan and symbiotics on production performance, serum indices and muscle composition in growing-finishing pigs[J]. *Journal of Domestic Animal Ecology*, 2016, 37(4): 27-31.
- [15] LIU Y Y, LI F N, KONG X F, et al. Signaling pathways related to protein synthesis and amino acid concentration in pig skeletal muscles depend on the dietary protein level, genotype and developmental stages[J]. *PLoS One*, 2015, 10(9): e0138277.
- [16] YANG Min, MI Yong, WU De. Research progress on factors affecting sow farrowing duration[J]. *Swine Industry Science*, 2015, 32(7): 108-109.
- [17] YOUNGQUIST R S, THRELFALL W R. *Current therapy in large animal theriogenology*[M]. 2nd ed. St. Louis, MO: Elsevier, 2006.
- [18] LIN Lijuan, CAI Jieqiong, ZHANG Tingting, et al. Study on in vitro and in vivo antibacterial effects of oligo-chitosan against *Salmonella pullorum*[J]. *Cereal and Feed Industry*, 2012(12): 48-50.
- [19] CI L, LIU Z Q, GUO J, et al. The influence of maternal dietary fat on the fatty acid composition and lipid metabolism in subcutaneous fat of progeny pigs[J]. *Meat Science*, 2015, 108: 82-87.
- [20] DU Mingqing. Effects of different amino acid levels on production performance, blood indices and milk amino acid concentration in lactating sows[D]. Master's thesis. Ya'an: Sichuan Agricultural University, 2010: 13-21.
- [21] WANG Cheng. Effects of fatty acid composition in sow diets on fatty acid composition in sow milk, blood and piglet blood[J]. *Feed Wide Angle*, 2012(20): 30-33.
- [22] FLYNN N E, KNABE D A, MALLICK B K, et al. Postnatal changes of plasma amino acids in suckling pigs[J]. *Journal of Animal Science*, 2000, 78(9): 2369-2375.
- [23] LIU Y Y, KONG X F, JIANG G L, et al. Effects of dietary protein/energy ratio on growth performance, carcass traits, meat quality, and plasma metabolites in pigs of different genotypes[J]. *Journal of Animal Science and*

Biotechnology, 2015, 6: 36.

[24] JI Yaohu, DOU Huating, WU Houjiu, et al. Research progress on antimicrobial activity of oligosaccharides[J]. Food Science, 2016, 37(13): 237-242.

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